Integrating new discoveries into the “vicious cycle” paradigm of prostate to bone metastases

Leah M. Cook¹, Gemma Shay¹, Arturo Aruajo², and Conor C. Lynch¹

¹Departments of Tumor Biology, Moffitt Cancer Center and Research Institute, Tampa, Florida, USA.

²Integrated Mathematical Oncology, Moffitt Cancer Center and Research Institute, Tampa, Florida, USA.

Summary

In prostate to bone metastases, the “vicious cycle” paradigm has been traditionally used to illustrate how metastases manipulate the bone forming osteoblasts and resorbing osteoclasts in order to yield factors that facilitate the growth and establishment. However, recent advances have illustrated that the cycle is far more complex than this simple interpretation. In this review, we will discuss the role of exosomes and hematopoietic/mesenchymal stem cells facilitate the establishment and activation of prostate metastases and, how cells including such as myeloid derived suppressor cells, macrophages, T-cells and nerve cells contribute to the momentum of the vicious cycle. The increased complexity of the tumor-bone microenvironment requires a systems level approach. The evolution of computational models to interrogate the tumor-bone microenvironment is also discussed and the application of this integrated approach should allow for the development of effective therapies to treat and cure prostate to bone metastases.

Keywords

Prostate cancer; Bone metastasis; Stem Cells; Immune Cells; Mathematical modeling; Vicious Cycle

1 Introduction

Prostate to bone metastases stimulate enhanced osteoblast and osteoclast activity that in turn promotes the release of growth factors from the bone matrix creating a “vicious cycle” of cancer growth. This feedback loop results in mixed lesions that are comprised of areas of extensive osteolysis (bone degradation) and osteogenesis (bone formation) with the latter being a hallmark of the disease [1]. While prostate to bone metastases remain incurable, therapeutic developments have begun to improve patient quality of life and begun push overall survival rates higher. Denosumab (Xgeva®), a monoclonal antibody to RANKL, has recently been approved for the treatment of prostate cancer patients. Denosumab has been

Corresponding Author Conor C. Lynch. Tumor Biology Department 12902 Magnolia Dr., SRB-3 Tampa, FL, USA. 33612 TEL: 813-745-8094 FAX: 813-745-3829 conor.lynch@moffitt.org.

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shown in clinical trials to significantly increase the time to SREs in comparison to bisphosphonates but does not appear to improve overall survival [2]. Another breakthrough has come in the form of radium-233 chloride (Alpharadin/Xofigo®) [3]. This alpha emitting radiopharmaceutical is similar to calcium in structure and targets the bone tissue. The short wavelength of the radiation ensures less damage to surrounding tissues when compared to beta or gamma emitting sources of radiation. In clinical trials of men with castrate resistant prostate cancer (CRPC) bone metastases, alpharadin was found to significantly improve overall survival by 3-4 months and is the first therapy to achieve this milestone [4]. Despite these advances, there is room for improvement. New studies have begun to define the precise cellular and molecular players that drive the establishment and evolution of prostate to bone metastases thus presenting novel targets for therapeutic intervention (Table 1). These advances and how to integrate them on a global level are the focus of the current review.

2 Preparing the “soil”

In 1889, Stephen Paget classically described the “seed and soil” hypothesis as the propensity of circulating tumor cells (CTCs) to home to specific organs, irrespective of the nearest anatomical location to the primary tumor [5]. CTCs preferentially metastasize and “seed” organs with the proper “soil.” While empirical evidence has identified genetic programs that facilitate the homing and initial establishment of prostate and breast cancer cells to bone such as Interleukin-11 (IL-11), connective tissue growth factor (CTGF), chemokine receptor 4 (CXCR4), matrix metalloproteinase-1 (MMP-1) and vascular cellular adhesion molecule-1 (VCAM-1), studies have also shown that primary tumor-derived signals can prepare the distant soil in pre-ordained areas for metastases [6-9]. This has been particularly true for lung metastasis in which studies using a B16 melanoma model identified that primary tumor-secreted factors induced the mobilization of immune-derived cells to niches in the lung where they promote angiogenesis and extracellular matrix remodeling [10, 11]. These niches then become the sites of future metastases. Understanding the factors that control the formation of the pre-metastatic niche in bone, could yield insights into identifying therapeutic targets that will prevent metastasis and therefore, have a significant impact on cancer related deaths. Recently, studies have implicated tumor derived microvesicular liposomes known as exosomes in facilitating the initial communication between the primary tumor and the site of metastasis [12, 13].

2.1 Exosomes and Oncosomes

Exosomes were first identified in the mid-1980s as recycled portions of intravesicular membranes released by reticulocytes following endocytosis of the membrane receptor, transferrin [14, 15]. Exosomes are utilized for the packaging and recycling of membrane and cytosolic proteins [16]. Produced by a variety of cells, exosomes vary in size from 30-100nm in diameter and can contain numerous proteins that control for example, membrane fusion and transport (annexins, small GTPase Rab proteins and Lamp proteins) [17]. Despite the variability in exosome contents and membrane proteins, components can be transferred both homo- and heterotypically between multiple cell types [18].
Significant roles for exosomes in the establishment of the pre-metastatic niche and metastasis have been recently described [13]. Systemic delivery of fluorescent exosomes from metastatic B16F10 melanoma cells have been shown to localize to lungs and other tissues including bone. Subsequent experiments identified that the growth and metastasis of subcutaneous B16F10 metastatic melanoma tumors was significantly increased in C57BL/6J mice that received exosomes from B16F10 cells, compared to mice that received exosomes from congenic non-metastatic B16F1 cells [13]. Furthermore, the developing metastases co-localized at the sites of exosome deposition with the underlying mechanism being implicated as exosome derived MET, receptor for hepatocyte growth factor (HGF), transfer to bone marrow-derived cells (BMDCs). These data illustrate a role for exosomes in the development of metastatic lesions. Supporting this observation, knockdown of Rab27a, a gene necessary for exosome release, significantly reduced the number of circulating exosomes and the number of metastases, demonstrating that metastatic burden was dependent on the number of circulating exosomes [13, 19]. Clinically, the detection of TYRP2 and MET in exosomes derived from melanoma patient plasma is already proving informative as a prognostic indicator of disease progression [13]. MET is involved in normal prostate development and promotes progression of malignant prostate cells. Androgen ablation therapy increases MET expression by bone marrow stromal, osteoblastic and prostate tumor cells promoting the transition of localized prostate cancer to a more metastatic phenotype [20]. Thus exosome-mediated transfer of MET to bone marrow stromal cells may play a large role in prostate tumor progression in bone.

In prostate cancer, the number of oncosomes (vesicles larger than exosomes that range in size from 1-10 μm) in patient plasma was found to correlate with Gleason scores of 7 and higher while exosomes isolated from LNCaP, DU145, and PC3 prostate cancer cell lines have been found to enhance osteoblast differentiation and to induce the differentiation of bone marrow-derived mesenchymal stem cells (MSCs) into myofibroblasts that promote tumor growth [21-23]. Collectively, these data suggest a role for oncosomes in prostate cancer malignancy and potentially in the establishment of pre-metastatic niches in the bone microenvironment.

Advances in our understanding of how exosomes can contribute to tumor progression have provided new angles for prognostic evaluation and potentially for therapeutic intervention [13]. Cabozantinib (XL184), a vascular endothelial growth growth factor (VEGF) and MET inhibitor, has had very promising results in phase II clinical trials of men with advanced prostate cancer. Indications suggest significant decreases in tumor burden and increases in overall survival [24]. MET transfer from exosomes to bone marrow derived cells is an important step in generating the pre-metastatic niche so it is possible cabozantinib may also interfere with the process. Approval of cabozantinib will add to the clinician’s ability to extend overall survival of patients with bone metastases and also supports the approach of developing targeted therapies to cells and factors that control the vicious cycle (Figure 1). Preventing exosome release by targeting proteins involved in the process such as lysosome associated membrane proteins (Lamps) could have a significant impact on cancer progression. Exosomes are non-immunogenic and therefore could be the perfect means through which to deliver therapies. In this regard, cationic liposomes that release their contents via exosome release were loaded with siRNA to Plekho1, a factor responsible for...
tagging osteogenic genes such as Runx2 for ubiquitination, and were found to increase osteoblast mediated bone formation and demonstrate the potential feasibility of the approach in the clinical setting [25].

3 New roles for stem cells in the establishment of prostate to bone metastases

Extravasation and colonization in bone is tightly regulated by adhesion molecules such as VCAM-1, which has been shown to be critical in the initial interaction between CTCs and stromal cells [26, 27]. Additionally, VCAM-1-mediated osteoclast activity releases disseminated tumor cells (DTCs) from dormancy, converting indolent metastasis to actively growing lesions demonstrating an important role for VCAM-1 in both the initial entrance and activation of DTCs in bone [6]. Once in the bone marrow, DTCs must survive in the new microenvironment and communicate with the host cells in order to generate active bone metastases. A number of new exciting studies illustrate how metastatic prostate cancer cells can interact with hematopoietic stem cell niches and mesenchymal stromal cells to facilitate establishment in the bone microenvironment (Figure 2).

3.1 Hematopoietic Stem Cells (HSCs)

HSCs reside in specialized niches within the bone marrow and are the source of myeloid and lymphoid cell lineages [28]. HSCs can mobilize from the marrow to distant regions in response to injury or primary cancer and subsequently return to bone marrow stem cell niches where they remain quiescent until needed again [29]. Niches in the bone marrow can be found proximal to the sinusoidal vasculature or the endosteal surface at the bone-marrow interface. Lining the endosteal surface are osteoblasts to which HSCs adhere via interactions with VCAM-1, selectins (P- and E-selectin), angiopoietin (via interactions with Tie2), or Kit-ligand [30, 31]. Osteoblasts are critical for the maintenance of HSC quiescence in the endosteal niches but can activate HSC expansion via the expression of chemokine ligand 12 (CXCL12) and its interaction with CXCR4 on the HSC cell surface. Osteoclasts also play a role in mobilizing HSCs from the endosteal niche via the expression of matrix metalloproteinase-9 (MMP-9) that in turn processes Kit-ligand, a mechanism that facilitates the release of the HSCs from the niche and increases CXCL12 and VEGF expression in the bone marrow compartment. Newly arriving prostate to bone metastases can utilize the factors involved in the maintenance of the HSC niche. For example, SUMO-specific protease 1 (SENP1), a member of the deSUMOylation protease family, has been recently identified as a regulator of MMP9 expression by prostate tumors and promotes prostate cancer metastasis to bone [32]. Additionally, CXCR4 expression by the prostate cancer cells is involved in the homing of prostate metastases to the bone, in particular to areas that are rich in CXCL12 such as the endosteal HSC niche [33].

Recent evidence has identified how metastatic prostate cancer cells directly compete with HSCs to engraft within the bone marrow compartment [34]. Disseminated cells shed from implanted subcutaneous prostate cancers (PC3 or C42B) were found to inhibit HSCs engraftment after bone marrow transplant [35]. Results show that the HSCs and disseminated prostate metastases co-localized to osteoblasts expressing CXCL12 and
annexin 2, an adhesion molecule utilized by HSCs to adhere to osteoblasts in the endothelial stem cell niche. Increasing osteoblast number via parathyroid hormone (PTH) treatments and thereby, the number of niches also increased the number of metastatic cells that engrafted within the bone marrow. Likewise, ganciclovir-mediated ablation of the osteoblasts decreased the number of endothelial stem cell niches and thus the number of engrafted metastatic cells in osteoblast specific (Col2.3Δ-TK) transgenic mice, demonstrating the critical role of osteoblasts and the HSC stem cell niche in prostate cancer engraftment within the bone marrow. Interestingly, prostate metastases occupying the endosteal niche in tumor bearing mice can also be mobilized back into the circulation by administrating CXCL12 blocking antibodies or treatment with granulocyte-colony stimulating factor (G-CSF) [35].

In addition to utilizing the same pathways as HSCs to home and graft into bone niches, prostate cancer cells can also manipulate HSC differentiation to facilitate the induction of the vicious cycle. Using osteolytic and osteogenic cell lines, PC3 and C42B respectively, studies have shown that in the presence of RANKL, HSCs derived from mice bearing subcutaneous PC3 tumors can promote the differentiation of naïve HSCs into osteoclasts compared to HSCs derived from C42B bearing mice with PC3 derived interleukin-6 (IL-6) being implicated as the underlying mechanism involved [36]. IL-6 is important for prostate tumor growth in bone as well as for progression of primary prostate tumors via cross-activation of insulin-like growth factor (IGF) signaling, one of the most abundant factors released from the bone matrix upon osteolysis [37, 38]. These data demonstrate the effects that prostate cancer cells can elicit on the HSC compartment to initiate the lytic compartment of the vicious cycle. Intriguingly, the authors also show in converse experiments that HSCs isolated from C42B tumor bearing animals exhibited increased BMP2 and BMP6 expression and promoted the differentiation of naïve bone marrow derived mesenchymal stromal cells (MSCs) into osteoblasts more efficiently than HSCs derived from PC3 tumor bearing animals thus implicating potential roles for MSCs in prostate cancer induced osteogenesis.

These advances offer a means to therapeutically prevent bone metastasis formation by targeting underlying molecular mechanisms such as VCAM-1 and CXCL12. In recent years, there have been an increasing number of clinical trials for CXCL12/CXCR4 antagonists as anticancer agents. Nox-A12, a CXCL12 antagonist, is currently in clinical development as an anticancer agent and, has recently been introduced into Phase III clinical trials of myeloma patients, a cancer known to promote increased osteolysis in bone. Additionally, AMD3100 was recently FDA-approved for the use of stem cell mobilization in Non-Hodgkins lymphoma and multiple myeloma along with other CXCL12 inhibitors including BKT140, POL6326, and TG-0054 [39, 40]. However, MSX-122 is the first reported CXCL12 inhibitor in Phase 1 clinical trials for solid tumor cancers. Activation of peroxisome proliferator-activated receptor-α (PPARα) with Wy14643 has been shown to inhibit VCAM-1, abrogating HSC interactions with the endosteal niche, and promote HSC mobilization [41]. Inhibition of VCAM-1 in prostate cancer patients with early stage disease may prevent both DTC extravasation into bone and localization to HSC niches and should be further investigated. Therapeutic targeting of the CXCL12/CXCR4 or VCAM-1 pathway in combination with cytokine targeted therapies (e.g. Tocilizumab, monoclonal antibody to
IL-6) may be beneficial for preventing DTC localization to the HSC niche and prostate tumor progression in bone. It is currently unknown whether these CXCL12-targeted therapies will mobilize tumor cells that may metastasize to other areas or “re-seed” the primary tumor as has been noted [35]. This, however, is a profound observation that has implications for not only preventing bone metastases but also in considering how applied therapies may affect disseminated prostate cancer cells.

### 3.2 Mesenchymal Stromal Cells (MSCs)

Approximately 0.001% to 0.01% of the marrow compartment is comprised of mesenchymal stromal/stem cells (MSCs) [42]. Given the correct cues, MSCs can differentiate into fibroblasts, chondrocytes, myocytes, adipocytes or osteoblasts and are involved in the maintenance and regeneration of skeletal tissues, including connective tissues, cartilage, muscle, fat tissue and bone [43]. MSCs are multipotent, have limitless proliferative abilities, maintain the HSC niche and can be mobilized from the marrow to sites of tissue injury for repair. Virchow has described cancer as a wound that does not heal and therefore, it is unsurprising that MSCs have been identified as contributing to the progression of primary cancers. In breast cancer, chemokine ligand 5 (CCL5) plays a key role in the recruitment of MSCs to the primary tumor microenvironment [44]. Heightened MSC infiltration results in enhanced expression of lysyl oxidase (LOX) and angiogenesis that in turn promotes the invasion and metastasis of the cancer cells [44, 45]. In primary prostate cancer, CXCL16 facilitates MSC recruitment via the receptor CXCR6 to the tumor microenvironment and their subsequent differentiation into cancer-associated fibroblasts (CAFs). The MSC-derived CAFs were found to be critical for promoting tumor growth, epithelial to mesenchymal transition (EMT), and metastasis to bone [46]. CAFs exhibit CD44-mediated increased TWIST expression, a downstream target of transforming growth factor β (TGFβ) that promotes EMT, differentiation of MSCs into CAFs and osteoclastogenesis [47-49]. CD44 and TWIST may play a large role in recruiting MSCs and in stimulating the differentiation of tumor-promoting CAFs in bone.

It is important to note that while these studies have highlighted pro-tumorigenic roles for MSCs in cancer progression, other reports have documented protective roles. For example, MSCs have been found to home to and, potently inhibit, cancer proliferation and angiogenesis by decreasing Akt signaling and causing endothelial cell death via reactive oxygen species (ROS) respectively [50]. In contrast, VEGF is produced by MSCs and promotes angiogenesis, MSC migration and osteoblast differentiation, demonstrating the multiplicity of MSC function [51-53]. Expression of Wnt antagonist Dickkopf-1 (Dkk-1), by MSCs can suppress breast cancer growth but conversely enhances prostate cancer metastasis [54, 55]. Clearly, more investigation is required to define how MSCs contribute to disease progression in different contexts. Complicating this picture is the fact that the “stemness” of MSCs has been difficult to define despite the availability of many markers. Recent studies have also identified that the source of tissue from which stem cells are derived (adipose vs. bone marrow for example) should be a critical consideration for testing the roles of MSCs in cancer progression [56].
Since bone is a major reservoir for MSCs and the MSC niche is thought to be proximal to sinusoidal vasculature, prostate cancer metastases arriving in the bone marrow have a unique opportunity to interact with MSCs. MSC-derived factors such as IL-6 and VEGF directly promote secretion of tumor-derived factors that induce precursor osteoblast proliferation and differentiation (such as endothelin 1 (ET1) and TGFβ). However, despite the ability of MSCs to differentiate into osteoblasts and prostate cancers to express potent mediators of osteoblast differentiation, no studies to date have examined the interplay between prostate to bone metastases and MSCs in the generation of osteogenic lesions. For example, Wnt7B, a mediator of MSC differentiation, was recently found to be highly expressed by castration-resistant prostate cancer (CRPC) cells and in osteoblastic xenografts and is important for their proliferation, implicating a dual role for Wnt signaling in promoting MSC-mediated bone formation as well as tumor growth [57]. The differentiation of MSCs into other lineages such as adipocytes for example could also impact prostate cancer progression in bone [58]. Bone is a rich source of adipocytes and adipokines such as leptin and adiponectin can promote osteoblast differentiation, proliferation and mineralization while inhibiting osteoclastogenesis [59]. Understanding the roles of MSCs in the prostate cancer-bone vicious cycle will be challenging but has the exciting potential to yield new insights into how prostate metastases establish and grow in bone.

Given the multiple roles MSCs and MSC derived factors in primary and bone metastatic prostate cancer, a clear opportunity for therapeutic intervention exists (Figure 2). For example, VEGF, is produced by MSCs in large quantities and, in addition to promoting angiogenesis, directly promotes MSC migration and osteoblast differentiation. Thus targeting of VEGF may inhibit tumor growth as well as the tumor-promoting effects of MSCs in the tumor-bone microenvironment [51-53]. Preclinical studies investigating VEGF-targeted therapies, such as bevasuzimab, have successfully inhibited prostate cancer growth in bone. However, MET expression can be increased by VEGF inhibition thus dual targeting of both pathways with carbozantenib, for example, could overcome these effects [60-62]. The use of MSCs for targeted gene therapy is also a potentially powerful way to specifically deliver therapies to prostate to bone metastases but key to this approach is ensuring that the MSCs do not exacerbate prostate cancer progression in bone. Therapies such as autologous stem cell transplant could be used to isolate individual patient MSCs and induce or manipulate them to express anti-cancer factors. For example, injection of bone marrow-derived MSCs exogenously expressing osteoprotegrin into PC3 prostate cancer lesions significantly inhibited osteoclastogenesis [63]. Further characterization will be required to understand the optimal approach for the therapeutic targeting or use of MSCs in the treatment of prostate to bone metastases.

4 Integrating immune cell functions into the vicious cycle

Compelling evidence has emerged in recent years indicating that immune cell polarization in response to factors in the tumor microenvironment is a powerful regulator of cancer progression and metastasis. The bone marrow is a reservoir for immune cells such as: macrophages, myeloid derived suppressor cells (MDSCs), dendritic cells, and various T-cell subsets that can directly impact the steps of the vicious cycle (Figure 3).
4.1 MDSC and macrophages

In the bone marrow of healthy individuals, naïve immature myeloid cells (iMCs) do not have immune suppressive capabilities [64]. iMCs are poised to rapidly differentiate into mature macrophages, granulocytes and neutrophils. However, in response to tumor-derived factors (e.g. VEGF, SDF-1 and IL-3), differentiation is blocked, giving rise to MDSCs that have immune suppressive capabilities (widely defined as CD11b+GR1+ in mice), and can be identified in the bone marrow, spleen, blood and solid tumors [65-67]. MDSCs are defined as a heterogeneous population of activated, immature myeloid cells with the ability to suppress T-Cells and promote tumor growth [68]. This heterogeneous population of cells can be divided into two major subgroups in mice, granulocytic (CD11b+Ly6G+Ly6Clow) and monocytic MDSCs (CD11b+Ly6G-Ly6Chigh) that differ not only by their phenotype and morphology but also in their mechanism of suppression [68]. In addition to immune suppression, MDSCs can directly promote tumor growth by releasing pro-angiogenic factors and incorporating into developing vasculature [69]. MDSCs can also directly influence the vicious cycle due to their ability to differentiate into bone-resorbing osteoclasts in vivo [70-72]. Although iMC and MDSC can differentiate into osteoclasts in vitro, MDSCs isolated from the bone marrow of tumor bearing mice were found to be significantly primed for osteoclastogenesis compared to MDSCs derived from non-skeletal tissue [70]. Additionally, independent of their ability to differentiate into osteoclasts, MDSCs produce TGF-β that in turn can further promote cancer cell derived para-thyroid hormone related protein (PTHRP) expression thereby, accelerating the vicious cycle. Myeloid-derived TGF-β has been shown to be essential for tumor metastasis, with mice deficient in myeloid specific TGF-β, showing a significant reduction in metastasis due to IFN-y activation of CD8+ T-Cells [73]. MDSCs can play major roles in facilitating cancer progression in several sites including the metastatic bone microenvironment. Given the importance of MDSCs in cancer progression, they provide an interesting therapeutic target in the treatment of bone metastasis. Such therapeutic strategies include: inhibiting MDSC expansion (e.g. gemcitabine), stimulating their differentiation into mature antigen presenting cells (e.g. Trans retinoic acid) and inhibiting their function (e.g. Cox-2 inhibitors) [74-77]. These therapies, and others, are currently under investigation as adjuvants for immunotherapies in a variety of cancer types, with a significant enhancement of immune interventions by reversing MDSC induced immune suppression [78].

Tumor associated macrophages (TAMs) play pivotal roles in cancer progression and metastasis with the density of TAM infiltration correlating with a poor prognosis for several cancer types, including prostate cancer [79, 80]. Naïve macrophages polarize in response to microenvironment signals into an “anti” or “pro” tumor phenotypes that have a broad spectrum of functions but are classically defined as M1 and M2 macrophages respectively. However in cancer, the majority of TAMs are M2 orientated and largely contribute to tumor immunoevasion via the secretion of anti-inflammatory cytokines such as IL-10 [81]. Independent of their immune regulatory effects, TAMs can promote angiogenesis by expressing proteinases including MMP-9 that in turn regulates the bio-availability of growth factors such as VEGF [82]. Osteal macrophages (osteomacs) also contribute to bone healing [83]. In the context of prostate to bone metastases, the expression of chemokine ligand-2

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(CCL2), also known as monocyte chemoattractant protein-1 (MCP-1), by prostate cancer cells promotes the recruitment of TAMs and osteoclast precursors to the prostate bone microenvironment and the growth of the metastases implicating important roles for macrophages in the prostate cancer vicious cycle of bone metastasis [84]. In addition, bone marrow macrophages (BMMs) have been shown to highly express osteogenic-genes CCL-2, COX-2 and Cathepsin K. The overexpression of Cathepsin K by BMMs in bone tumors has been shown to promote tumor progression in a prostate cancer model by up regulating CCL-2 and COX-2 pathways in the bone microenvironment [85]. Such potential influences include the stimulation of αvβ3 integrin expression on prostate cancer cells increasing tumor cell migration and invasion by CCL-2 [86]. Targeting tumor and macrophage derived CCL-2 significantly increases survival of prostate cancer-bearing mice [87, 88]. Preclinical studies are investigating the reprogramming of macrophage polarization from an “M2” to a “M1” phenotype by targeting NFκB and COX-2 as a potential therapeutic strategy [89-91]. Inhibition of NFκB in TAMs resulted in the expression of pro-inflammatory cytokines such as IL-12 that were cytoxotic to tumor cells [89]. Several animal studies have indicated that nitrogen-containing bisphosphonates also reduce pro-angiogenic MMP-9, as well as skewing tumor associated macrophages to a M1 phenotype, by a yet to be understood mechanism [92, 93].

4.2 Dendritic cells

Dendritic cells (DCs) are derived from the myeloid lineage and act as messengers between the adaptive and innate arms of the immune system, specifically by presenting antigens primarily to adaptive T-cells. DCs are divided into two main groups; myeloid (mDC) and plasmacytoid (pDC) with the latter resembling plasma cells and expressing large amounts of interferon alpha (IL-α). Recently, pDCs have been implicated in regulating osteolysis. Primary cancers including prostate, are often heavily infiltrated by pDCs [94]. Activation of pDCs led to elevated levels of systemic circulating cytokines such as IL-15, CCL5 and CCL2 that in turn stimulate osteoclast activity in bone [84, 95]. These data imply roles for pDCs in the primary tumor microenvironment regulating systemic bone turnover and suggest that pDCs may play a role in the generation of pre-metastatic niches in the bone microenvironment as depletion of the pDCs in vivo also decreases the number of bone metastases [96]. Tumor-derived CXCL12 is important for the recruitment of pDCs to the tumor site, providing a potential therapeutic strategy by inhibiting the CXCR4 receptor (eg. Small molecule inhibitor AMD3100 [97]). Autologous dendritic cell transplant therapies with “educated” DCs (Provenge®) has proven an effective way to stimulate T-cell mediated recognition and attack of the prostate cancer cells, an approach that has been successful in extending the overall survival of prostate cancer patients [94]. It will be interesting to determine if the re-education of the dendritic cells also impacts bone remodeling and prostate to bone metastases.

4.3 T-cells

T-cell subclasses (eg. CD4+, CD8+ T-cells) originate from HSCs in the bone marrow and are activated in the thymus prior to migrating to their target tissue [98]. T-cell control of normal bone homeostasis and osteolytic inflammatory bone diseases such as rheumatoid arthritis and cancer-induced bone disease has been described [99]. In normal bone remodeling,
CD4+/CD8+ T-cell derived cytokines such as, IL-17 and TNF-α can act in an autocrine manner to stimulate RANKL expression. Thus, T-cells negate the requirement for the interaction of osteoclast precursors with osteoblasts and directly stimulate osteoclastogenesis [100]. However, IFN-γ expression by T-cells can prevent osteoclastogenesis and therefore, is a potential mechanism through which T-cells can dampen osteoclastogenesis and bone resorption [101, 102]. In the tumor-bone microenvironment T-cells can either promote or protect against cancer progression. Phospholipase C gamma-2 (PLC 2−/−) null mice have defects in osteoclast function and an impairment in T-cell activation. Surprisingly, the inoculation of tumors into these animals stimulated tumor growth in bone, despite less osteolysis. Rescue experiments in which the delivery of activated CD8+ T-Cells into PLC 2−/− resulted in reduced tumor burden and suggested a protective role for this T-cell subset [103]. Patients whose primary tumors have high levels of IFN type I were also found to be protected from bone metastasis. The secretion of IFN type I by tumor cells results in the expansion of CD8+ T-cells and NK cells, which is associated with a decrease in metastasis to the bone. The importance of NK cells and CD8+ T-cells was confirmed as mice lacking the IFN receptor, or depletion in these cells, had increased bone metastasis [104].

Regulatory T-cell (Treg CD4+/CD25+) infiltration of tumors can suppress immune mediated cytotoxicity and result in an immune-privileged tumor-microenvironment. Tregs are recruited to the bone marrow of prostate cancer patients with bone metastasis via CXCR4/CXCL12 signaling, where dendritic cells induce their expansion within the marrow. Tregs inhibit osteoclast differentiation in vitro and adoptive transfer of Tregs in a mouse model of prostate cancer also increased bone mineral density [105]. The authors suggest that this inhibitory effect of Tregs on osteoclasts may be responsible for the progression of osteoblastic prostate cancer. This interaction between T-cells, and osteoclasts is not solely one sided, with osteoclasts capable of influencing T-cell function. For example, osteoclasts secrete chemokines (in particular CCL4, CXCL5, CXCL1) that can recruit CD8+ T-cells. Upon recruitment, osteoclasts activate the CD8+ T-cells via antigen-presentation, to express cytokines including IL-2 and IFN-γ. These activated T-cells express FOXP3 that in turn inhibits the priming of naive CD8+ T-cell activation by dendritic cells. This interaction provides evidence of a feedback loop between osteoclasts and T-cells. [101]. It has been suggested that patients with bone metastasis may benefit from IFN-γ based therapies, with mice deficient in IFN-γ receptor, or NK and CD8+ T-cells more susceptible to bone metastasis [106]. In addition, treatment with standard of care nitrogen-containing bisphosphonates induces the expansion of a particular subset of γδ T-cells in some patients receiving the drugs intravenously. γδ T-cells are involved in tumor surveillance and upon target recognition can express pro-inflammatory cytokines that promotes cancer cell elimination. Interestingly, γδ T-cells themselves have the potential to inhibit osteoclast formation, via the secretion of IFN-γ [107]. Prostate cancer patients receiving zoledronate treatment that exhibited expansion in γδ T-cells, were found to have a reduction in disease progression, indicating a potential therapeutic approach [108]. In a similar manner, dendritic cell autologous therapies such as Provenge® can promote the activation of cytotoxic T-cells and may prove important in preventing prostate metastases and/or cancer progression in bone.
5 Blood-derived contributors to metastasis to bone

Immune cells such as B-cells and mast cells have been described as playing critical roles in metastasis, which is not surprising given the importance of other immune cells in the process [109, 110]. Interestingly, cells in the bone marrow microenvironment previously thought to be bystanders such as megakaryocytes, have been identified as playing active roles in the vicious cycle of prostate to bone metastases.

5.1 Megakaryocytes

Mature megakaryocytes are primarily located adjacent to blood vessels within the bone, where they migrate or protrude through the endothelium and deliver platelets directly into the circulating blood. Emerging evidence suggests that megakaryocytes can also regulate bone homeostasis. Mice with abnormal megakaryocytopoiesis have high bone content due to an increased number of osteoblasts, but a reduced number of osteoclasts. Megakaryocytes have also been shown to inhibit osteoclast differentiation via the release of soluble factors such as osteoprotegrin (OPG), an endogenous inhibitor of RANKL [111, 112]. Additionally, megakaryocytes influence bone formation by stimulating the proliferation and differentiation of osteoblasts via a cell-contact dependent mechanism suggesting that the number and activity of megakaryocytes within the bone marrow could potentially be important in the prostate to bone metastasis vicious cycle [113]. This potential has recently been explored. In an in vivo prostate cancer model the expansion of megakaryocytes (following treatment with thrombopoetin) within the bone marrow significantly reduced tumor burden [114]. Further, direct co-culture of megakaryocytes with prostate cancer cells reduced the prostate cancer growth by decreasing proliferation and inducing apoptosis. These data suggest that high numbers of megakaryocytes could play protective roles in prostate cancer to bone metastasis but it is possible that megakaryocytes could also promote cancer metastasis given the roles of platelets in the process. Several therapies are currently under clinical investigation for megakaryocyte expansion, predominately for treatment of thrombopenia associated with diseases such as acute myeloid leukemia and chronic hepatitis C. These therapies include Thrombopoetin mimetics such as Romiplostim, that also acts to increase megakaryocyte colony forming cells and Elthombopag, that increases megakaryocyte proliferation and maturation [115].

5.2 Platelets

Platelets are small anuclear cells derived from megakaryocytes that function primarily to induce coagulation at sites of tissue injury. Injury-induced platelet activation results in aggregation at the injury site and adhesion to the endothelium (via integrin and selectin interactions) as well as a subsequent release of growth-promoting and wound-healing factors including, VEGF, CXCL12, PDGF, IGF, and TGFβ from platelet granules. However, platelet aggregation and interaction with tumor cells enhances invasion and tumor metastasis via the formation of tumor emboli thereby; allowing the evasion of immune surveillance in circulation, adhesion of the emboli to the endothelium, and facilitating extravasation at distant tissue sites [116, 117]. Tumor-induced platelet cell activation can promote the release of platelet-derived growth factors that further aid in tumor progression at secondary tissue sites. Understanding the mechanisms that control platelet-cancer cell interaction could offer...
unique opportunities to prevent prostate to bone metastasis. The platelet membrane glycoprotein complex GPIIb/IIIa (now known as integrin αIIbβ3) is the most abundant integrin solely expressed by platelets and megakaryocytes. αIIbβ3 facilitates platelet binding to fibronectin and von Willebrand factor (vWF) on B16 melanoma cells with αIIbβ3 blocking antibodies decreasing platelet binding to melanoma cells by ~60% and significantly reducing the number of pulmonary metastases [118]. αIIbβ3 expression has been implicated as an important facilitator of bone metastasis. Mice that are null for β3 integrin and thus unable to form heteromeric complexes with αIIb and/or αv integrins, exhibited fewer metastases after intracardiac injection of B16 melanoma cells in immunocompetent mice [119]. Additionally, β3 null platelets significantly abrogated platelet aggregation with circulating tumor cells and resulted in reduced cell “seeding” within bone. There was also a significant reduction in tumor-induced osteolysis, a result of a dual role of β3 integrins in both platelet aggregation and osteoclastogenesis. Expression of E-selectin and integrins by platelet-tumor emboli are critical for prostate cancer extravasation. CCL2, a chemokine that promotes tumor progression in bone via osteoclast recruitment has recently been shown to induce αvβ3 expression, thereby promoting prostate cancer migration [84]. These and other findings highlight the importance of cell adhesion molecules in platelet interaction with both tumor cells and stromal cells in the bone [86, 120, 121].

Understanding the factors that control platelet activation can lead to therapeutic intervention opportunities. Targeting β3 integrins may prove an effective means at not only preventing metastasis by reducing the ability of cancer cells to form emboli by inhibiting platelet activation and osteoclast mediated bone resorption. Originally approved for prevention of thrombosis associated with cardiovascular disease, Abciximab (ReoPro®) and XV454 are potent antagonists of platelet GPIIb/IIIa receptors (including αvβ3) and have recently been shown to prevent angiogenesis, tumor growth and metastasis in melanoma and lung cancer models [122]. Therefore, targeting β3 integrins may be an effective means to prevent prostate cancer metastasis and bone lesions. Platelets derived from prostate cancer patients with metastases were found to be more reactive than those with only primary disease and platelet depletion in an orthotopic LNCaP-C42 model of metastatic prostate carcinoma prevented bone metastasis and tumor induced osteogenesis [123]. Additionally, platelet ablation reduced serum levels of osteocalcin and alkaline phosphatase in tumor bearing mice suggesting decreases in bone-matrix turnover. Analysis of platelet granules derived from tumor-bearing mice revealed a significant increase in VEGF and TGFβ, factors both shown to modulate osteoblast maturation and bone formation, as well as several MMPs, including MMP-1, -3, and -13, also known to be important in bone matrix remodeling. Collectively, these studies implicate that cancer induced release of platelet stored factors contributes to bone remodeling and may be critical for the colonization and progression of prostate cancer to bone metastases. By proxy, the data also suggest indirect roles for megakaryocytes in promoting bone metastasis.

6 Innervation of the vicious cycle

Cancer associated bone pain is common in patients with prostate to bone metastases since the bone tissue is considerably innervated by both sensory and sympathetic nerve fibers [124, 125]. Studies have highlighted that heightened osteoclast activity can lead to acidosis.
and the resultant protons can stimulate nociceptors on the surface of sensory neurons [126]. Recently, new data has also revealed the importance of neuronal factors and the sympathetic nervous system in controlling bone homeostasis and cancer progression in the bone microenvironment.

6.1 Sympathetic nervous system (SNS)

The SNS has been shown to be primarily involved in early stages of tumor development, with the parasympathetic nervous system important for metastasis to the bone. The density of sympathetic and parasympathetic nerve fibers within and surrounding the human prostate samples is associated with the aggressiveness of the tumor and a poor clinical outcome [127]. Recent studies have highlighted the roles of the SNS and the PNS in driving prostate cancer progression and metastasis [127]. However, once tumor cells metastasize to the bone, the SNS may also play a role in cancer progression. The SNS maintains homeostasis of many essential functions, as well as responding to stress signals as a part of the “fight or flight” response. The role of the sympathetic nervous system in bone homeostasis was first observed due to the effect of adipocyte secreted hormone leptin on decreasing bone mass [128]. Leptin stimulates the sympathetic nerves to release the neurotransmitter noradrenaline that in turn acts on beta-androgenic receptors (βAR) on the osteoblast surface leading to reduced activity. βARs are expressed on a variety of cells involved in the vicious cycle including: tumor cells, osteoclasts, osteoblasts and macrophages. Activation of the sympathetic nervous system in a stress-induced breast cancer model resulted in a significant increase in the number of osteolytic lesions. This was primarily due to the βAR activation on osteoblasts leading to elevated levels of RANKL [129]. These findings are in agreement with studies illustrating the direct effects of βAR stimulation on osteoclastogenesis via reactive oxidase species [130]. Activation of βAR can also impact prostate cancer cells by inhibiting apoptosis and increasing migration in vitro. Induction of stress in a xenograft model of prostate cancer enhanced cancer growth and could be inhibited via the application of a βAR inhibitor [131]. Targeting the sympathetic nervous system, in particular βAR receptors, with agents such as beta-blockers is likely to significantly contribute to our ability to treat prostate to bone metastases. In several recent retrospective studies, the use of beta-blockers was shown to increase survival in a number of solid tumors, including in prostate cancer patients receiving androgen deprivation therapy [132].

7. Interrogating the expanded vicious cycle

While our understanding of the cellular and molecular factors that control the vicious cycle has become clearer, the degree of complexity as to how the multiple cell types interact with each other over time represents a major challenge to determining the impact of specific growth factors/cytokines or targeted therapies in the progression of prostate to bone metastases. Recent advances in computational modeling however and their integration with biological models of the disease may provide a way forward for researchers to reduce background noise and focus in on major processes being utilized by cancer cells to establish and grow in the bone microenvironment.

Computational modeling of cancer has become a practical tool to understanding fundamental principles and specific features in cancer [133]. These models have been
proven useful in the development of new hypotheses, in the tackling of complex interactions and testing putative therapies [134, 135]. With the increasing power of computer technology, it is now feasible and practical to directly address the interactions between the individual components of the prostate cancer bone microenvironment over clinically relevant periods [136]. In this regard, hybrid cellular automata (HCA) are beginning to be implemented as a means to model key aspects of cancer dynamics. HCA are a class of spatially and temporally discrete dynamic systems [137]. They can incorporate many features of self-organizing, complex systems, and are able to exhibit emergent behavior. Cancer models that rely on cellular automata consider the simulation of a biological process that utilizes known biological parameters to inform rules of interaction [138]. Actions in the model are carried out by these predefined rules whose conditions may or may not be satisfied by a given situation but ultimately yield a deterministic outcome that can be predictive and tested biologically. For example, HCAs have been successfully used to model complex cancer phenomena such as angiogenesis and how the stroma impacts the somatic evolution of prostate cancer cells [139, 140]. Computational modeling is also being utilized to understand the cellular dynamics of the bone microenvironment and partial differential equations (PDEs) have been generated that describe the spatio-temporal interactions between osteoblasts, osteoclasts and osteocytes [141-143]. In this model, RANKL and OPG were considered to be the principal biochemical factors controlling the interactions between the bone cells during trabecular bone remodeling. The output of the model hypothesizes that the spatial localization of RANKL and OPG are critical in directing the bone remodeling unit. Importantly, the model can assess bone formation over clinical relevant periods of time (6 to 12 months) in a short simulation period, a characteristic would be difficult and expensive to determine using in vivo models. Applying a higher-level paradigm of modeling such as HCAs will allow us to assess the biological stochasticity of multiple cells and factors of interest over time in the setting of prostate to bone metastases. A major challenge however, is that the design of the computational model is critically dependent on the quality of the biological information used for parameterization. Therefore, utilizing quality clinical and biological empirical information will be key for the development of robust models that can be used to investigate regulators of the vicious cycle of prostate to bone metastases.

8 Conclusions

For several years, the paradigm of the vicious cycle has been limited to three major cell types, namely, prostate cancer cells, osteoblasts and osteoclasts and the factors that facilitate the interaction between these cell types. These studies have yielded important therapeutic targets such as RANKL based inhibitors and bisphosphonates. In the last 5 years, there has been a significant expansion in our knowledge of the cellular and molecular factors that drive the vicious cycle, especially in regards to stem and immune cell contributions. It stands to reason that if clinical successes such as Zometa and Xgeva® can be born out of studies into the factors that control the vicious cycle, then further dissection of how stem cells, immune cells and neurons control the process can yield similar successes that not only improve quality of life but significantly extend overall survival. Understanding how multiple cellular interactions occur over time, especially in the presence of putative inhibitors, represents a major challenge given the current biological tools. However, the integration of
global system level approaches such as computational model with biological data offers a unique means with which to tackle these complexities and accelerate the discovery of therapies that will cure prostate to bone metastases.

Acknowledgments

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References


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Figure 1. Role of exosomes, HPCs and MSCs in the establishment and outgrowth of prostate to bone metastases
1. Prior to the arrival of the prostate metastases in bone, exosomes released from the primary tumor can facilitate the establishment of the pre-metastatic niche via mechanisms such as transfer of MET. 2. Activated metastases establish and grow in the bone microenvironment via the expression of VCAM-2 and the recruitment of osteoclast precursors. 3. Tumor derived CXCL16 and CCL2 can promote MSC recruitment and differentiation into CAFs or osteoblasts. 4. Enhanced osteoblast activity and expression of RANKL can in turn drive osteoclastogenesis and bone resorption. 5. Cancer recruitment of HPCs can also lead to
osteoclast mediated bone resorption resulting in a release of growth factors that further stimulate the growth of the metastases. Inhibitors of the various molecules that control cell-cell interactions are highlighted in red.
Figure 2. Immune regulation of bone metastasis

1. Dendritic cells in the bone microenvironment can contribute to T-cell polarization resulting in an immuno-evasion of the prostate metastases. Dendritic cells can also influence osteoclast behavior via a number of molecules.

2. T-cells can act directly or indirectly through the regulation of osteoclast function on the growth of prostate to bone metastases.

3. MDSCs can impact the function of several cell types in the tumor-bone microenvironment and can also directly contribute to prostate cancer growth and osteoclastogenesis.

4. Macrophages, are commonly found in the bone microenvironment and can be recruited by cancer derived factors such as CCL2. They can promote cancer cell invasion and angiogenesis while in parallel suppress cytotoxic T-cell activity. Macrophages have also been identified to play roles in regulating bone resorption. Therapies targeting the cellular and molecular mediators are highlighted in red.
Figure 3. Megakaryocytes/platelets and the sympathetic nervous system (SNS) contribute to tumor progression in bone

1. Megakaryocyte-derived platelets produce tumor-promoting growth factors and cytokines promoting extravasation and colonization of the bone compartment by prostate to bone metastases. 2. Resident megakaryocytes in the bone marrow stimulate osteoblast activity and inhibit osteoclast differentiation by secreting osteoprotegrin. 3. Nerve cells have recently been shown to play major roles in the cancer-bone microenvironment by regulating osteoblast differentiation, osteoclast activity and tumor growth. Events for potential therapeutic intervention and respective available therapies are highlighted in red.
Table 1

Cell Types/Interactors and their roles in the expanded vicious cycle of prostate to bone metastases.

<table>
<thead>
<tr>
<th>Cell Types/Interactors</th>
<th>Function</th>
<th>References</th>
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<tbody>
<tr>
<td>Exosomes/Oncosomes</td>
<td>• Educates bone-marrow derived cells (development of pre-metastatic niche)</td>
<td>[13, 22]</td>
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<td></td>
<td>• Induces differentiation of MSCs into fibroblasts</td>
<td></td>
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<tr>
<td>Hematopoietic Stem Cells</td>
<td>• Colocalize with PCa in bone</td>
<td>[35, 36]</td>
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<td></td>
<td>• Can be stimulated by PCa to differentiate into osteoclasts</td>
<td></td>
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<tr>
<td></td>
<td>• Promote differentiation of MSCs into osteoblasts upon cues from PCa</td>
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<tr>
<td>Mesenchymal Stem Cells</td>
<td>• Regulates HSC niche</td>
<td>[42, 44, 45, 50, 55]</td>
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<tr>
<td></td>
<td>• Home to tumor and produces LOX</td>
<td></td>
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<td></td>
<td>• Produces Dkk-1</td>
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<tr>
<td></td>
<td>• Inhibits angiogenesis and reduces growth by inhibiting Akt activation</td>
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<tr>
<td>Cancer-associated fibroblasts</td>
<td>• Promotes PCa growth in bone</td>
<td>[46]</td>
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<tr>
<td>Adipocytes</td>
<td>• Increases osteoblast differentiation, proliferation, and mineralization</td>
<td>[58, 59]</td>
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<tr>
<td></td>
<td>• Increases PCa migration and proliferation</td>
<td></td>
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<tr>
<td>Megakaryocytes</td>
<td>• Inhibit osteoclast differentiation</td>
<td>[114]</td>
</tr>
<tr>
<td></td>
<td>• Stimulate osteoblast proliferation and differentiation</td>
<td></td>
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<tr>
<td></td>
<td>• Decrease tumor cell proliferation and induce apoptosis</td>
<td></td>
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<tr>
<td>Platelets</td>
<td>• Inhibit tumor adhesion to endothelium</td>
<td>[116, 117, 123]</td>
</tr>
<tr>
<td></td>
<td>• Release VEGF and TGFβ</td>
<td></td>
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<tr>
<td></td>
<td>• Modulates osteoblast differentiation and bone formation</td>
<td></td>
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<tr>
<td>MDSCs</td>
<td>• Suppress T-cells activity</td>
<td>[68, 70-72]</td>
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<tr>
<td></td>
<td>• Pro-angiogenic</td>
<td></td>
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<tr>
<td></td>
<td>• Differentiate into osteoclasts, promoting osteolysis</td>
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<tr>
<td></td>
<td>• Secrete TGFβ</td>
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<tr>
<td>Dendritic Cells</td>
<td>• Secrete cytokines (eg. CCL2) which stimulate osteoclast activity</td>
<td>[96]</td>
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<tr>
<td></td>
<td>• &quot;Educated&quot; dendritic cells can stimulate T-cells to attack tumor cells</td>
<td></td>
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<tr>
<td>T Cells</td>
<td>• Express RANKL stimulating osteoclast formation</td>
<td>[100-102, 108]</td>
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<td></td>
<td>• Express IFN-gamma which inhibit osteoclast formation</td>
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<td></td>
<td>• Activate gamma-delta T-cells produce pro-inflammatory cytokines</td>
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<td>Nervous System</td>
<td>• Increased pain due osteoclast proton stimulation of nociceptors</td>
<td>[125, 129]</td>
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<tr>
<td></td>
<td>• Reduce osteoblast activity</td>
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<tr>
<td></td>
<td>• Increase RANKL expression on osteoblast, increasing osteoclast formation</td>
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