Published in final edited form as:

Phys Rev E. 2020 June ; 101(6-1): 062405. doi:10.1103/PhysRevE.101.062405.

Relationship between velocities, tractions, and intercellular stresses in the migrating epithelial monolayer

Yoav Green^{1,2,*}, Jeffrey J. Fredberg¹, James P. Butler^{1,3}

¹Harvard T.H. Chan School of Public Health, Boston, Massachusetts 02115, USA

²Department of Mechanical Engineering, Ben-Gurion University of the Negev, Beer-Sheva 8410501, Israel

³Department of Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston, Massachusetts 02115, USA

Abstract

The relationship between velocities, tractions, and intercellular stresses in the migrating epithelial monolayer are currently unknown. Ten years ago, a method known as monolayer stress microscopy (MSM) was suggested from which intercellular stresses could be computed for a given traction field. The core assumption of MSM is that intercellular stresses within the monolayer obey a linear and passive constitutive law. Examples of these include a Hookean solid (an elastic sheet) or a Newtonian fluid (thin fluid film), which imply a specific relation between the displacements or velocities and the tractions. Due to the lack of independently measured intercellular stresses, a direct validation of the 2D stresses predicted by a linear passive MSM model is presently not possible. An alternative approach, which we give here and denote as the Stokes method, is based on simultaneous measurements of the monolayer velocity field and the cell-substrate tractions. Using the same assumptions as those underlying MSM, namely, a linear and passive constitutive law, the velocity field suffices to compute tractions, from which we can then compare with those measured by traction force microscopy. We find that the calculated tractions and measured tractions are uncorrelated. Since the classical MSM and the Stokes approach both depend on the linear and passive constitutive law, it follows that some serious modification of the underlying rheology is needed. One possible modification is the inclusion of an active force. In the special case where this is additive to the linear passive rheology, we have a new relationship between the active force density and the measured velocity (or displacement) field and tractions, which by Newton's laws, must be obeyed.

I. INTRODUCTION

During wound healing, cancer metastasis, development, and asthmatic airway remodeling, cells comprising a confluent epithelial layer migrate collectively [1–5]. Within the epithelial monolayer, each constituent cell exerts intercellular stresses on neighboring cells and exerts traction forces [6] on its substrate. While traction forces exerted by a monolayer have been measured for two decades [7–9], their relationship to measured cellular velocities remains

* yoavgreen@bgu.ac.il.

unknown, and the relationship between intercellular stresses and tractions remains unresolved [10].

Ten years ago [11,12], our group suggested a method to recover the induced stresses σ within a monolayer given a measured traction vector field T . The method was termed monolayer stress microscopy (MSM) [11,12], and we denote the recovered stresses as $\sigma_{\text{MSM}}(T)$. The principle assumption was that the stresses could be described by the same equations describing simple passive systems such as Hookean solids [13] or Newtonian fluids [14,15], where there is a linear relationship between stress and strain [13] or stress and strain rate [15], respectively. Since our first work [11], this approach has been adopted by many, and various derivative formulations have been suggested (Hookean solid [10–12,16–29] or Newtonian fluid [10,18,20,30–32], see recent reviews [10,33] for more details).

It is clear that in one dimension this approach is exact and a constitutive equation (whether it be linear or nonlinear, passive or active, elastic solid or viscous fluid, viscoelastic, etc.) is not required. By contrast, in two dimensions the situation is more complex because a constitutive law is required. Additional difficulties can be attributed to the effects of the boundary conditions (described below). Tambe *et al.* [11,12] addressed the Poisson ratio issue by computing intercellular stresses with a variety of Poisson ratios. They found the dependence upon the Poisson ratio to be small, thus lending credence to the MSM approach. On the other hand, it has also been suggested that under certain conditions, including a Poisson ratio of zero, the two-dimensional (2D) problem decouples into two one-dimensional (1D) (exact) problems. This is true only in the exceptional case when shear stresses (equivalently, off-diagonal elements in the strain tensor) are zero. Interestingly, the principle of plithotaxis holds that cells migrate along trajectories that minimize intercellular shear stresses [11,12], thus making such an assumption a plausible approximation, although this argument in turn rests on the validity of MSM, which we question. In general, however, even when the Poisson ratio is zero, the stresses depend on both x and y tractions, and the equations do not decouple into a pair of exact 1D problems.

Nevertheless, MSM remains to be validated in the following sense: the tractions are the independent variable, along with boundary conditions at the edge of the monolayer or field of view (see Sec. IIE below), from which $\sigma_{\text{MSM}}(T)$ is calculated. But we currently have no direct measures of intercellular stress by which to compare these. Hence, currently MSM relies on testing the self-consistency of MSM within the MSM framework (Tambe *et al.* [11,12] and the above-mentioned works). An equivalent test validating the use of a passive constitutive law is to note that a measured local velocity or displacement field, together with an assumed viscosity or shear modulus, suffices to compute the tractions. And these in turn can then be compared with the experimentally measured tractions using traction force microscopy (TFM) [7–9]. This comparison is the focus of this work. We find that tractions computed from the measured velocity field are uncorrelated with tractions measured by TFM. This result implies a shortcoming in the assumptions that underlie both the MSM and the Stokes approach used here.

This paper is structured as follows. In Sec. II we review MSM used for stress recovery within a monolayer [11]. Since its conception ten years ago [11], many derivative models

have been suggested that use either a Hookean solid [10–12,16–28] or a Newtonian fluid [10,18,20,30–32] constitutive equation. We review both formulations and the embedded assumptions of MSM. In Sec. III, we present our alternative approach and show that MSM is an incomplete theory. Section IV discusses the implications of our result, which is that the constitutive equation needs modification. One candidate is inclusion of an additive active stress field. A variety of such active stress formulations have been put forward on an *ad hoc* basis [10,17,19–21,26–33]. Under this restrictive assumption of additivity, we derive a relationship that links active force density to substrate tractions and velocity fields. It does not cover issues such as nonlinearity, plasticity, and other rheological features. Nevertheless, to the extent that one posits such an additive contribution of active forces, this relationship is general and robust. Finally, we conclude in Sec. V with a few short remarks.

II. MONOLAYER STRESS MICROSCOPY

In this section we review MSM as suggested by us and its implementation by others with various derivative models. Section IIA presents the general three-dimensional (3D) governing equations. Section IIB discusses constitutive equations, while Sec. IIC discusses incompressibility. In Sec. IID, we reduce the governing equation to two dimensions (2D), while Sec. IIE discusses the various boundary conditions (BCs) needed at the edge of the monolayer.

A. Three-dimensional model

In a 3D bulk where inertia is neglected and there are no body forces [Fig. 1(a)], Newton's second law is

$$\nabla_{3D} \cdot \sigma_{3D} = 0, \quad (1)$$

where σ_{3D} are the 3D stresses. In 3D, the 2D tractions $\mathbf{T}$ are accounted for as BCs [Fig. 1(b)] while BCs at the edge of the monolayer are discussed in Sec. IIE. In the remainder of this work, we refer to $\mathbf{T}$ as those measured by TFM [8,9] (i.e., the forces the cells apply on the substrate). Equation (1) is an indeterminate vector of three equations comprising six different terms (σ_{ij} , $i, j = x, y, z$). Hence, solution of this equation requires additional equations in the form of either a constitutive equation or compatibility equation.

B. Elastic and viscous constitutive laws

The 2D equations comprise three terms (σ_{xx} , σ_{yy} , σ_{xy}). To close these set of equations, the Beltrami-Mitchell compatibility equation was supplemented [11,12]. However, using this equation is tantamount to assuming that the monolayer obeys a simple Hookean constitutive law [13] characterized by two material constants: shear modulus G and Poisson ratio ν . In this work, for the sake of simplicity, we consider the case of $\nu = 1/2$, which is the limiting case of an incompressible elastic material (i.e., volume conservation, see Sec. IIC). For an incompressible elastic medium the constitutive law [13]

$$\sigma = \frac{1}{3} \text{tr}(\sigma) \mathbf{I} + G (\nabla_{3D} \mathbf{u}_{3D} + \nabla_{3D} \mathbf{u}_{3D}^T), \quad (2)$$

where $\mathbf{u}_{3D}$ is the 3D displacement vector field. However, without time-dependent terms in Eq. (1), the equations for an incompressible Hookean solid and incompressible Newtonian fluid are similar, except for a time derivative in the latter case. This is easily seen by inspection of the constitutive equation for an incompressible fluid [14,15],

$$\boldsymbol{\sigma} = \frac{1}{3}\text{tr}(\boldsymbol{\sigma})\mathbf{I} + \mu(\nabla_{3D}\mathbf{v}_{3D} + \nabla_{3D}\mathbf{v}_{3D}^T), \quad (3)$$

where $\mathbf{v}_{3D}$ and μ are respectively the 3D velocity vector field and viscosity. It is immediately clear that Eqs. (2) and (3) are analogous with the identification of displacement with velocity and shear modulus with viscosity. How are these two distinctly different physical models related? From the experimental perspective, what is typically measured is the displacement of cells $\mathbf{u}$ between two consecutive frames, say at $t = 0$ and $t = \Delta t$, yielding the velocity, $\mathbf{v} = \mathbf{u}/\Delta t$. Importantly, if the stress field at $t = 0$ is assumed to be zero, then whether one uses the displacement field, Eq. (2), or the velocity field, Eq. (3), the stresses from both approaches are necessarily the same. This addresses the issue of whether a Hookean description can be valid when cells are clearly moving. The answer is yes insofar as for any pair of images, the assumption of a stress-free state in the first image yields the same stress field as the Newtonian fluid description. Importantly, the fluid description is more general in that does not require an assumption of a zero-stress state in any of the image frames, nor does it require an assumption of small strains.

While these models are similar, here we adopt the viscous formulation as it requires fewer assumptions from the experimental perspective. In what follows we use the more conventional notation for the fluid's constitutive equation,

$$\boldsymbol{\sigma} = -p\mathbf{I} + \mu(\nabla_{3D}\mathbf{v}_{3D} + \nabla_{3D}\mathbf{v}_{3D}^T), \quad (4)$$

whereby the pressure p takes the place of the trace of the stress tensor, $p \triangleq -\frac{1}{3}\text{tr}(\boldsymbol{\sigma})$.

Regardless of the chosen experimental interpretation, since both models are similar and the formulations are similar, they both share the discrepancy discussed in Sec. III.

Inserting any of these constitutive equations [Eqs. (2)–(4)] into Eq. (1) leads to

$$-\nabla p + \mu\nabla^2\mathbf{v}_{3D} = 0. \quad (5)$$

C. Incompressibility

Cells are comprised mainly of water and as such are essentially incompressible. In either the Hookean solid formulation [13] or Newtonian fluid formulation [14,15], an incompressible material must also satisfy volume conservation. In 3D, this reads as

$$\nabla_{3D} \cdot \mathbf{v}_{3D} = 0. \quad (6)$$

Nevertheless, cells are not strictly isovolumic insofar as water can be transported into or out of the cell via osmotic stress or water channels. Intercellular fluid flow within a monolayer

occurs over timescales of hours [34–36]. These water fluxes comprise a source term in the continuity equation [Eq. (6)]. Indeed, it has been shown that osmotic pressure results in cell area and volume oscillations with a period of 4 h and amplitude of 20% [34,35]. Further, proliferation and apoptosis, and cell extrusion from the layer represent additional source terms [26,37]. Because these phenomena are slow ($\sim$ hours) compared with the timescales of interest here ($\sim$ minutes), we take the monolayer to be isovolumic, as have others [10,19,20,24,38–42].

D. Reduction from three to two dimensions

In the case of plane stress, the 3D equations for either elastic bodies or fluids can be reduced to a thin fluid film in 2D [13]. The reduction is based on the assumption that the height of the monolayer h is much smaller than characteristic in-plane lengths and is taken to be uniform [9,10,18].

Here we give a brief derivation in the incompressible case (see Chap. 67 of Sokolnikov [13]). In plane stress, the shear stresses in the xz and yz planes are zero,

$$\sigma_{xz} = \sigma_{yz} = 0, \quad (7)$$

and the z component of the normal stress is zero ($\sigma_{zz} = 0$). The constitutive law [Eq. (4)] yields

$$\sigma_{zz} = -p + 2\mu v_{z,z} = 0 \Rightarrow v_{z,z} = p/(2\mu). \quad (8)$$

The comma subscript denotes partial differentiation. From 3D incompressibility [Eq. (6)] we have

$$\nabla_{3D} \cdot \mathbf{v}_{3D} = 0 \Rightarrow v_{z,z} = -v_{x,x} - v_{y,y} = -\nabla_2 \cdot \mathbf{v}_{2D}, \quad (9)$$

where $\nabla_2 \equiv (\partial_x, \partial_y)$ is a 2D operator and $\mathbf{v}_{2D} = (v_x, v_y)^T$. Equating Eqs. (8) and (9) yields the 2D incompressibility condition:

$$\nabla_2 \cdot \mathbf{v}_{2D} = -p/(2\mu). \quad (10)$$

In contrast to a 3D formulation, where the tractions are imposed as boundary conditions, in the plane stress reduction the tractions assume the role of a body force [10–12,16–24,30]. It is common to write the 2D governing equation of force balance as

$$\nabla_{2D} \cdot \boldsymbol{\sigma}_{2D} = \mathbf{T}/h. \quad (11)$$

However, it is beneficial to use the constitutive equation [Eq. (4)]. Then, this reads as

$$\sigma_{ij,j} = -p_{,i} + \mu(v_{i,jj} + v_{j,ji}) = T_i/h. \quad (12)$$

Using Eq. (10), Eq. (12) becomes

$$\sigma_{ij,j} = -\frac{3}{2}p_{,i} + \mu v_{i,jj} = -T_i/h, \quad (13)$$

or, alternatively, in vector notation,

$$-\frac{3}{2}\nabla_{2D}^2 p + \mu \nabla_{2D}^2 \mathbf{v}_{2D} = \mathbf{T}/h. \quad (14)$$

Typically, MSM is presented by Eq. (11) along with the incompressible Beltrami-Mitchell equation:

$$\nabla_{2D}^2(\sigma_{xx} + \sigma_{yy}) = \frac{3}{2}(\nabla \cdot \mathbf{T})/h. \quad (15)$$

We call these two equations [Eqs. (11) and (15)] the classical MSM formulation, in which the tractions are the independent variable and the stresses are the dependent variable. By contrast, in Eqs. (10) and (14), the tractions measured by TFM are again the independent variable, but now the velocities are the dependent variable, from which the stresses can be calculated. We call these two equations the Stokes formulation. If all the BCs at the edge of the monolayer are given in terms of the stresses, then Eqs. (11) and (15) can be solved. However, as we discuss in the next section, typically the BCs are given in term of the velocities, and as such, Eqs. (11) and (15) cannot be solved without introduction of an intermediate variable of displacement or velocity through the constitutive law. By contrast, Eqs. (10) and (14) can be solved directly and locally for tractions with velocities taken as the independent variable without the need for specific BC assumptions. This will be used below in Sec. III, where the tractions obtained from Eqs. (10) and (14) are used to directly compare with experimentally measured tractions. In the remainder of this work, we consider the 2D MSM problem; for the sake of brevity, we drop the 2D subscript.

E. Edge boundary conditions

Originally, MSM [11,12] estimated intercellular stresses directly from the traction field. Solution of the governing equations requires assumptions about the boundary conditions at the free edges of the cellular domain or at the boundary of the field of view. This latter boundary was termed the “optical edge” [12]. In what follows we will refer to the conditions at both the free and optical edges as “edge conditions” to distinguish these from the 3D BCs at the top and bottom surfaces, discussed in Fig. 1.

Here we revisit the question of what are the most appropriate edge conditions necessary for a computation of intercellular stresses. We begin with a short review of our previous discussion [12]. Figure 2 shows a schematic representation of an island of cells expanding into free space where four scenarios are considered. The island’s boundary is given by a solid black line. The red circle represents an obstacle. Each square represents a possible field of view and is color coded for four possible cases. In all cases, we assume velocities (or displacements) and tractions are accessible through measurement.

Case 1 (dashed purple line).—The entire island is imaged. In this situation, the edge conditions at the free edges are taken to be stress free:

$$\boldsymbol{\sigma} \cdot \mathbf{n} = 0, \quad \boldsymbol{\sigma} \cdot \mathbf{t} = 0. \quad (16)$$

Case 2 (dashed-dotted green line).—Only part of the island is imaged. For the free edges, the edge conditions are given by Eq. (16), as in case 1. However, for the optical edges (at the boundaries of the field of view) an assumption must be made. Previously [12], we argued that at the “optical” boundaries the condition is zero normal flux and zero shear stress:

$$\mathbf{v} \cdot \mathbf{n} = 0, \quad \boldsymbol{\sigma} \cdot \mathbf{t} = 0. \quad (17)$$

Here we suggest that instead of this *ad hoc* assumption it be replaced by using additional measurements of velocities or displacements at the optical edge (e.g., with PIV) such that at the optical edge,

$$\mathbf{v} = \mathbf{v}_{\text{meas}}. \quad (18)$$

Other approaches are possible, including requiring zero tractions at the cell free boundary [23,24], with an apparent stress jump at the same boundary, the origins of which remain open.

Case 3 (dotted blue line).—This is solely determined by optical edges, for which Eq. (18) is taken to be the appropriate edge condition.

Case 4 (long-dashed dotted red line in Fig. 2).—Here cells have contacted an obstacle and we must make a distinction between two kinds of obstacles, each of which has two limiting subtypes. The first is a physical obstacle. In this case, the first subtype is no-slip and no flux, for which, at the interface,

$$\mathbf{v} = 0. \quad (19)$$

The second subtype is a pure slip (i.e., zero shear stress) with no flux, for which, again at the interface, the edge condition is

$$\boldsymbol{\sigma} \cdot \mathbf{T} = 0, \quad \mathbf{v} \cdot \mathbf{n} = 0. \quad (20)$$

The other kind of obstacle is chemical, for example, the boundary between a region coated with collagen and region without. Here, the first subtype is where there are no cellular interactions outside the observed boundary of the island, for which the edge condition is stress-free [Eq. (16)]. The second subtype is where there may be cellular interactions and nonzero tractions outside the visible island. For example, the presence of protruding lamellipodia may exert tractions but may not be visible on a phase image. In this case, it is most appropriate to use zero normal velocity and zero stress [Eq. (20)]. Which condition is most appropriate depends on the nature of the obstacle and demands attention to the biology of cell/extracellular-matrix interactions.

Finally, we note the approach of two recent works [31,32]. These works differ from this one, whose starting point is a purely passive constitutive equation, in that their initial assumption was that the stress tensor included an active term [similar to what will be discussed in Sec. IV]. The inclusion of this stress term added an additional free parameter that could be determined by applying a tangential and two normal BCs at the free edge,

$$\boldsymbol{\sigma} \cdot \mathbf{T} = 0, \quad \boldsymbol{\sigma} \cdot \mathbf{n} = 0, \quad \mathbf{v} \cdot \mathbf{n} = 0. \quad (21)$$

It should be noted that this approach worked in their quasi-1D geometry of a circular island. However, it remains to be seen if a single fitting parameter is suitable for arbitrary 2D geometries.

III. MSM VALIDATION

This section is divided as follows. Section IIIA discusses the inherent difficulties in validating MSM. Section IIIB presents our approach, which bypasses all these difficulties and allows for validation. Section IIIC gives a short review of the experimental results used in this work. In Sec. IIID we compare theory and experiments, and show a striking discrepancy. The implications of this discrepancy are given in Sec. IIIE.

A. Difficulties in validating MSM

A direct validation of MSM would require an independent experimental measurement of stresses, which does not presently exist. In the Stokes formulation, an independent measurement of stress is no longer required, because the stresses follow directly from Eq. (4). Rather, the validation requires that both the tractions $\mathbf{T}$ and velocities $\mathbf{v}$ (denoted by $\mathbf{T}^{\text{meas}}$ and $\mathbf{v}^{\text{meas}}$, respectively) be simultaneously measured. Fortunately, both are easily measured: $\mathbf{T}^{\text{meas}}$ by TFM [7–9] and $\mathbf{v}^{\text{meas}}$ by PIV, optical flow, or other experimental methods.

For the Stokes approach, one end goal would be an analytical solution for the velocity given a traction field, $\mathbf{v}^{\text{calc}}(\mathbf{T})$, in 2D or 3D, and comparing this with $\mathbf{v}^{\text{meas}}$. However, this is not without difficulties. First, the governing equations [Eqs. (10) and (14)] are a set of coupled linear partial differential equations. Second, these equations are subject to varying types of BCs (see Sec. IIE). Third, unless the geometry is extremely simple, an analytical solution is typically not possible, thus necessitating numerical evaluation.

Even if we could calculate $\mathbf{v}^{\text{calc}}(\mathbf{T})$ and compare it to $\mathbf{v}^{\text{meas}}$, any variation between theory and experiment could be attributed to an incorrect choice of appropriate BCs for those images that are larger than the field of view. This also holds for the 2D comparison of stresses. To bypass this BC dependency, here we adopt an alternative approach that negates the need for solving for $\mathbf{v}(\mathbf{T})$ [or $\boldsymbol{\sigma}(\mathbf{T})$]. This is described in the next section. However, we did find two $\mathbf{v}(\mathbf{T})$ solutions: Appendix A provides a derivation of a 3D solution, $\mathbf{v}_{3D}(\mathbf{T})$, for a monolayer where we assume that the monolayer is infinite in the x, y plane and solves the governing equation via a Fourier transform. However, due to the complexity of the expressions in $\mathbf{k}$ space, the final calculation requires numerical evaluation. Appendix B provides a derivation of the simpler 2D plane-stress problem, which is typically the most

relevant for MSM. We calculated these inverse functions and give an exact solution for $v_{2D}(T)$.

B. Alternative approach to MSM

Once more, the governing equations in 2D are

$$\nabla \cdot \mathbf{v} = -p/(2\mu), \quad (22)$$

$$-\frac{3}{2}\nabla p + \mu\nabla^2 \mathbf{v} = \mathbf{T}/h. \quad (23)$$

Inserting Eq. (22) into Eq. (23) leads to [13]

$$\nabla^2 \mathbf{v} + 3\nabla(\nabla \cdot \mathbf{v}) = \mathbf{T}/(\mu h) \equiv \mathbf{t}. \quad (24)$$

In classical MSM [11,12,16–25,30], $\sigma_{\text{MSM}}(\mathbf{T})$ is calculated. In the Stokes formulation, Eq. (24) is solved for $\mathbf{v}(\mathbf{T})$ [and then $\sigma_{\text{Stokes}}(\mathbf{T})$ is calculated via Eq. (4)]. As previously noted, solving for $\mathbf{v}(\mathbf{T})$ is nontrivial.

These difficulties can be entirely circumvented. As $\mathbf{T}$ and $\mathbf{v}$ are both measured, our alternative approach is to solve for $\mathbf{T}$ given $\mathbf{v}$, denoted $\mathbf{T}^{\text{calc}}(\mathbf{v})$. Here we compute calculated tractions, normalized by μh , denoted $\mathbf{t}^{\text{calc}} = \mathbf{T}^{\text{calc}}(\mathbf{v}^{\text{meas}})/\mu h$, which depends only on the derivatives of the velocity [Eq. (24)]. In contrast to $\mathbf{v}(\mathbf{T})$, which is difficult to calculate and BC dependent, calculating $\mathbf{t}^{\text{calc}}(\mathbf{v})$ is simple; it is algebraic, and in-plane edge conditions are not needed. Also, $\sigma_{\text{Stokes}}(\mathbf{v})$ stresses can be calculated directly (up to a factor of μh) from the velocities given in Eqs. (4) and (22).

We compared $\mathbf{t}^{\text{calc}}$ with $\mathbf{T}^{\text{meas}}$ based upon measurements from our previous work, wherein $\mathbf{v}^{\text{meas}}$ and $\mathbf{T}^{\text{meas}}$ were obtained independently and concurrently in the same experiment [16]. There are two extreme cases in this comparison of $\mathbf{t}^{\text{calc}}$ with $\mathbf{T}^{\text{meas}}$. First, $\mathbf{t}^{\text{calc}}$ and $\mathbf{T}^{\text{meas}}$ are proportional. This would imply that MSM is self-consistent and accurate. Second, $\mathbf{t}^{\text{calc}}$ and $\mathbf{T}^{\text{meas}}$ are unrelated. This would imply that MSM is not self-consistent. Being incomplete, the intercellular stresses computed by MSM must be taken with caution. Here we show that the latter is true. We use the Pearson correlation test, which is independent of μh , and find that $\mathbf{t}^{\text{calc}}$ and $\mathbf{T}^{\text{meas}}$ are essentially completely uncorrelated.

C. Experimental setup

Briefly, cells were cultured until they were confluent in a confined region bounded by a barrier [16]. Subsequently, the barrier was lifted so that the monolayer could migrate into a cell-free space containing a circular obstacle into which the cells cannot migrate. The velocity field was measured by particle image velocimetry (PIV) [9], and the traction field was measured by TFM [8,9]. To smooth the spontaneous spatial fluctuations that are known to characterize collective cellular migration, velocity and traction fields were averaged over six identical experiments. To enhance the ensemble averaging, each field was folded top half to bottom half averaged over (see methods of Ref. [16] for details regarding ensemble averaging). These fields were smoothed further using a Gaussian filter with a two-pixel

standard deviation [44,45]. To ensure that the lack of correlation is not due to the ensemble average or the effect of the interaction with the obstacle, we also compared the measured and calculated tractions from a single (nonaveraged) dataset for pre- and post-interaction with the obstacle [43]; $\mathbf{t}^{\text{calc}}$ and $\mathbf{T}^{\text{meas}}$ for these datasets were also uncorrelated.

The advancing monolayer encountered and then encompassed the circular obstacle [Fig. 3(a)] [16]. The corresponding velocity components (v_x , v_y) around the circular obstacle resemble those of flow around a cylinder [14], in which case the flow field divides at an upstream stagnation point [Figs. 3(b) and 3(c)].

D. Comparison of $\mathbf{T}^{\text{meas}}$ and $\mu h \mathbf{t}^{\text{calc}}$

After applying the same Gaussian smoothing filter [44,45] to the measured x component of traction T_x^{meas} , the heterogeneity and punctate nature of this field became apparent [Fig. 4(a)]. The corresponding x component of traction, $\mu h t_x^{\text{calc}}$, is also heterogeneous but more highly punctate due to its origin in the derivatives of the velocity field [Fig. 4(b)]. The values chosen for the height and viscosity are discussed below (see Sec. IVA and Appendix C) and are used for illustrative purposes so that $\mu h t_x^{\text{calc}}$ and T_x^{meas} can be described by the same heat map and have the same units.

A pixel-by-pixel scatter plot of these two traction fields reveals that the peak of T_x^{meas} is centered around 4 Pa while that of $\mu h t_x^{\text{calc}}$ is centered at around zero (Fig. 5). This difference can also be seen in Figs. 4(d) and 4(e), which show the heat maps of $|\mathbf{T}^{\text{meas}}|$ and $|\mu h \mathbf{t}^{\text{calc}}|$, respectively, as well as their vector fields. It is evident that $\mathbf{T}^{\text{meas}}$ [Fig. 4(d)] appears to be dominated by the x component, T_x^{meas} , and is pulling away from the free edge [9,16]; $\mu h \mathbf{t}^{\text{calc}}$ [Fig. 4(e)] appears to lack a distinct direction and is diffuse. Equation (24) provides a relation between $\mathbf{t}^{\text{calc}}$ and $\mathbf{v}^{\text{meas}}$ that is independent of μh . Had the calculated tractions, $\mathbf{t}_x^{\text{calc}}$, been proportional to the measured tractions, T_x^{meas} , then the resultant fields would look similar up to a multiplicative factor of μh . This is clearly not the case.

To quantify the degree of correlation, we computed the Pearson correlation coefficient between T_x^{meas} and t_x^{calc} , given by $\rho_x = \text{cov}(T_x^{\text{meas}}, t_x^{\text{calc}}) / [\text{std}(T_x^{\text{meas}}) \text{std}(t_x^{\text{calc}})]$. This is the covariance of the measured and calculated tractions divided by their respective standard deviations and is therefore dimensionless and independent of μh . The calculated correlation coefficients are $\rho_x = -0.022$ and $\rho_y = -0.012$, indicating little correlation (the y -component data are given in the Supplemental Material [43]).

E. Implications for MSM

As noted above in the Introduction, the foundations of 2D $\sigma_{\text{MSM}}(\mathbf{T})$ have not been verified experimentally because there are no independent measurements of internal stress, i.e., to the best of our knowledge there is no current method for measuring all three components of the stress tensor. On the other hand, any constitutive law, including MSM as a subset, can be tested for internal consistency by a specific comparison of $\mathbf{T}^{\text{meas}}$ and $\mathbf{T}^{\text{calc}}(\mathbf{v}^{\text{meas}})$. Here we have done that. Our finding of a lack of correlation between $\mathbf{T}^{\text{meas}}$ and $\mathbf{t}^{\text{calc}}(\mathbf{v}^{\text{meas}})$ necessarily implies $\sigma_{\text{Stokes}}[\mathbf{T}^{\text{calc}}(\mathbf{v}^{\text{meas}})] \neq \sigma_{\text{MSM}}(\mathbf{T}^{\text{meas}})$. In words, intercellular stresses computed from measured tractions are not the same as intercellular stresses computed from

the measured velocity field. The next section (Sec. IV) discusses the implications of this result, namely, that the constitutive equation requires modification both in MSM as well as in the Stokes approach. One of the suggested modifications, discussed below, is the inclusion of active stresses. By contrast to the 2D situation, stress recovery using MSM in 1D is straightforward and unambiguous, and a constitutive law is not required. For example, a simple balance of forces requires that for stresses in the x direction, $\sigma_{xx} = h^{-1} \int T_x dx$. This approach has been successfully used to calculate 1D stresses [9,17,19,20,26–28,31,32] in cases where the symmetry of the geometry (planar or circular) allows, through averaging, a reduction to 1D.

More complex geometries require solutions in 2D and the need for compatibility equations arising from a constitutive law. For example, Zimmerman *et al.* [21] recently suggested that perhaps stress recovery was method independent. In that work [21], they compared 2D stresses recovered from 2D tractions using MSM versus the stresses recovered using the molecular dynamic simulation method of Hardy [46]. Heat maps of recovered stresses in that report [21] show close agreement between the two methods at longer length scales, supporting their claim that stress recovery is method independent, but at shorter length scales indicate discrepancies that approach 400%. At this scale intercellular stress recovery is strongly method dependent, and as such, the issue of intercellular stress recovery remains open.

Assessing errors in MSM and the Stoke stresses is currently not possible. This can be attributed to a number of uncertainties that still require further investigation. Namely, independent measurements of the viscosity, the stresses, or the constitutive equation are needed. This is left for future work.

IV. GOING BEYOND MSM AND STOKES

The main finding of this work is that tractions calculated from the measured velocity field, $T^{\text{calc}}(\mathbf{v}^{\text{meas}})$, do not correspond to tractions that are measured directly, T^{meas} . Hence, the underlying assumptions of both MSM and the equivalent Stokes approach require reevaluation. In particular, the constitutive equations must be modified. There are three natural candidates: a linear viscoelastic rheology, a nonlinear rheology (visco-plasto-elastic, glassy, or otherwise), or an active rheology.

As pointed out previously, classical MSM and the fluid Stokes approach here both assume that the constitutive law is that of a linear elastic material or a linear viscous material. However, as correctly pointed out to us by one of the referees, a possible resolution is linear and/or nonlinear visco-plasto-elastic approaches, or even the soft glassy rheology [47,48]. Linear viscoelastic approaches have been adopted in various biological systems (cell aggregate under aspiration [40], cell aggregate spreading [49], and single-cell twisting via magnetic tweezers [47,50]). To the best of our knowledge, such modeling has not been attempted on monolayers. We have yet to attempt to address the issue of viscoelasticity as it goes beyond our main question: Is a simple (viscous or elastic) linear passive formulation of MSM consistent? Future work on monolayer dynamics should consider both linear and nonlinear rheologies.

Finally, we consider the last of these candidates—active rheology. We do this because currently this is the most favored approach in investigating monolayer dynamics [10,17,19–21,26–33] and there is some experimental evidence supporting this approach. Below, we show a relation for the behavior of active stresses that can be derived from Newton's law and requires minimal assumptions.

A. Active stresses as a resolution

Previous works [10,17,19–21,26–33] considered the possibility that the stress tensor additively comprises both a passive σ^p and active σ^a term, such that

$$\sigma = \sigma^p + \sigma^a. \quad (25)$$

It is then assumed that [10,17,19–21,26–28,30–32] the passive term is either an elastic solid [Eq. (2)] or viscous fluid [Eq. (4)]. To the extent that the velocities are representative of the motion of the composite monolayer, we insert $\sigma^p = \sigma^{\text{viscous}}$ and Eq. (25) into Newton's law [Eq. (11)]:

$$\nabla \cdot (\sigma^a + \sigma^p) = T/h. \quad (26)$$

The active force density, denoted F^{active} , is defined as

$$F^{\text{active}} = \nabla \cdot \sigma^a. \quad (27)$$

From Eqs. (4), (26), and (27) we have F^{active} explicitly in terms of measured quantities:

$$\begin{aligned} hF^{\text{active}} &= T^{\text{meas}} - \mu h t^{\text{calc}} \\ &= T^{\text{meas}} - \mu h \left[\nabla^2 v^{\text{meas}} + 3 \nabla (\nabla \cdot v^{\text{meas}}) \right]. \end{aligned} \quad (28)$$

F^{active} includes not only the tractions and velocities but also the height h and the viscosity μ . It follows from Eq. (28) that in the limit of low viscosity F^{active} is dominated by the tractions T^{meas} , whereas in the limit of high viscosity F^{active} is dominated by passive component $\mu h t^{\text{meas}}$.

Note that while h may be estimated accurately [9,10,18], the same is not true for μ . Appendix C shows a number of estimates using different methods [43]. Here we illustrate F_x^{active} for the case where $h = 5 \mu\text{m}$ and $\mu = 23 \text{ kPa s}$, the latter being our best estimate resting on the fewest assumptions, and also falling in an intermediate range such that the contributions of both T^{meas} and $\mu h t^{\text{meas}}$ can both be seen [Fig. 4(c)]. For these values, and for the data set described above, we used Eq. (28) to compute hF_x^{active} [Fig. 4(c)], which shows clear contributions from both the traction term [Fig. 4(a)] and the passive velocity terms [Fig. 4(b)]. For example, it can be observed that hF_x^{active} is similar to T_x^{meas} in that the tractions at the free edge are largely pointing away from the obstacle [Fig. 4(a)], as was previously shown [9]. Similarly, there is fine scale structure in hF_x^{active} which reflects its origin in the passive component [Fig. 4(b)]. Figure 4(f) is a vector map showing the directions F^{active} where the length of the arrows are proportional to $|F^{\text{active}}|$. Note that the

active forces are largely pointing away from the obstacle, similar to the tractions, as was shown in [9,16].

B. Origin of active stresses

Recent experiments have confirmed the existence of active forces in epithelial monolayers. For example, in Ref. [26] it was found that 1D stresses are linearly associated with strains but with a nonzero offset, expressed as $\sigma = Ee + \sigma^a$. This is interpreted as reflecting an active component in both the modulus and the offset [26] and supports the active rheology suggested by Eq. (25). The main finding in Ref. [26] is that $\sigma = Ee + \sigma^a$ was a global statement relating stresses and strains as the tractions were averaged both in the perpendicular direction as well as in the center of their expanding island experiment. This approach is perhaps instructive as it suggests that the additive stress relation of Eq. (25) holds. However, this approach cannot be generalized to 2D nor does it account for local changes as does our model. Two recent works also used Eq. (25) in their derivations to explain wetting [32] and fingering instabilities [31] observed in epithelial monolayers. However, they did not consider the question of whether or not the passive formulation is incomplete. In both works of Treppe and co-workers [26,32], blebbistatin was used to inhibit the activity motors; however, even then σ^a was not zero. As discussed below, this too implies that the origin of active stresses is still not understood. Whether Eq. (25) holds is a topic for future work.

With respect to the origin of active forces, velocities, and tractions, several previous works have suggested possible phenomenological forms for the active stress [10]. These potential origins of $\mathbf{F}^{\text{active}}$ differ substantially in the literature. Some works suggest the origin is in the concentration of active contractile units [19,20], while others suggest in-plane cell polarization [19,20,29–32,51], nonzero tension [26], and cell division [26,37]. Importantly, without a compatibility equation for σ^a , integrating Eq. (28) to calculate σ^a , and thus the total stress σ , is not possible. The current state of knowledge regarding $\mathbf{F}^{\text{active}}$ is inconclusive as to which of the above models is most appropriate [10]. Nevertheless, Eq. (28) describing the active stress is derived from Newton's laws, with minimal assumptions, and must be satisfied.

V. CONCLUSIONS

We have shown that based on a linear passive constitutive law (either elastic or viscous), tractions computed from the measured velocity field are uncorrelated with measured tractions. This implies a shortcoming in the rheological assumptions that underlie both the MSM and the Stokes approach. As such, rheology of the cell layer requires further evaluation.

We suggest that intercellular active stresses may resolve any inconsistency but that if the active stresses are not derivable from a constitutive law, then it may not be possible to explicitly compute intercellular stresses. However, we do give an explicit expression [Eq. (28)] for the active force density as a function of the measured tractions and velocities for the restrictive case when the active stresses are additive to a linear passive constitutive law.

In this case, regardless of the proposed origin of the active stresses, this condition, derived from Newton's second law, must be satisfied.

ACKNOWLEDGMENTS

We thank Dr. Jae Hun Kim for generously providing us with the raw experimental data used in Ref. [16] and used in this work. We also thank Dr. Bo Lan for helpful discussions. This work was supported by National Institutes of Health (NIH) Grants No. U01CA202123, No. P01HL120839, No. R01HL148152, and No. T32HL007118. Finally, we thank both reviewers for their helpful and constructive comments throughout.

APPENDIX A:: THREE-DIMENSIONAL SOLUTION FOR $\mathbf{v}(\mathbf{T})$

1. Governing equations in real space

In 3D, the governing Stokes equations for an incompressible fluid of uniform density are

$$\nabla \cdot \mathbf{v} = 0, \quad (\text{A1})$$

$$-\nabla p + \mu \nabla^2 \mathbf{v} = 0, \quad (\text{A2})$$

where $\mathbf{v} = (v_x, v_y, v_z)^T$ is the 3D velocity vector in Cartesian coordinates and p is the pressure [Fig. 1(a)]. Equations (A1) and (A2) express conservation of mass and force balance, respectively.

For boundary conditions, we take the top (apical) surface, $z = h$, to be stress free ($\boldsymbol{\sigma} \cdot \mathbf{n} = 0$) [Fig. 1(b)]:

$$\mu(v_{x,z} + v_{z,x})|_{z=h} = 0, \quad (\text{A3})$$

$$\mu(v_{y,z} + v_{z,y})|_{z=h} = 0, \quad (\text{A4})$$

$$(2\mu v_{z,z} - p)|_{z=h} = 0. \quad (\text{A5})$$

The comma subscript denotes partial differentiation. Note that we assume the film height to change only slowly ($\nabla h \ll 1$) such that curvatures terms are negligible [52–54]. At the bottom (basal) surface that is in contact with the substrate, $z = 0$, the shear stress must be balanced by the tractions, and we take zero normal flux:

$$v_{x,z}|_{z=0} = T_x/\mu, \quad (\text{A6})$$

$$v_{y,z}|_{z=0} = T_y/\mu, \quad (\text{A7})$$

$$v_z|_{z=0} = 0. \quad (\text{A8})$$

2. Governing equations in k space

The form of Eqs. (A1)–(A8) suggests using two-dimensional Fourier transforms, defined for any function $f(\mathbf{r})$ by $\tilde{f}(\mathbf{k}) = \int d^2r f(\mathbf{r}) e^{-i\mathbf{k} \cdot \mathbf{r}}$, integrated over the plane. Here $\mathbf{r} = (x, y)^T$, the wave vector $\mathbf{k} = (\alpha, \beta)^T$, $k = |\mathbf{k}|$, $r = |\mathbf{r}|$, and $i = \sqrt{-1}$ is the imaginary unit. With this notation, Eqs. (A1)–(A8) transform to

$$i\alpha\tilde{v}_x + i\beta\tilde{v}_y + \tilde{v}_{z,z} = 0, \quad (\text{A9})$$

$$-i\alpha\tilde{p} + \mu(\tilde{v}_{x,zz} - k^2\tilde{v}_x) = 0, \quad (\text{A10})$$

$$-i\beta\tilde{p} + \mu(\tilde{v}_{y,zz} - k^2\tilde{v}_y) = 0, \quad (\text{A11})$$

$$-\tilde{p}_{,z} + \mu(\tilde{v}_{z,zz} - k^2\tilde{v}_z) = 0. \quad (\text{A12})$$

Note that this approach to solving the field equations is exactly the same as that used in Ref. [9], albeit with different top and bottom boundary conditions. In the current application, the transformed stress-free boundary conditions are given by, for the top surface,

$$(\tilde{v}_{x,z} + i\alpha\tilde{v}_z)|_{z=h} = 0, \quad (\text{A13})$$

$$(\tilde{v}_{y,z} + i\beta\tilde{v}_z)|_{z=h} = 0, \quad (\text{A14})$$

$$(2\mu\tilde{v}_{z,z} - \tilde{p})|_{z=h} = 0, \quad (\text{A15})$$

and for the tractions and homogeneous flux at the bottom,

$$\tilde{v}_{x,z}|_{z=0} = \tilde{T}_x/\mu, \quad (\text{A16})$$

$$\tilde{v}_{y,z}|_{z=0} = \tilde{T}_y/\mu, \quad (\text{A17})$$

$$\tilde{v}_z|_{z=0} = 0. \quad (\text{A18})$$

3. Three-dimensional solution in k space

There are six solutions to Eqs. (A9)–(A12), given in Ref. [9], the two linear combinations of which satisfy the four homogeneous conditions [Eqs. (A13)–(A15) and (A18)] and can be written in vector notation:

$$\boldsymbol{\varphi}_1(z) = \frac{\cosh[k(h-z)]}{\cosh kh} \begin{bmatrix} \beta \\ -\alpha \\ 0 \end{bmatrix}, \quad (\text{A19})$$

$$\begin{aligned} \boldsymbol{\varphi}_2(z) = & -\frac{1+2k^2h^2+\cosh 2hk}{\phi} \begin{bmatrix} \alpha \cosh kz \\ \beta \cosh kz \\ -ik \sinh kz \end{bmatrix} + \begin{bmatrix} \alpha \sinh kz \\ \beta \sinh kz \\ -ik \cosh kz \end{bmatrix} \\ & + kz \begin{bmatrix} \alpha \cosh kz \\ \beta \cosh kz \\ -ik \sinh kz \end{bmatrix} - \frac{2(\cosh hk)^2}{\phi} kz \begin{bmatrix} \alpha \sinh kz \\ \beta \sinh kz \\ -ik \cosh kz \end{bmatrix} \\ & + \begin{bmatrix} 0 \\ 0 \\ ik(\cosh kz - 2\phi^{-1}(\cosh hk)^2 \sinh kz) \end{bmatrix}, \end{aligned} \quad (\text{A20})$$

where $\phi = 2hk + \sinh(2hk)$. The corresponding solutions for the pressure are

$$p_1 = 0, \quad p_2 = -2ik^2 \sinh kz + \frac{4ik^2(\cosh hk)^2}{\phi} \cosh kz. \quad (\text{A21})$$

In these terms, we write the linear combination as

$$\boldsymbol{v} = \begin{bmatrix} \tilde{v}_x \\ \tilde{v}_y \\ \tilde{v}_z \end{bmatrix} = A\boldsymbol{\varphi}_1 + B\boldsymbol{\varphi}_2, \quad p = A\mu p_1 + B\mu p_2, \quad (\text{A22})$$

where A, B are chosen to satisfy the inhomogeneous traction conditions [Eqs. (A16) and (A17)]. Explicitly, this requires

$$M \begin{bmatrix} A \\ B \end{bmatrix} = \begin{bmatrix} \tilde{T}_x/\mu \\ \tilde{T}_y/\mu \end{bmatrix}, \quad (\text{A23})$$

where

$$M = k \begin{bmatrix} -\beta \tanh kh & 2\alpha \\ \alpha \tanh kh & 2\beta \end{bmatrix}. \quad (\text{A24})$$

This leads to

$$\begin{bmatrix} A \\ B \end{bmatrix} = M^{-1} \begin{bmatrix} \tilde{T}_x/\mu \\ \tilde{T}_y/\mu \end{bmatrix}. \quad (\text{A25})$$

Specifically,

$$A = \frac{\alpha T_y - \beta T_x}{\mu k^3} \coth kh, B = \frac{\alpha T_x + \beta T_y}{2\mu k^3}. \quad (\text{A26})$$

Note that in experiments with transverse averaging (such that $v_y = 0$) and where the height is spatially uniform, there is a simple volume conservation relation between $v_z(z = h)$ and the height averaged v_x at the two edges, say $x = a$ and $x = b$. Explicitly,

$$h[v_x(x = b) - v_x(x = a)] + \int_a^b v_z(x'; z = h) dx' = 0. \quad (\text{A27})$$

In 2D expanding islands, the equivalent expression of volume conservation is

$$h \oint_C ds \mathbf{n} \cdot \mathbf{v} + \int_D dx dy v_z(x, y; z = h) = 0, \quad (\text{A28})$$

where C and D are the boundary and domain, respectively. If proliferation is present, these expressions give the volume-averaged mean proliferation rate, given measurements of both edge and height velocities.

APPENDIX B:: TWO-DIMENSIONAL SOLUTION FOR $\mathbf{v}(\mathbf{T})$

1. Two-dimensional solution in $\mathbf{k}$ space

The 2D governing equations are

$$\nabla \cdot \mathbf{v} = -p/(2\mu), \quad (\text{B1})$$

$$-\frac{3}{2}\nabla p + \mu \nabla^2 \mathbf{v} = \mathbf{T}/h. \quad (\text{B2})$$

The solution of Eqs. (B1) and (B2) for the case of infinite and periodic edge BCs (similar to Appendix A) yields

$$\tilde{p} = \tilde{\mathbf{G}}_p(\mathbf{k}) \cdot \tilde{\mathbf{T}}, \quad (\text{B3})$$

$$\tilde{\mathbf{G}}_p(\mathbf{k}) = \frac{i\mathbf{k}}{2hk^2}, \quad (\text{B4})$$

$$\mu \tilde{\mathbf{v}} = \frac{3(\mathbf{k} \cdot \tilde{\mathbf{T}})\mathbf{k}}{4hk^4} - \frac{\tilde{\mathbf{T}}}{hk^2}. \quad (\text{B5})$$

With tractions as the only input, inserting Eqs. (B3)–(B5) into the constitutive law [Eq. (4)] confirms the scaling argument in Refs. [11,12] that the stresses are independent of the viscosity (or shear modulus).

2. Two-dimensional solution in real space

We can now calculate Green's function in real space, i.e., the impulse response corresponding to a delta function of tractions at the origin. This is given by the inverse Fourier transform $f(\mathbf{r}) = (4\pi^2)^{-1} \int d^2k \tilde{f}(\mathbf{k}) e^{i\mathbf{k} \cdot \mathbf{r}}$. We start with the pressure [Eq. (B3)],

$$\begin{aligned}
 G_p(\mathbf{r}) &= \frac{1}{4\pi^2} \int_{-\infty}^{\infty} d^2k \tilde{G}_p(\mathbf{k}) e^{i\mathbf{k} \cdot \mathbf{r}} = \frac{1}{4\pi^2} \int_{-\infty}^{\infty} d^2k \frac{i\mathbf{k}}{2hk^2} e^{i\mathbf{k} \cdot \mathbf{r}} \\
 &= \frac{1}{4\pi^2} \int_{-\infty}^{\infty} d^2k \frac{\nabla e^{i\mathbf{k} \cdot \mathbf{r}}}{2hk^2} \\
 &= \frac{1}{8\pi^2 h} \int_0^{\infty} dk \frac{1}{k} \nabla \int_0^{2\pi} d\theta e^{ikr \cos \theta} \\
 &= \frac{1}{8\pi^2 h} \int_0^{\infty} dk \frac{2\pi}{k} \nabla J_0(kr) \\
 &= -\frac{\nabla \mathbf{r}}{4\pi h} \int_0^{\infty} dk J_1(kr) = -\frac{\mathbf{r}}{4\pi h r^2}.
 \end{aligned} \tag{B6}$$

Thus, the pressure is given by the convolution, integrated over all space:

$$p = - \int d^2r' \frac{\tilde{T}(\mathbf{r}') \cdot (\mathbf{r} - \mathbf{r}')}{4\pi h |\mathbf{r} - \mathbf{r}'|^2}. \tag{B7}$$

For the velocity, we identify that we need to calculate three different terms: $\alpha^2 k^{-4}$, $\beta^2 k^{-4}$, $\alpha\beta k^{-4}$, $[\mathbf{k} = (\alpha, \beta)^T]$. The first term is

$$\begin{aligned}
 FT^{-1}\left(\frac{\alpha^2}{k^4}\right) &= \frac{1}{4\pi^2} \int_{-\infty}^{\infty} d^2k \frac{\alpha^2}{k^4} e^{i\mathbf{k} \cdot \mathbf{r}} \\
 &= \frac{1}{4\pi^2} \int_{-\infty}^{\infty} d^2k \frac{-\alpha}{2} \left[\frac{\partial}{\partial \alpha} \left(\frac{1}{k^2} \right) \right] e^{i\mathbf{k} \cdot \mathbf{r}} \\
 &= \frac{i}{8\pi^2} \frac{\partial}{\partial x} \int_{-\infty}^{\infty} d^2k \left[\frac{\partial}{\partial \alpha} \left(\frac{1}{k^2} \right) \right] e^{i\mathbf{k} \cdot \mathbf{r}} \\
 &= -\frac{1}{8\pi^2} i \frac{\partial}{\partial x} \left[(ix) \int_{-\infty}^{\infty} d^2k \frac{1}{k^2} e^{i\mathbf{k} \cdot \mathbf{r}} \right].
 \end{aligned} \tag{B8}$$

To find $\int_{-\infty}^{\infty} d^2k k^{-2} e^{i\mathbf{k} \cdot \mathbf{r}}$, consider the impulse response to the 2D Laplace equation, $\nabla^2 G = \delta^2(\mathbf{r})$, for which the solution is $(2\pi)^{-1} \ln r$. Its 2D Fourier transform is $\tilde{G} = -k^{-2}$. This implies that $\int_{-\infty}^{\infty} d^2k k^{-2} e^{i\mathbf{k} \cdot \mathbf{r}} = -(2\pi)^{-1} \ln r$, from which it is immediate that

$$\begin{aligned}
 FT^{-1}\left(\frac{\alpha^2}{k^4}\right) &= -\frac{1}{8\pi^2} \frac{\partial}{\partial x} \left[(2\pi)^{-1} x \ln r \right] \\
 &= -\frac{1}{16\pi^3} \left(\frac{x^2}{r^2} + \ln r \right),
 \end{aligned} \tag{B9}$$

$$FT^{-1}\left(\frac{\beta^2}{k^4}\right) = -\frac{1}{16\pi^3}\left(\frac{y^2}{r^2} + \ln r\right), \quad (\text{B10})$$

$$FT^{-1}\left(\frac{\alpha\beta}{k^4}\right) = -\frac{1}{16\pi^3}\left(\frac{xy}{r^2} + \ln r\right). \quad (\text{B11})$$

APPENDIX C:: VISCOSITY ESTIMATES

Despite the apparent inconsistencies of using only a passive stress tensor, we use the pure fluid dynamical description to estimate the monolayer viscosity. We employ two different independent methods. First, consider the scaling argument that viscous stresses in a thin film balance the tractions, $\mu \sim Th/u$. For typical values of $T \sim 10$ [Pa], $u \sim 10$ [nm/s], and $h \sim 5$ [μm] [9,11,17], one gets $\mu_{\text{cell}} \sim 5[\text{kPa s}] \sim 5 \times 10^6 \mu_{\text{water}}$ with $\mu_{\text{water}} = 10^{-3}[\text{Pa s}]$. Next, we considered a simple coarse-grained calculation $\mu = \langle T^{\text{meas}} \rangle_{\text{rms}} / \langle |t^{\text{calc}}| h \rangle_{\text{rms}}$, where $\langle \rangle_{\text{rms}}$ is the root-mean-square average; this gives the result $\mu/\mu_{\text{water}} \approx O(2.3 \times 10^7)$. These values together with others from the literature are shown in Table I. Our values are on the lower end of the reported range, but we note that all these estimates are extremely large compared with what we consider physically reasonable. This is another indication that a purely passive viscous description is inadequate and points to the necessity of active stresses to account for the relationship between velocities and tractions.

References

1. Friedl P and Gilmour D, Collective cell migration in morphogenesis, regeneration and cancer, *Nat. Rev. Mol. Cell Biol* 10, 445 (2009). [PubMed: 19546857]
2. Friedl P, Sahai E, Weiss S, and Yamada KM, New dimensions in cell migration, *Nat. Rev. Mol. Cell Biol* 13, 743 (2012). [PubMed: 23072889]
3. Atia L, Bi D, Sharma Y, Mitchel JA, Gweon B, Koehler SA, DeCamp SJ, Lan B, Kim JH, Hirsch R, Pegoraro AF, Lee KH, Starr JR, Weitz DA, Martin AC, Park J-A, Butler JP, and Fredberg JJ, Geometric constraints during epithelial jamming, *Nat. Phys* 14, 613 (2018). [PubMed: 30151030]
4. Park J-A, Kim JH, Bi D, Mitchel JA, Qazvini NT, Tantisira K, Park CY, McGill M, Kim S-H, Gweon B, Notbohm J, Steward R Jr., Burger S, Randell SH, Kho AT, Tambe DT, Hardin C, Shore SA, Israel E, Weitz DA, Tschumperlin DJ, Henske EP, Weiss ST, Manning ML, Butler JP, Drzen JM, and Fredberg JJ, Unjamming and cell shape in the asthmatic airway epithelium, *Nat. Mater* 14, 1040 (2015). [PubMed: 26237129]
5. Bazellères E, Conte V, Elosegui-Artola A, Serra-Picamal X, Bintanel-Morcillo M, Roca-Cusachs P, Muñoz JJ, Sales-Pardo M, Guimerà R, and Trepas X, Control of cell-cell forces and collective cell dynamics by the intercellular adhesome, *Nat. Cell Biol* 17, 409 (2015). [PubMed: 25812522]
6. Roca-Cusachs P, Conte V, and Trepas X, Quantifying forces in cell biology, *Nat. Cell Biol* 19, 742 (2017). [PubMed: 28628082]
7. Dembo M and Wang Y-L, Stresses at the cell-to-substrate interface during locomotion of fibroblasts, *Biophys. J* 76, 2307 (1999). [PubMed: 10096925]
8. Butler JP, Toli -Nørrelykke IM, Fabry B, and Fredberg JJ, Traction fields, moments, and strain energy that cells exert on their surroundings, *Am. J. Physiol. - Cell Physiol* 282, C595 (2002). [PubMed: 11832345]
9. Trepas X, Wasserman MR, Angelini TE, Millet E, Weitz DA, Butler JP, and Fredberg JJ, Physical forces during collective cell migration, *Nat. Phys* 5, 426 (2009).

10. Banerjee S and Marchetti MC, Continuum models of collective cell migration, in *Adv. Exp. Med. Biol* (Springer International Publishing, Cham, Switzerland, 2019), pp. 45–66.
11. Tambe DT, Corey Hardin C, Angelini TE, Rajendran K, Park CY, Serra-Picamal X, Zhou EH, Zaman MH, Butler JP, Weitz DA, Fredberg JJ, and Trepats X, Collective cell guidance by cooperative intercellular forces, *Nat. Mater* 10, 469 (2011). [PubMed: 21602808]
12. Tambe DT, Croult U, Trepats X, Park CY, Kim JH, Millet E, Butler JP, and Fredberg JJ, Monolayer stress microscopy: Limitations, artifacts, and accuracy of recovered intercellular stresses, *PLoS ONE* 8, e55172 (2013). [PubMed: 23468843]
13. Sokolnikov IS, *Mathematical Theory of Elasticity* (Krieger Publishing, Malabar, FL, 1983).
14. Currie IG, *Fundamental Mechanics of Fluids* (McGraw-Hill, New York, 1974).
15. Batchelor GK, *An Introduction to Fluid Dynamics* (Cambridge University Press, Cambridge, England, 2000).
16. Kim JH, Serra-Picamal X, Tambe DT, Zhou EH, Park CY, Sadati M, Park J-A, Krishnan R, Gweon B, Millet E, Butler JP, Trepats X, and Fredberg JJ, Propulsion and navigation within the advancing monolayer sheet, *Nat. Mater* 12, 856 (2013). [PubMed: 23793160]
17. Serra-Picamal X, Conte V, Vincent R, Anon E, Tambe DT, Bazellieres E, Butler JP, Fredberg JJ, and Trepats X, Mechanical waves during tissue expansion, *Nat. Phys* 8, 628 (2012).
18. Kondo Y, Aoki K, and Ishii S, Inverse tissue mechanics of cell monolayer expansion, *PLOS Comput. Biol* 14, e1006029 (2018). [PubMed: 29494578]
19. Banerjee S, Utuje KJC, and Marchetti MC, Propagating Stress Waves During Epithelial Expansion, *Phys. Rev. Lett* 114, 228101 (2015). [PubMed: 26196647]
20. Notbohm J, Banerjee S, Utuje KJC, Gweon B, Jang H, Park Y, Shin J, Butler JP, Fredberg JJ, and Marchetti MC, Cellular Contraction and Polarization Drive Collective Cellular Motion, *Biophys. J* 110, 2729 (2016). [PubMed: 27332131]
21. Zimmermann J, Hayes RL, Basan M, Onuchic JN, Rappel W-J, and Levine H, Intercellular Stress Reconstitution from Traction Force Data, *Biophys. J* 107, 548 (2014). [PubMed: 25099794]
22. Nier V, Jain S, Lim CT, Ishihara S, Ladoux B, and Marcq P, Inference of Internal Stress in a Cell Monolayer, *Biophys. J* 110, 1625 (2016). [PubMed: 27074687]
23. Das T, Safferling K, Rausch S, Grabe N, Boehm H, and Spatz JP, A molecular mechanotransduction pathway regulates collective migration of epithelial cells, *Nat. Cell Biol* 17, 276 (2015). [PubMed: 25706233]
24. Vishwakarma M, Russo JD, Probst D, Schwarz US, Das T, and Spatz JP, Mechanical interactions among followers determine the emergence of leaders in migrating epithelial cell collectives, *Nat. Commun* 9, 3469 (2018). [PubMed: 30150695]
25. Jang H, Notbohm J, Gweon B, Cho Y, Park CY, Kee S-H, Fredberg JJ, Shin JH, and Park Y, Homogenizing cellular tension by hepatocyte growth factor in expanding epithelial monolayer, *Sci. Rep* 7, 45844 (2017).
26. Vincent R, Bazellieres E, Pérez-González C, Uroz M, Serra-Picamal X, and Trepats X, Active Tensile Modulus of an Epithelial Monolayer, *Phys. Rev. Lett* 115, 248103 (2015). [PubMed: 26705659]
27. Uroz M, Wistorf S, Serra-Picamal X, Conte V, Sales-Pardo M, Roca-Cusachs P, Guimerà R, and Trepats X, Regulation of cell cycle progression by cell-cell and cell-matrix forces, *Nat. Cell Biol* 20, 646 (2018). [PubMed: 29802405]
28. Sunyer R, Conte V, Escibano J, Elosegui-Artola A, Labernadie A, Valon L, Navajas D, García-Aznar JM, Muñoz JJ, Roca-Cusachs P, and Trepats X, Collective cell duro-taxis emerges from long-range intercellular force transmission, *Science* 353, 1157 (2016). [PubMed: 27609894]
29. He S, Green Y, Saeidi N, Li X, Fredberg JJ, Ji B, and Pismen LM, A theoretical model of collective cell polarization and alignment, *J. Mech. Phys. Solids* 137, 103860 (2020).
30. Blanch-Mercader C, Vincent R, Bazellieres E, Serra-Picamal X, Trepats X, and Casademunt J, Effective viscosity and dynamics of spreading epithelia: A solvable model, *Soft Matter* 13, 1235 (2017). [PubMed: 28098306]
31. Alert R, Blanch-Mercader C, and Casademunt J, Active Fingering Instability in Tissue Spreading, *Phys. Rev. Lett* 122, 088104 (2019). [PubMed: 30932560]

32. Pérez-González C, Alert R, Blanch-Mercader C, Gómez-González M, Kolodziej T, Bazellieres E, Casademunt J, and Trepát X, Active wetting of epithelial tissues, *Nat. Phys* 15, 79 (2019). [PubMed: 31537984]
33. Alert R and Trepát X, Physical models of collective cell migration, *Annu. Rev. Condens. Matter Phys* 11, 77 (2020).
34. Zehnder SM, Wiatt MK, Uruena JM, Dunn AC, Sawyer WG, and Angelini TE, Multicellular density fluctuations in epithelial monolayers, *Phys. Rev. E* 92, 032729 (2015).
35. Zehnder SM, Suaris M, Bellaire MM, and Angelini TE, Cell volume fluctuations in MDCK monolayers, *Biophys. J* 108, 247 (2015). [PubMed: 25606673]
36. Guo M, Pegoraro AF, Mao A, Zhou EH, Arany PR, Han Y, Burnette DT, Jensen MH, Kasza KE, Moore JR, Mackintosh FC, Fredberg JJ, Mooney DJ, Lippincott-Schwartz J, and Weitz DA, Cell volume change through water efflux impacts cell stiffness and stem cell fate, *Proc. Natl. Acad. Sci* 114, E8618 (2017). [PubMed: 28973866]
37. Ranft J, Basan M, Elgeti J, Joanny J-F, Prost J, and Jülicher F, Fluidization of tissues by cell division and apoptosis, *Proc. Natl. Acad. Sci* 107, 20863 (2010). [PubMed: 21078958]
38. Köpf MH and Pismen LM, A continuum model of epithelial spreading, *Soft Matter* 9, 3727 (2013).
39. Pawlizak S, Fritsch AW, Grosser S, Ahrens D, Thalheim T, Riedel S, Kießling TR, Oswald L, Zink M, Manning ML, and Käs JA, Testing the differential adhesion hypothesis across the epithelial-mesenchymal transition, *New J. Phys* 17, 083049 (2015).
40. Guevorkian K, Colbert M-J, Durth M, Dufour S, and Brochard-Wyart F, Aspiration of Biological Viscoelastic Drops, *Phys. Rev. Lett* 104, 218101 (2010). [PubMed: 20867138]
41. Basan M, Elgeti J, Hannezo E, Rappel W-J, and Levine H, Alignment of cellular motility forces with tissue flow as a mechanism for efficient wound healing, *Proc. Natl. Acad. Sci* 110, 2452 (2013). [PubMed: 23345440]
42. Basan M, Joanny J-F, Prost J, and Risler T, Undulation Instability of Epithelial Tissues, *Phys. Rev. Lett* 106, 158101 (2011). [PubMed: 21568616]
43. See Supplemental Material at <http://link.aps.org/supplemental/10.1103/PhysRevE.101.062405> for supplementary figures of non-averaged datasets.
44. MATLAB (The MathWorks, Natick, MA, 2016).
45. To further reduce the noise we applied a Gaussian smoothing kernel with a standard deviation of 2 pixels (“imgaussfilt” in MATLAB [44]). The derivatives of the velocity were calculated in MATLAB using the function ‘gradient [44].
46. Hardy RJ, Formulas for determining local properties in molecular-dynamics simulations: Shock waves, *J. Chem. Phys* 76, 622 (1982).
47. Fabry B, Maksym GN, Butler JP, Glogauer M, Navajas D, and Fredberg JJ, Scaling the Microrheology of Living Cells, *Phys. Rev. Lett* 87, 148102 (2001). [PubMed: 11580676]
48. Sollich P, Lequeux F, Hébraud P, and Cates ME, Rheology of Soft Glassy Materials, *Phys. Rev. Lett* 78, 2020 (1997).
49. Douezan S, Guevorkian K, Naouar R, Dufour S, Cuvelier D, and Brochard-Wyart F, Spreading dynamics and wetting transition of cellular aggregates, *Proc. Natl. Acad. Sci* 108, 7315 (2011). [PubMed: 21504944]
50. Bonakdar N, Gerum R, Kuhn M, Spörrer M, Lippert A, Schneider W, Aifantis KE, and Fabry B, Mechanical plasticity of cells, *Nat. Mater* 15, 1090 (2016). [PubMed: 27376682]
51. Banerjee S and Marchetti MC, Substrate rigidity deforms and polarizes active gels, *EPL Europhys. Lett* 96, 28003 (2011).
52. Oron A, Davis SH, and Bankoff SG, Long-scale evolution of thin liquid films, *Rev. Mod. Phys* 69, 931 (1997).
53. Craster RV and Matar OK, Dynamics and stability of thin liquid films, *Rev. Mod. Phys* 81, 1131 (2009).
54. Howison S, *Practical Applied Mathematics: Modelling, Analysis, Approximation* (Cambridge University Press, Cambridge, England, 2005).

FIG. 1.

(a) 3D schematic of a thin film of fluid (or epithelial monolayer) on a rigid substrate. Not drawn to scale—the length and width of the film are substantially larger than the height. (b) $x - z$ cross section. The top surface is stress free while at the bottom, tractions are balanced by viscous shear stresses. In this work, tractions (T_x and T_y) refer to the measured tractions by the cells on the substrate, $T_{\text{cell on substrate}}$. The dashed lines refer to boundary conditions at the edges, discussed in Sec.III.

FIG. 2.

Schematic of a cell island approaching an obstacle. The cell island is given by a solid black line. Different scenarios, where the cell island is imaged fully or partially, are color coded. See text for case 4 edge conditions.

FIG. 3.

(a) Phase image of the migrating cells encompassing the obstacle. The field of view is $725 \times 725 \mu\text{m}^2$. Measured velocities (b) v_x and (c) v_y (color bar is in units of 10 nm/s). Raw data from Ref. [16].

FIG. 4.

(a) Measured tractions T_x^{meas} [Pa] used in Ref. [16]. (b) Calculated tractions $\mu h t_x^{\text{calc}}$ [Pa] with $h = 5 \mu\text{m}$ and $\mu = 23 \text{ kPa s}$. (c) The active force density, $h F_x^{\text{active}} = T_x^{\text{meas}} - \mu h t_x^{\text{meas}}$ [see Eq. (28) below]. (d)–(f) A zoomed view of the marked green box from panel (c), showing (d) $|T^{\text{meas}}|$ [Pa], (e) $|\mu h t^{\text{calc}}|$ [Pa], and (f) $|h F^{\text{active}}|$ [Pa]. The green arrows show the direction of each respective vector of (d)–(f) where the lengths of the arrows are proportional to the magnitude.

FIG. 5.

Scatter plot of T_x^{meas} [Pa] vs $\mu h t_x^{\text{calc}}$ [Pa]. The color bar is proportional to the count density.

TABLE I.

Viscosity estimates from various experimental setups. The first two rows are from this work.

Method [References]	$10^6 \times (\mu/\mu_{\text{water}})$
Scaling (Th/u)	5
Coarse graining ($\langle T^{\text{meas}} \rangle_{\text{rms}}/\langle \mu^{\text{calc}} h \rangle_{\text{rms}}$)	23
Low frequency G' [47]	1
Micropipette aspiration [40]	200
Expanding island assays [30]	$10^2\text{--}10^4$
Expanding island-active stresses [31,32]	$(0.5\text{--}5)10^3$

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript