Genetic analysis reveals the wild ancestors of the llama and the alpaca

Miranda Kadwell¹, Matilde Fernandez², Helen F. Stanley¹, Ricardo Baldí³, Jane C. Wheeler⁴, ⁵, Raul Rosadio² and Michael W. Bruford¹, ²

¹Institute of Zoology, Regent’s Park, London NW1 4RY, UK
²Facultad de Medicina Veterinaria, Universidad Nacional Mayor de San Marcos, Apartado 41-0068, Lima 41, Peru
³Centro Nacional Patagónico, Consejo Nacional de Investigaciones Científicas y Tecnológicas, 9120 Puerto Madryn, Argentina
⁴CONOPA, Las Cerezos 106, Salamanca-Lima 3, Peru
⁵Biodiversity and Ecological Processes Group, Cardiff School of Biosciences, Cathays Park, Cardiff CF10 3TL, UK

The origins of South America’s domestic alpaca and llama remain controversial due to hybridization, near extinction during the Spanish conquest and difficulties in archaeological interpretation. Traditionally, the ancestry of both forms is attributed to the guanaco, while the vicuña is assumed never to have been domesticated. Recent research has, however, linked the alpaca to the vicuña, dating domestication to 6000–7000 years before present in the Peruvian Andes. Here, we examine in detail the genetic relationships between the South American camels in order to determine the origins of the domestic forms, using mitochondrial (mt) and microsatellite DNA. MtDNA analysis places 80% of llama and alpaca sequences in the guanaco lineage, with those possessing vicuña mtDNA being nearly all alpaca or alpaca–vicuña hybrids. We also examined four microsatellites in wild known-provenance vicuña and guanaco, including two loci with non-overlapping allele size ranges in the wild species. In contrast to the mtDNA, these markers show high genetic similarity between alpaca and vicuña, and between llama and guanaco, although bidirectional hybridization is also revealed. Finally, combined marker analysis on a subset of samples confirms the microsatellite interpretation and suggests that the alpaca is descended from the vicuña, and should be reclassified as Vicugna pacos. This result has major implications for the future management of wild and domestic camels in South America.

Keywords: domestication; alpaca; guanaco; llama; vicuña; mitochondrial DNA

1. INTRODUCTION

Four South American camelids are recognized today, two of which are wild species, the guanaco (Lama guanicoe, Muller 1776) and the vicuña (Vicugna vicugna, Molina 1782), and two of which are domestic forms, the alpaca (Lama pacos L) and the llama (Lama glama L), whose evolutionary origins are debated (Wheeler 1995). The guanaco and vicuña diverged from a common ancestor around two million years ago, and are the only representatives of the lamid to survive the Pleistocene period (Stanley et al. 1994; Wheeler 1995). Archaeological evidence from the central Peruvian Andes links alpaca origins to the vicuña at 6000–7000 years before present (Wheeler 1995). Because all potential ancestral forms are extant, South American camelid domestication represents an unusual and useful opportunity to gain insight into the origin and biodiversity of domesticated animals, an issue which is of increasing interest due to the recognized potential economic benefits of indigenous genetic resources and the threats that face marginal and extensive agriculture today (Hall & Bradley 1995). In contrast with many other domestic farm animals, there is no written history associated with the llama and alpaca. Orally transmitted herding knowledge was largely lost during the Spanish conquest, and breeds disappeared as both the human and the native domestic livestock populations were reduced by 80–90% during the first 100 years of contact (Wheeler 1995). At present, although llama and alpaca rearing is a central element of the economy in the high Andes, it is often not profitable due to the poor quality of the animals and their fibre. The reconstruction of fine-fibre breeds and the breeding strategies needed are therefore uniquely dependent upon the contributions of archaeozoology and genetic analysis (Wheeler et al. 1995).

Understanding the domestication of the llama and alpaca using morphological analyses has been hampered by the lack of species-specific post-cranial skeletal characteristics, and because archaeological remains are often in poor condition (Bahn 1994). Fibre analysis has been pursued at sites where fleece has been preserved (Dramart 1991a; Reigadas 1993; Wheeler et al. 1995), but the decimation of traditional llama and alpaca breeds at the time of the conquest, and probable subsequent hybridization among the survivors, make inference from phenotypic characters problematic.

Molecular genetic analysis using methods such as mitochondrial DNA (mtDNA) sequencing and microsatellite and Y-chromosome analysis has recently proved illuminating when examining the origins of modern domestic livestock, even in the face of often overwhelming recent changes in domestic populations due to genetic drift, selection and/or hybridization (e.g. Bradley et al. 1996; MacHugh et al. 1997; Hendler et al. 1998; Luikart et al. 2001; Vila et al. 2001). Here, we apply mitochondrial cytochrome b sequence and microsatellite typing to the
question of the evolutionary relationships between and
domestication of South American camelids throughout
their range in South America today.

2. MATERIAL AND METHODS

(a) Samples

Sample collection sites span as far as possible the geo-
ographical range of the two wild species (see figure 1). Our
sample comprises: two vicuña subspecies (*V. v. vicugna* and
*V. v. menoloides*) from Argentina, Chile and Peru, guanaco
(*L. g. guanicoe* and *L. g. caullaini*) from Argentina and Peru,
llama (a range of morphological types) from Argentina, Bolivia
and Peru, alpaca (including ‘surí’ and ‘huacaya’ fleece types)
from Argentina, Bolivia, Chile and Peru, and 10 known hybrids
(seven between llama and alpaca (locally known as warí) and
three between alpaca and vicuña (locally known as pacovi-
űña). Samples were taken from only those individuals whose
phenotype corresponded to accepted morphological criteria for
domestic forms. The localities and genetic data for each indivi-
dual included in this study can be found at http://www.cardiff.
ac.uk/bioscience/biodiversity/staff/mh.html.

(b) MitDNA analysis

The phylogenetic affiliations of the llama and alpaca were
first confirmed by sequencing a short (138 bp) but highly infor-
mative region of the cytochrome b gene of the mitochondrial
genome *sensu* Stanley *et al.* (1994) (GenBank accession
numbers U06425-U06430). In total, 21 South American camelids were
analysed from throughout the geographical range (comprising
21 guanaco, 42 vicuña, 54 llama, 84 alpaca and the 10 hybrids
described in table 1). One Arabian camel was also analysed.
Briefly, DNA was extracted from blood or skin using standard
proteinase-K digestion followed by organic extraction using
phenol and phenol–chloroform, and total DNA was precipitated
in 100% ethanol (Stanley *et al.*, 1994; Bruford *et al.*, 1998). DNA
samples were stored in TE buffer (10 mM Tris–HCl, 1 mM
EDTA, pH 8.0). The cytochrome b primers LH742 and HH4900
were used for polymerase chain reaction (PCR), which was
carried out as in Stanley *et al.* (1994). PCR products were puri-
fied and DNA sequencing was carried out as previously
described (Stanley *et al.*, 1994).

Sequences were aligned manually, and unique haplotypes have
been deposited in GenBank under accession numbers
AF373809-AF373833. Mitochondrial haplotype divergence and
frequency were analysed for the guanaco, vicuña, llama and
alpaca samples as previously described (Stanley *et al.*, 1994), and
a minimum-spanning network (Kruskal 1956; Bandelt *et al.*, 1999)
was generated using the program MrNeXet (Excoffier 1993;
figure 2). The distribution patterns of domestic South American
camelid haplotypes were then compared with those of the wild
South American camelid sample.

(c) Microsatellite analysis

Since the strict maternal inheritance of mtDNA in most
mammals restricts its use in studies of hybridization, especially
dominant livestock (e.g. MacHugh *et al.*, 1997), we also applied
nuclear DNA markers. Four microsatellite loci (*YW11* 38,
*YW11* 43, *YW11* 46 and *LCA* 19; Lang *et al.*, 1996; Penedo
*et al.*, 1998) were typed for 669–711 individuals, including the 21
individuals for cytochrome b (table 1). The genetic distances
between the four South American camelid taxa were measured
in three ways: first, by using 1 - (proportion of shared alleles);
second, by using Reynolds’s distance (Reynolds *et al.*, 1983), a
measure commonly used in livestock analysis where genetic drift
has a major impact on allele frequencies; and finally, by using
$\delta^2$ (Goldstein *et al.*, 1995), which calculates population sub-
division using allele size differences under a stepwise mutation
model. All distances were estimated using the program
Microsat v. 1.5d (Minch 1999).

Factorial correspondence analysis was then performed on
pairwise allele frequency differences using GENETIX v. 4 (Belkhir
1999). Here, the genetic differentiation between populations (in
this case vicuña, guanaco, llama and alpaca) is expressed as
factors, which explain the correspondence between samples in a
number of dimensions (Benzécri 1973). Thus, the relationships
among populations can be judged by examining how individuals
from each population cluster in two or three dimensions.

(d) Combined analysis

To assess the extent of introgression in llama and alpaca
populations, we used both the microsatellite and the mitochondrial
data to calculate the admixture proportions mC and mY
(mC: estimator based on allele frequencies, Chakraborty
*et al.*, 1992; mY: estimator based on the number of substitutions
for mtDNA and squared allele size difference for microsatellite
data, Bertorelle & Excoffier (1998) for alpaca and llama com-
pared with the vicuña and guanaco. This was implemented
using the program Amix 1.0 (Bertorelle 1998).

Finally, to examine the concordance between mitochondrial
and microsatellite data and patterns of introgression in more
detail, we returned to the 21 individuals typed for both mtDNA
and microsatellites. Genotypes were coded ‘V’ (vicuña) or ‘G’
(guanaco) for mtDNA and ‘V’; ‘G’ or ‘H’ (hybrid) for *YW11* 46
and *LCA* 19, depending on their allele sizes with reference to
the guanaco and vicuña ranges, and we examined the data for
each locus separately and combined.

3. RESULTS

(a) Mt DNA

We found 26 unique haplotypes within the 21 South
American camelids analysed from throughout the geo-
ographical range. Uncorrected distances within the South
American camelids ranged from 0.006 (one substitution)
 to 0.089 (14 substitutions). The minimum-spanning
network (figure 2) recovers two groups, recapitulating
the reciprocally monophyletic clades found previously
(Stanley *et al.*, 1994), the first group contains all vicuña
(V), and the other contains all guanaco (G). Both groups
are connected by 21 substitutions to the Arabian camel.
The domestic South American camelids are found in
both groups, but 81% (120 out of 148, including 61 out of
84 alpaca) are found within the G group. A minority
(28 out of 148) comprising alpaca (n = 23), pacovićuña
(n = 3) and llama (n = 2) are found within the V group.

(b) Microsatellites

A feature of the microsatellite loci analysed here is the
large number of private alleles found when comparing
vicuña and guanaco. These private alleles, which range
between 33% for *YW11* 38 and 100% for *LCA* 19 and
YW11 46, occupy predominantly different allele size
ranges. Figure 3a(1) and 3b(1) show allele frequency
histograms for *LCA* 19 and YW11 46 for wild vicuña
and guanaco: the allele sizes do not overlap between the

two species. These loci therefore provide potentially powerful tools for the discrimination of ancestral genomes in modern domestic stock. Figure 3a(ii) and 3b(ii) show histograms for the same loci in llama and alpaca, which display similar patterns. However, the patterns of genetic similarity are in contrast to those
Table 1. South American camelid samples analysed for mitochondrial DNA (cytochrome b) and four microsatellite loci.

<table>
<thead>
<tr>
<th></th>
<th>cytochrome b</th>
<th>YWLL 38</th>
<th>LCA 19*</th>
<th>YWLL 46*</th>
<th>YWLL 43</th>
</tr>
</thead>
<tbody>
<tr>
<td>vicuña</td>
<td>42</td>
<td>434</td>
<td>439</td>
<td>438</td>
<td>440</td>
</tr>
<tr>
<td>guanaco</td>
<td>21</td>
<td>112</td>
<td>120</td>
<td>122</td>
<td>115</td>
</tr>
<tr>
<td>wari</td>
<td>7</td>
<td>7</td>
<td>7</td>
<td>7</td>
<td>7</td>
</tr>
<tr>
<td>pacovicuña</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
<td>3</td>
</tr>
<tr>
<td>llama</td>
<td>54</td>
<td>50</td>
<td>60</td>
<td>60</td>
<td>60</td>
</tr>
<tr>
<td>alpaca</td>
<td>84</td>
<td>63</td>
<td>138</td>
<td>141</td>
<td>103</td>
</tr>
</tbody>
</table>

* Indicates loci with non-overlapping allele size ranges.

Figure 2. Minimum-spanning network representing the relationships between cytochrome b mitochondrial haplotypes as circular nodes, where the number of substitutions (if greater than one) are represented by multiple dashes or numbers on each connecting line. The relative frequency of each haplotype is represented by the area of the circle. Unfilled circles indicate vicuña haplotypes, and filled circles represent guanaco haplotypes. Wild samples are specifically referred to where present. Phylogenetic analysis (maximum parsimony and neighbour joining based on uncorrected p, JC and K2P sequence distances) recovered an equivalent pattern to the network, and high (> 90%) bootstrap support was always found for the major split between ‘V’ and ‘G’ haplotypes (not shown).

revealed by mtDNA. Visual inspection reveals strong similarities between the allele size distributions of vicuña and alpaca, and between guanaco and llama. For example, for YWLL 46, the 98 bp allele has a frequency of 0.95 in the vicuña sample and 0.75 in the alpaca, while the 104 and 106 bp alleles have a combined frequency of 0.91 in the guanaco and 0.64 in the llama. Analysis of all four loci reveals that genetic distances between the vicuña and alpaca and between the guanaco and llama (table 2) are almost always much lower than those between the vicuña and guanaco, the vicuña and llama or the guanaco and alpaca. Distances between the alpaca and llama are intermediate, with the exception of δμ^2, where the distance between the guanaco and llama is slightly larger.

However, a second feature of the histogram in figure 3b is the presence, at low frequencies, of ‘vicuña’ alleles in the llama sample and of ‘guanaco’ alleles in the alpaca sample. For example, in LCA 19, the 99 bp and 103 bp ‘vicuña’ alleles are present at a combined frequency of 0.063 in the llama sample, and the 87 bp ‘guanaco’ allele has a frequency of 0.119 in the alpaca. The presence of these alleles in our sample confirms bidirectional introgression in both domestic forms.

A striking pattern emerges from the factorial correspondence analysis (figure 4), where almost half of the
Figure 3. Allele frequency histograms for a large sample of (i) vicuña and guanaco and (ii) llama and alpaca for (a) LCA 19 and (b) YWLL 46. Distributions were generated using the following: guanaco, \( n = 104 \) (LCA 19) and \( n = 177 \) (YWLL 46); llama, \( n = 56 \) (LCA 19) and \( n = 58 \) (YWLL 46); alpaca, \( n = 80 \) (LCA 19) and \( n = 82 \) (YWLL 46); and vicuña, \( n = 227 \) (LCA 19) and \( n = 231 \) (YWLL 46).
Table 2. Pairwise genetic distances between the four South American camels. (a) Above diagonal: Reynolds distance; below diagonal: $\delta^2$. (b) Below diagonal: 1 – proportion of shared alleles.

<table>
<thead>
<tr>
<th></th>
<th>vicuña</th>
<th>guanaco</th>
<th>alpaca</th>
<th>llama</th>
</tr>
</thead>
<tbody>
<tr>
<td>vicuña</td>
<td>—</td>
<td>0.729</td>
<td>0.173</td>
<td>0.627</td>
</tr>
<tr>
<td>guanaco</td>
<td>28.928</td>
<td>—</td>
<td>0.433</td>
<td>0.174</td>
</tr>
<tr>
<td>alpaca</td>
<td>1.089</td>
<td>19.781</td>
<td>—</td>
<td>0.267</td>
</tr>
<tr>
<td>llama</td>
<td>17.162</td>
<td>10.784</td>
<td>9.892</td>
<td>—</td>
</tr>
</tbody>
</table>

(b)

<table>
<thead>
<tr>
<th></th>
<th>vicuña</th>
<th>guanaco</th>
<th>alpaca</th>
<th>llama</th>
</tr>
</thead>
<tbody>
<tr>
<td>vicuña</td>
<td>—</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>guanaco</td>
<td>0.963</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>alpaca</td>
<td>0.337</td>
<td>0.841</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>llama</td>
<td>0.522</td>
<td>0.522</td>
<td>0.616</td>
<td>—</td>
</tr>
</tbody>
</table>

Figure 4. Two-dimensional factorial correspondence plot for allele frequencies at four microsatellite loci in all South American camelids. Almost half of the explained correspondence (15%) is found in factors 1 and 2, represented on the horizontal and vertical axes, respectively.

explained variance (15%) is found in factors 1 and 2, represented on the horizontal and vertical axes, respectively. It can be clearly seen that the guanaco (black squares) and vicuña (grey, filled diamonds) form two tightly clustered and highly distinct groups. The alpaca (grey, filled triangles) also form a cohesive group and cluster strongly with the vicuña. In contrast, the llamas (open squares) and hybrids form a much more diffuse group. The llamas, although tending to cluster with guanaco on axis 1, are more intermediate with respect to the wild species when compared with the alpaca sample, and are also the most genetically diffuse group on axis 2.

(c) Combined data

The admixture results are non-concordant between the mitochondrial and microsatellite analyses in the alpaca, where the estimated microsatellite proportion of vicuña genome is two to three times higher than the proportion estimated using mtDNA (0.310 ± 0.121 for mtDNA; 0.903 ± 0.108 for mtC and 0.823 ± 0.087 for mtY for microsatellites). Furthermore, in the llama, although both estimates are relatively low, the microsatellite admixture proportions are an order of magnitude higher (0.0231 ± 0.242 for mtDNA; 0.220 ± 0.088 for mtC and 0.389 ± 0.109 for mtY for microsatellites). Importantly, it is evident using both mtC and mtY that the proportion of vicuña DNA is much lower in llama than in alpaca.

The combined three-locus analysis of the 211 individuals produces striking results (table 3). Out of the 54 llamas, 32 (96%) possessed a ‘G’ mtDNA haplotype, and 47 (90%) and 33 (61%) possessed a pure ‘G’ genotype for LCA 19 and YWIL 46, respectively. Out of the 84 alpacas, only 23 (27%) possessed a ‘V’ mtDNA haplotype, while 63 (75%) and 66 (79%) possessed pure ‘V’
Table 3. Three-locus genotypes for samples where all three types of data are available. Loci are ordered mitochondrial DNA, LCA 19 and YWLL 46; so, for example, GVH indicates guanaco mitochondrial DNA, vicuña genotype at LCA 19 and a hybrid genotype at YWLL 46 (X signifies that the sample could not be typed).

<table>
<thead>
<tr>
<th></th>
<th>vicuña</th>
<th>alpaca</th>
<th>guanaco</th>
<th>llama</th>
<th>wari</th>
<th>pacovicuña</th>
</tr>
</thead>
<tbody>
<tr>
<td>GGG</td>
<td></td>
<td></td>
<td>21</td>
<td>32</td>
<td></td>
<td></td>
</tr>
<tr>
<td>GGH</td>
<td></td>
<td></td>
<td></td>
<td>15</td>
<td></td>
<td></td>
</tr>
<tr>
<td>GGV</td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>GHH/GHX/GXH</td>
<td>2/1/1</td>
<td></td>
<td></td>
<td>2</td>
<td></td>
<td></td>
</tr>
<tr>
<td>GXV</td>
<td></td>
<td>1</td>
<td></td>
<td>2</td>
<td></td>
<td></td>
</tr>
<tr>
<td>GVH/GHV</td>
<td>18</td>
<td></td>
<td></td>
<td>1</td>
<td></td>
<td></td>
</tr>
<tr>
<td>GVG</td>
<td></td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>GVV/GXV</td>
<td>34/1</td>
<td></td>
<td></td>
<td>4</td>
<td></td>
<td></td>
</tr>
<tr>
<td>VVV</td>
<td>42</td>
<td>17</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>VVH/VHV</td>
<td>3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>VGG</td>
<td></td>
<td></td>
<td></td>
<td>2</td>
<td></td>
<td></td>
</tr>
<tr>
<td>VGV</td>
<td></td>
<td>1</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>VHH</td>
<td></td>
<td>2</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

LCA 19 and YWLL 46 genotypes, respectively. Out of the llamas tested in this study, 32 (59%) exhibited a ‘GGG’ three-locus genotype; only 17 (20%) alpacas exhibited a ‘VVV’ three-locus genotype. Extensive nuclear introgression is detected in the llamas studied here, with 20 (37%) showing one or more ‘vicuña’ alleles at LCA 19 and/or YWLL 46. In contrast, much of the presumed introgression in the alpaca is mitochondrial, with 34 (40%) samples showing a ‘GVV’ three-locus genotype.

4. DISCUSSION

The data resulting from this study present a complex picture of how modern-day domestic South American camelid genetic diversity has been shaped by domestication, selection, hybridization and genetic drift, with the added uncertain impact of events relating to the Spanish conquest. Such complexity is not without precedence in studies of domestic livestock (e.g. MacHugh et al. 1997), and requires detailed analysis and cautious interpretation.

Taken in isolation, the expanded mtDNA analysis presented here largely supports the findings of Stanley et al. (1994). Wild vicuña and guanaco mtDNA is reciprocally monophyletic with 5.8-8.9% uncorrected sequence divergence being found between the two lineages, recapitulating the suggestion in Stanley et al. (1994) that these species diverged from a common ancestor two to three million years ago. Furthermore, the finding that nearly all modern llamas possess a ‘guanaco’ haplotype is also supported by the present dataset (table 3), where all except two llamas are from a sample of 54 individuals had guanaco mtDNA. However, our much expanded alpaca dataset reveals a different pattern from that previously found, with only 27% of individuals possessing vicuña mtDNA (table 3), in contrast to the 50% reported by Stanley et al. (1994). It is worth noting here that our sample comprises 84 individuals (as opposed to 14) sampled from throughout the geographical range. In isolation, the finding that a large proportion of modern-day alpacas possess guanaco mtDNA is in accordance with hypotheses that alpacas, in common with llamas, are descended from the guanaco. However, clearly, as in Stanley et al. (1994), the presence of substantial numbers of alpaca possessing vicuña mtDNA also raises the possibility that the alpaca is of mixed origin or has undergone substantial hybridization during domestication or subsequently. However, the limitations of mtDNA, which is maternally inherited, in the context of gene flow and evolution in domestic populations are obvious, since historical and modern-day agricultural practices have often used phenotypically desirable males to sire large numbers of females. Therefore, analysis of biparentally inherited markers (in this case microsatellites) capable of discriminating between the wild ancestors was desirable.

The microsatellite data provide a contrast to the mtDNA data, and the existence of two loci with non-overlapping allele size ranges in the wild ancestors allowed us to compare patterns of divergence in relatively large numbers of domestic animals. In contrast to the mtDNA results, visual inspection of allele frequency distributions, genetic-distance analysis using three measures with contrasting models and assumptions, and factorial correspondence analysis all reveal a striking similarity between the alpaca and the vicuña (figures 3a,b, 4 and table 2a,b). Each genetic-distance estimate is lowest for the alpaca–vicuña comparison, and the factorial correspondence analysis (figure 4) shows that the alpaca and vicuña occupy an almost overlapping region of the plot. These data, therefore, point towards a very close genetic affinity between the alpaca and vicuña, a finding in complete contrast to the mtDNA data.

The microsatellite data in part agree with the mtDNA data by supporting a close relationship between the llama and guanaco. Out of the genetic-distance estimates, both Reynold’s and allele sharing distances are second lowest for the guanaco–llama comparison (table 2a,b), and the Reynold’s distance estimate is almost identical to that for the alpaca–vicuña comparison. However, other data are more equivocal, with the factorial correspondence plot revealing a dispersed pattern for the llama, which could be described as intermediate between the vicuña and guanaco, and the $\delta_{hu}$ distances being slightly lower for the llama–alpaca comparison than for the llama–guanaco comparison. Although none of the above are indicative of a close relationship between the llama and vicuña, they suggest either that there has been nuclear
gene flow between the llama and vicuña (or, more likely, between the llama and alpaca) or that the ancestral stock for the present-day llama was genetically highly diverse and that this is reflected in present-day microsatellite allelic diversity. The guanaco and llama have much greater geographical ranges than the vicuña and alpaca, which may have led to greater historical intraspecific differentiation, reflected in greater diversity in nuclear and mtDNA (where we found 21 guanaco haplotypes, as opposed to five in the vicuña). Analysis of greater numbers of microsatellite loci (only four were used here) is highly desirable, and could include some of those described by Sarno et al. (2000), where at least two additional loci suggest non-overlapping allele distributions in the wild species. Archaeozoological evidence is inconclusive concerning llama domestication, but it is possible that there may have been more than one centre (Wheeler 1995).

The suggestion of substantial mitochondrial introgression in the alpaca and nuclear introgression in the llama is substantiated when admixture is measured for both marker types. The relatively low estimated admixture proportion of vicuña mtDNA present in the alpaca (0.31) is in contrast with the high proportion estimated for the microsatellites (0.82–0.90). Further, the extremely low admixture proportion of vicuña mtDNA in the llama (0.02) contrasts strongly with microsatellite estimates (0.22–0.39), which also suggest substantial nuclear admixture in the llama. These data must, however, be treated with caution, since the assumptions underlying the admixture models used in this analysis are highly likely to be violated in South American camels, where introgression events are likely to have been recurrent, and may have increased in frequency during and after the Spanish conquest. Furthermore, the drastic reductions in domestic South American camelid populations associated with the conquest are likely to have had significant and unpredictable impacts on allele frequencies.

Inspection of the distribution of the three-locus genotypes in modern alpacas and llamas confirms many of the above findings. Only 27% of alpacas are mitochondrially ‘vicuña’, although 40% of alpaca possess only vicuña microsatellite alleles with guanaco mtDNA haplotypes. Such a pattern suggests that introgression of guanaco (or, more likely, llama) mtDNA some time in the past may have occurred recurrently within alpaca populations, but may have been accompanied more recently by a reversion to line or stock breeding within local alpaca populations.

The lack of written records means that any such inference is speculative. Table 3 suggests that, in contrast, mitochondrial introgression has occurred much less frequently in the llama. However, although table 3 suggests that nuclear introgression is similar in the alpaca and llama, the expanded microsatellite dataset suggests that it has occurred at a higher level in the llama (possibly two or three times higher than in the alpaca from the admixture analysis), which may partly account for the more dispersed factorial correspondence pattern; this warrants further investigation.

The implications of these data are potentially important for the way in which these genetic resources are managed in the future. In our sample, only 35% of domestic animals have not undergone any detectable hybridization based on mtDNA and the two diagnostic microsatellites. In particular, there are a very large number of detectable hybrids within the alpaca population (80%), which are accentuated when using mtDNA. Also, 40% of llama show detectable signs of hybridization, but mitochondrial introgression is virtually absent.

During the last 20–25 years large-scale hybridization between llamas and alpacas has been carried out in Peru (Bustinza 1989). Specifically, male alpacas have been bred to female llamas to increase the population of animals producing higher priced ‘alpaca’ fibre, and male llamas have been bred to female alpacas to obtain greater fleece weights and, thus, increased income. With sale price traditionally determined by weight, and no consideration given to fineness, the quality of alpaca fibre has decreased markedly over the past 25 years. Indigenous Quechua- and Aymara-speaking herders subdivide the hybrids into llamawari or waritu (llama-like) and pacowari or wayki (alpaca-like), respectively, depending upon physical appearance (Flores Ochoa 1977; Dransart 1991b). The offspring are fertile, tend to be intermediate in size and can be backcrossed to either parental type. Further, recent intensive selection for white fleece in modern alpaca may also have involved bidirectional hybridization. A combination of these practices and our results could explain the taxonomic confusion surrounding the domestic forms in the recent past, as it is likely that many specimens used in previous taxonomic studies were hybrids.

Given the extreme hybridization in present-day alpacas, DNA analysis has been critical in resolving the origin of this domestic form. Since our results suggest the vicuña as the ancestor of the alpaca, we propose that the classification of the alpaca should be changed from Lama pacos L. to Vicugna pacos L.

We suggest, therefore, that the degeneration of quality and value in present-day alpacas and llamas has been due to extensive hybridization, probably beginning with the conquest and continuing to the present day. While it was believed that these crosses were between different forms of a single domestic animal descended from the guanaco, there was little concern about the economic impact of such introgression. However, given that the alpaca is probably descended from the vicuña, the negative impact of such crosses is now evident. The use of DNA analysis to identify and eliminate hybrid animals from the breeding pool is essential, since the antiquity of the ongoing hybridization process makes it impossible to accurately identify all hybrids on the basis of phenotypic characteristics. Additionally, the knowledge that the alpaca is descended from the vicuña opens new routes for the improvement of alpaca fibre production, not only through the identification of hybrids and their elimination from pure-bred elite herds, but also via the backcrossing of pure-bred alpacas to their vicuña ancestor in order, possibly, to improve fibre fineness.

While 90% of the alpaca fibre produced in Peru today has a diameter of greater than 25 μm and fetches low prices on the world market ($US3–30 per kg, 1980–1995), pre-conquest animals produced fibre of 17–22 μm in diameter (Wheeler et al. 1995), similar to cashmere (15–17 μm; $US60–120 per kg, 1980–1995). It is possible, therefore, that identification of the remaining pure
alpacas may aid in recovery of the fine fibre characteristics of pre-conquest animals. The knowledge that the alpaca is the domestic vicuña also necessitates a re-evaluation of vicuña conservation policy. Although the vicuña has been listed as endangered under the Convention on International Trade in Endangered Species of Wild Fauna and Flora (CITES Appendix I) since its inception in 1975, all Peruvian vicuñas and large segments of the Chilean, Argentine and Bolivian populations have been reclassified as threatened (CITES Appendix II), permitting controlled commercialization of live-shorn fibre. With unprocessed fibre currently valued at ca. US$405 per kg, vicuña fleece is the most expensive natural fibre in the world, and represents an important potential source of income for the extremely poor rural populations on whose lands the animals live. To date, Peru's rational use policy has produced an important increase in vicuña numbers, but demands for greater control over the species through construction of fences, intensive rearing and selection are growing. Judging by the alpaca, such interventions will, in the long run, lead to a deterioration of fibre quality and fineness (which, at 12–14 µm, is the basis of its value), and increased limitation on movement, especially of the non-territorial male bachelor bands, represents a significant new threat to this species (Wheeler et al. 2001).

We acknowledge the following people who generously helped by providing information, samples or permits for this study. In Argentina: M. Knobel, Argentine Guanaco Products, Esquel; S. Poncet; Eduardo Frank, Universidad de Cordoba; Freddy Sossa, EU Supreme Project, Jujuy; Gustavo Rebuffi, INTA Abra Pampa; Daniel Almeida, INTA Bariloche; Hector Guillermo Villanueva, Recursos Naturales, Salta; Teresa Raquel Chalaba, Universidad de Salta; G. Moeyle, BBRSIC-IGER, Aberystwyth, UK. In Chile: Hernan Torres, Conservation International, Washington DC, USA; Eduardo Nuñez and Rafael Fernández, CONAF, Arica; Calagongo Santoro, Universidad de Tarapacá, Arica. In Peru: Alfonso Martinez, Domingo Hoces, Jorge Herrera and Marco Antonio Zuñiga, CONACS, Lima; Alex Montufar and Marco Antonio Escobar, CONACS, Puno; Roberto Bohnett, CONACS, Junín; Carlos Ponce del Prado, Conservacion Internacional, Lima; Maximo Gaimarra y Santiago Baudillo, SAIS Tupac Amaru, Pachacauy; Rosa Perales, Jose Alva y Néstor Falcón, Facultad de Medicina Veterinaria, Universidad Nacional Mayor de San Marcos, Lima; Leoncio Ruiz Rios, Congreso Constituyente del Perú; Clive Woodham, Labvetsur, Arequipa; Felipe San Martin, Instituto Veterinario de Investigaciones Tropicales y de Altura, Lima. Nicola Anthony, Peter Arctander, Mark Beaumont, Jon Bridle, Kate Byrne, Louises Chikhi, Gordon Luikart, Benoit Goossens and two anonymous referees provided invaluable comments on the manuscript, and Georgio Bertorelle gave valuable advice on the admixture analysis. This study was supported by the Institute of Zoology, Instituto Veterinario de Investigaciones Tropicales y de Altura, Natural Environment Research Council grant GST/02/828 to H.F.S. and J.C.W. and Darwin Initiative grant 162/06/126 to H.F.S. J.C.W and M.W.B.

REFERENCES


Belkhir, L. 1999 GENETIX v. 4.0. Belkhir Biooff, Laboratoire des Genomes et Populations, Université Montpellier II, France.


Hendleder, S., Mainz, K., Plante, Y. & Lewalski, H. 1998 Analysis of mitochondrial DNA indicates that domestic sheep are derived from two ancestral maternal sources: no evidence for contributions from urial and argali sheep. J. Hered. 89, 113–120.


As this paper exceeds the maximum length normally permitted, the authors have agreed to contribute to production costs.