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Virus-Sized Gold Nanorods: Plasmonic Particles for Biology

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Conspectus

Plasmons, collective oscillations of conduction band electrons in nanoscale metals, are well-known phenomena in colloidal gold and silver nanocrystals that produce brilliant visible colors in these materials that depend on nanocrystal size and shape. Under illumination at or near the plasmon bands, gold and silver nanocrystals exhibit properties that enable fascinating biological applications: (i) the nanocrystals elastically scatter light, providing a straightforward way to image them in complex aqueous environments; (ii) the nanocrystals produce local electric fields that enable various surface-enhanced spectroscopies for sensing, molecular diagnostics, and boosting bound fluorophore performance; (iii) the nanocrystals produce heat, which can lead to chemical transformations at or near the nanocrystal surface, and can photothermally destroy nearby cells.

While all the above-mentioned applications have already been well-demonstrated in the literature, this Account focuses on several other aspects of these nanomaterials, in particular gold nanorods that are approximately the size of viruses (diameters ~10 nm, lengths up to 100 nm). Absolute extinction, scattering, and absorption properties are compared for gold nanorods of various absolute dimensions, and references for how to synthesize gold nanorods of four different absolute dimensions are provided. Surface chemistry strategies are detailed that coat nanocrystals with smooth or rough shells; specific examples include mesoporous silica and metal-organic framework shells for porous (rough) coatings, and polyelectrolyte layer-by-layer wrapping for “smooth” shells. For self-assembled monolayer molecular coating ligands, the smoothest shells of all, a wide range of ligand densities have been reported from many experiments, yielding values from less than 1 to nearly 10 molecules/nm² depending on nanocrystal size and ligand nature. Systematic studies of ligand density for one particular ligand with a bulky headgroup are highlighted, showing that the largest ligand density occurs for the smallest nanocrystals, even though these ligand headgroups are the most mobile as judged by NMR relaxation studies. Biomolecular coronas form around spherical and rod-shaped nanocrystals upon immersion into biological fluids; these proteins, and lipids, can be quantified, and their degree of adsorption depends on nanocrystal surface chemistry as well as biophysical characteristics of the adsorbing biomolecule. Photothermal adsorption and desorption of proteins on nanocrystals depend on the enthalpy of protein-nanocrystal surface interactions, leading to light-triggered alteration in protein concentrations near the nanocrystals. At the cellular scale, gold nanocrystals exert genetic changes at the mRNA level, with a variety of likely mechanisms that include alteration of local

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biomolecular concentration gradients, changing mechanical properties of the extracellular matrix, and physical interruption of key cellular processes - even without plasmonic effects. Microbiomes, both organismal and environmental, are the likely first point of contact of nanomaterials with natural living systems; we see a major scientific frontier in understanding, predicting, and controlling microbe–nanocrystal interactions, which may be augmented by plasmonic effects.

Graphical Abstract

1. Introduction

Nanoscale size and shape matters in biology: cells engulf 10-1000 nm objects by different mechanisms that depend on object size.^{1,2} Smaller colloidal nanoparticles are cleared from organisms faster than larger ones.³⁻⁵ Disc-shaped or rod-shaped colloidal particles appear to

evade the immune system better than spherical ones.^{6,7} Viruses are nanoscale objects: 18×300 nm are the dimensions of the common tobacco mosaic virus.⁸

Inorganic solids on the nanoscale show size- and shape-dependent properties. The mean free path of electrons in a metal at room temperature is ~ 10 - 100 nm, depending on the metal, while the excitonic Bohr radius in semiconductors is ~ 1 - 10 nm.^{9,10} Therefore, virus-sized metal nanocrystals and protein-sized semiconductor nanocrystals are expected, and have been found, to have fascinating size- and shape-dependent optical properties. Gold and silver nanocrystals of roughly 5 - 300 nm support a localized surface plasmon resonance (LSPR), which is the coherent oscillation of conduction band electrons at the metal surface upon illumination.¹¹⁻¹³ While nanospheres produce a single LSPR, it is well-known that nanorods exhibit two LSPRs: one corresponding to the transverse axis (transverse plasmon) and the other corresponding to the longitudinal axis (longitudinal plasmon). The longitudinal surface plasmon resonance can be tuned by varying either the length to width ratio (aspect ratio) or the effective volume of the nanorods. For gold and silver, these LSPRs are in the visible and near-infrared portions of the electromagnetic spectrum, due to their dielectric constants (Figure 1).

Extinction in these colloidal metal nanocrystals is the sum of absorption plus (elastic light) scattering, and larger nanocrystals scatter more.^{13,16} Thus, larger colloidal metal nanocrystals can serve as effective imaging agents just based on their elastic light scattering properties, with no bleaching and no blinking (Figure 2).¹⁷

The plasmons provide far more capabilities than elastic light scattering. Upon illumination, local electric fields are generated that allow for surface-enhanced molecular spectroscopies for local diagnostics, of which surface-enhanced Raman scattering is the most famous;¹⁸ and upon sufficient illumination, the large absorption cross-sections of metallic nanocrystals leads to heating.^{13,19} This plasmonic heating, described in more detail below, is sufficient to kill living cells, and there is great excitement in the field about photothermal cancer therapy that relies on this light-to-heat conversion.²⁰

2. Controlling the absolute dimensions of gold nanorods

The interplay of scattering and absorption of nanoscale metal particles as a function of absolute size means that the relative abilities of imaging, via scattering, and photothermal heating, via absorption, depend on absolute particle size. Early calculations of extinction, scattering and absorption cross-sections for gold nanorods (AuNRs) and gold nanospheres (AuNSs), show, for example, that 40 nm gold nanospheres scatter 6% of the light compared to that absorbed, while 80 nm spheres correspondingly scatter 68% .¹³

The most common way to make AuNRs is via seed-mediated growth methods in aqueous solution at room temperature, wherein nucleation and growth of the particle are separated.²¹ Typically, a concentrated solution of surfactant (cetyltrimethylammonium bromide, CTAB)-stabilized gold seeds is made via the reduction of auric ions with a strong reducing agent. The seeds are then placed into a growth solution containing more surfactant, auric ions, silver ions, and a weak reducing agent to directly reduce gold ions onto the surface of the

seed particles.²² The aspect ratio (AR; length/width) of the AuNRs is tunable depending on reagent concentration. More seeds mean less gold will be available per seed, so a smaller product size dominates, and vice versa for a smaller seed concentration. Overall, in the literature, four different absolute size regimes of AuNRs have been reported, for seemingly minor changes in synthetic protocol.²¹⁻²⁵ Figure 3 represents and Table 1 summarizes these four synthetic methods used to create AuNRs of controllable aspect ratio and of controlled absolute size.^{21,23-25}

3. Making gold nanorods look like viruses: porous coatings

The as-made AuNRs, no matter which of the four synthetic methods used, bear a bilayer coating of the cationic CTAB surfactant on the surface.¹⁴ As CTAB alone is cytotoxic to at least some cell lines at micromolar concentrations, chemical strategies to remove it, cover it up, or replace it have been employed, detailed below.

The surface coats of viruses can be thought of as patterned, not homogeneous. We have been able to create AuNRs that have regularly varying “rough” surfaces by using nanoporous coatings made of inorganic materials. Porous shells around AuNRs include mesoporous silica.³²⁻³⁴ Metal-organic frameworks (MOFs) are famed porous materials, although MOF shells are often not as uniform as their silica counterparts.^{29,35,36}

Mesoporous silica’s popularity stems from its chemical robustness and ease of synthesis, biocompatibility, and stability in a wide range of solvents.¹⁴ We developed a synthesis for mesoporous silica-coated gold nanorods with a shell thickness that can be tuned by altering the growth solution CTAB concentration (Figure 4a & 4b).³² We showed that controlling the shell thickness allows for optimization of plasmon-enhanced fluorescence of low-quantum-yield near-infrared fluorophores covalently anchored to the silica surface; a maximum 10-fold fluorescence enhancement was achieved with 17 nm shells.³²

The ~3 nm (and now larger) pores mesoporous silica coatings on gold nanorods can be used for molecular uptake and photothermal release of payloads.^{33,34,37} Development of additional methods for expanding the pore sizes would widen the range of applications, including the sequestration, sensing, and delivery of larger biomolecules.

MOFs have extremely high surface areas, defined pore channels, and pore sizes most often less than 3 nm.³⁸ AuNRs can be encapsulated with MOFs through either a one-pot reaction of AuNRs and the MOF precursors or a layer-by-layer process (LbL).^{29,35,36}

We developed a LbL process for the MOF HKUST-1 around AuNRs through alternating additions of copper acetate and an organic salt of the linker 1,3,5-benzenetricarboxylate.²⁹ With this method, the shells are more uniform and the thickness can be precisely controlled within single nanometers (Figure 4).²⁹ The HKUST-1 shell growth rate was found from the slope of the regression line in Figure 4d and it was determined the MOF shell grows by 0.178 ± 0.014 nm per layer. While MOFs with larger pores have been synthesized, there have not been reports of these being used as shell materials around nanoparticles. In order to have larger mesopores, the MOF shell has to be post-synthetically etched, which in turn loses the desirable defined pore channels. AuNR@MOF materials have demonstrated the

unique capability as a combined photothermal and chemotherapeutic agent through stimuli responsive drug release.^{35,39}

4. Making gold nanorods look like viruses: soft organic coatings

Gold nanorod surface modification can be achieved using LbL deposition of polyelectrolytes (PEs), which increases the thickness of the surface layer, and also allows the reversal of surface charge. This approach, similar to standard LbL techniques,⁴⁰ involves alternatively adding anionic and cationic PE layers onto the oppositely-charged AuNRs through electrostatic and hydrophobic interactions. For optimal PE coating the polymers should have a chain length comparable to the dimensions of the nanoparticles, and the ionic strength should allow the polymers to be in a flexible, extended conformation. Without these considerations the nanoparticles can flocculate due to multimer formation as a result of bridging, polymer rigidity, and/or insufficient electrostatic repulsion.⁴¹

UV-Vis spectroscopy can monitor the formation of PE coatings on AuNRs (Figure 5), where shifts in the plasmon peak maximum are observed, in an alternating fashion after each succession coating of the anionic PE poly(acrylic acid) (PAA), and cationic PE poly(allylamine hydrochloride) (PAH)⁴² These shifts are caused by the change in refractive index of the surrounding media.^{41,42} It is important to note particle loss after each layer-by-layer coating due to the remove of excess PE by centrifugation; successful coating is limited to about 10 layers.⁴³ The most effective way to characterize these successive PE depositions is by measuring ζ -potential to confirm the charge reversal after each successful coating (Figure 5c). The PE layer thickness (h) can be extracted from hydrodynamic diameter measurements from dynamic light scattering (DLS; Figure 5d):⁴²

$$d_H = \frac{k_B T}{3\pi\eta D_t} \quad (1)$$

$$D_t = \frac{k_B T \ln\left(\frac{L}{d_{CS}}\right) + C_t}{3\pi\eta L} \quad (2)$$

$$C_t = 0.312 + 0.565\left(\frac{d_{CS}}{L}\right) - 0.100\left(\frac{d_{CS}}{L}\right)^2 \quad (3)$$

$$L = L_{AuNR} + 2h \quad (4)$$

$$d_{CS} = d_{AuNR} + 2h \quad (5)$$

where d_H is the hydrodynamic diameter due to the translational diffusion of the AuNRs, k_B is the Boltzmann constant, T is temperature of the sample solution, η is the viscosity of the solution, D_t is the translational diffusion constant, L is the total length of the coated AuNR, d_{CS} is the total diameter of the coated AuNR, L_{AuNR} and d_{AuNR} are the bare average length and diameter of the AuNRs measured from transmission electron micrographs.

LbL PE deposition is a facile method to functionalize AuNR surfaces with controlled surface charge and coating thickness. This also allows for loading of molecules on or within the PE layers to make AuNRs more like soft, organic nanoparticles.¹⁴ In addition, molecular self-assembled monolayers (SAMs) can also be employed to modify the surface of AuNRs making them more “virus”- like. Hydrophobic octadecanethiol-coated AuNSs functionalized with lipids, or surfactants, form a “hybrid” lipid bilayer on AuNSs.⁴⁴ The non-polar tail of the lipid/surfactant associates with the non-polar chain of the self-assembled monolayer of octadecanethiol.

5. What does the molecular landscape look like on a gold nanocrystal?

Ligand density analysis.

Viruses are miracles of molecular-scale ordering over tens or hundreds of nanometers. Molecular self-assembled monolayers (SAMs) have been employed to modify the surface of nanoparticles. SAMs of thiolated molecules on gold surfaces are well-known and are usually considered to be “molecularly smooth”.⁴⁵ On flat gold surfaces, small thiols have ligand densities of 2-4 molecules/nm², while ~1-2 nm gold nanocrystals have been reported to have 9 thiol molecules/nm² (Table 2).⁴⁶⁻⁵⁴

We show literature ligand density values for different SAMs on gold (Table 2). In our own work,⁴⁶ we find that ligand density of a long-chain thiol with a bulky headgroup varies as a function of AuNS diameter, with the smallest particles having the largest ligand density (Figure 6b), yet the headgroup mobility, by NMR, is largest for the smallest nanoparticles (Figure 6a & 6c). In concert with molecular dynamics simulations, we reconcile these seemingly contradictory results by realizing that the bulky quaternary ammonium headgroup of the ligand jams for particle core sizes larger than 10 nm, but ligand flexibility on smaller particles allows both for higher packing densities and more headgroup mobility (Figure 6d).

6. Biomolecular coronas and their response to plasmonic heating

The surfaces of nanoparticles can be altered by their introduction into biological or natural environments, whereupon they acquire coatings of biomolecules, called coronas.⁵⁵ The formation and composition of these coronas are dependent upon both the physical and chemical properties of the nanoparticles, such as size, shape, and surface chemistry.

Denaturation behavior of a protein might be a useful predictor of adsorption behavior to AuNSs.⁵⁶ We studied protein adsorption to anionic citrate-capped AuNSs and to cationic PAH wrapped AuNSs for three proteins, α -amylase (A-Amy), β -lactoglobulin (BLG), and bovine serum albumin (BSA).⁵⁶ Langmuir adsorption isotherm results showed that BSA > BLG >> A-Amy affinities for citrate-capped AuNSs, while PAH-coated AuNSs displayed little affinity for these proteins, in accord with electrostatics.⁵⁶ Changes in the secondary structure of proteins on AuNSs did follow the trends predicted by acid denaturation characteristics for both citrate-AuNSs and PAH AuNSs.⁵⁶ While protein coronas have drawn much attention, other biomolecular coronas such as lipid coronas are equally important. We recently developed a LC/MS/MS method to assess the degree of lipid corona formation on 14 nm AuNSs upon incubation with lipid vesicles.⁵⁷ Cationic (16-Mercapto-hexadecyl) trimethylammonium bromide-coated AuNSs (MTAB-AuNPs) and PAH-AuNSs were compared, where MTAB displays a quaternary ammonium group to the solvent, while PAH displays primary amines. Cationic MTAB-AuNPs and PAH-AuNPs extracted 60–95% of the lipids available (Figure 7), while anionic citrate-AuNPs extracted almost none.⁵⁷ The primary amine polymer surfaces extracted more lipids than the quaternary ammonium surfaces.⁵⁷

The property of heat generation upon light excitation makes plasmonic nanoparticles extremely interesting for nanomedicine applications, such as controlled photothermal release of molecules, and for photothermal therapy. This heat generation phenomenon is explained by the strong absorption of the incident light from an external source (e.g. near-infrared (NIR) laser) by the plasmonic particles leading to the excitation of free electrons. Via electron-electron collisions, hot electrons are created. Then, the fast transfer of energy from the electron to the phonon via electron-phonon coupling generates heat with electron temperatures rising in the vicinity of plasmonic nanoparticles by few tens to thousands of degrees (on short time scales) (Figure 8).¹⁹

The composition of protein coronas formed on AuNRs after incubation in fetal bovine serum (~5000 different proteins at mM concentrations) differs significantly when subjected to plasmonic heating compared to conventional thermal heating.⁵⁸ Photoinduced heating lead to an increase in several biologically relevant proteins bound to the particles, including serum albumin, α -2-HS-glycoprotein precursor, apolipoprotein A-II precursor and apolipoprotein C-III precursor which have the potential to affect blood circulation time and the ability of particles to cross the blood-brain barrier.⁵⁸ Understanding how laser irradiation affects the corona composition of plasmonic nanoparticles and therefore their interactions with other biological entities is vital for the successful photothermal drug delivery and cancer therapy.⁵⁹

The photothermal effect produced by plasmonic nanoparticles upon irradiation has been explored for the thermally-induced release of molecules (e.g. drugs, DNA, siRNA, proteins) into cells for targeted therapeutics.⁶⁰ This strategy could significantly enhance treatment efficacy while reducing/eliminating the undesirable side effects of drugs, and prevents the therapeutic payload from degradation. However, cargo release using photothermal heating of plasmonic nanoparticles works only if the molecule-nanoparticle interaction is exothermic in nature, following Le Chatelier's principle.⁶¹ By investigating the interactions between

protein/PE-wrapped AuNR pairs that had either endothermic or exothermic associations, we found that the endothermic system increased protein absorption to the surface, while the opposite was observed in the exothermic one, (Figure 9) according to:

Therefore, Le Chatelier's principle can be applied to the nanoscale to predict protein-nanoparticle interactions and the successful release of biomolecules from the surface of nanoparticles upon irradiation, even when more complex biological environments are considered.

7. Photothermal Killing of Pathogenic Bacteria

Multidrug resistance (MDR) bacteria present a great threat to public health. The photothermal properties of gold nanocrystals have also been exploited to kill pathogenic bacteria, including MDR ones. We applied PE-coated AuNRs conjugated with antibodies as photothermal agents to selectively kill the Gram-negative MDR bacterium, *Pseudomonas aeruginosa*.⁶² (Figure 10). The treatment with AuNRs with plasmonic excitation led to a significant decrease in cell viability and irradiated cells showed irreparable cell membrane damage. Gold nanoparticle-mediated photothermal killing is effective against other different strains of bacteria, from Gram-positive to Gram-negative,⁶³ and it is a promising strategy to destroy bacterial biofilms.⁶⁴

8. Virus-Sized Gold Particles and Interactions with Biological Systems

The understanding of the interactions of nanoparticles with biological systems is essential for the safe human exposure to nanoscale materials. By investigating the interactions of gold nanocrystals with the extracellular matrix and monitoring cell behavior, we noticed changes in the mechanical properties of the extracellular matrix when these nanoparticles were added in the system, promoting the migration of cancer cells, which is a concern.⁶⁵

The surface chemistry is one of the major intrinsic parameters governing nanocrystal toxicity, playing a key role in nano-bio interactions. Transcriptomic analyses performed by us on two cell types exposed to four differently-coated AuNSs showed significant changes in the expression of genes involved in pathways such as cell proliferation, inflammation and cell death according to their surface chemistry. Surprisingly, the biomimetic lipid-coated AuNPs affected the highest number of genes, highlighting the complexity of nano-bio interactions.⁶⁶

Currently, very little is known about the long-term effects of nanocrystal exposure on living systems. To shed light on this issue, we assessed the effects of a low dose of AuNSs and AuNRs with different surface chemistries to human cells under chronic and acute exposures after 20 weeks.⁶⁷ Although no significant impact on cell viability was observed, analysis at

the molecular level revealed that the nanocrystals induced long-term changes in the expression levels of cell stress response genes. Intriguingly, the acute exposure (24h) led to a sustained cell stress response, detectable even 20 weeks after the stimulus, indicating that cells did not recover properly over time.⁶⁷ But upon chronic exposure, cells adopted an altered state in response to the continual stress (nanoparticles).⁶⁷ Therefore, cells adapt to nanocrystal stresses over time.

In vivo data on fate, bioaccumulation and toxicity of gold nanoplateforms over long time periods are still lacking. The El-Sayed group investigated the long-term toxic effects of AuNRs in mice and, although no evident toxicity was found at 15 months post-injection, most of the AuNRs still remained inside the hepatocytes and spleen cells.⁶⁸

9. Virus-sized gold nanoparticles in the environment: from soil to bacteria

Nano-enabled consumer products began to appear in the marketplace in 1999. These nanomaterials can be released into ecosystems through air, soil, sludge, wastewater, sea water, and freshwater from the beginning of manufacturing to the disposal of nanomaterial-enabled products. To understand how these engineered nanomaterials interact with ecosystems, we have investigated gold nanoparticle (AuNP) distributions and effects on soil,⁶⁹ groundwater,⁶⁹ and microbiomes.^{70,71}

Prior to studying NP influences on microbiomes, we examined how changes in physicochemical properties of AuNPs affect their soil retention and colloidal stability in groundwater.⁶⁹ AuNPs of different shapes, surface chemistry and charges were passed through soil columns. When eluted by simulated groundwater, four-fold more spherical AuNPs were retained in the soil column compared to rod-shaped counterparts, for various surface chemistries. However, we found that spherical AuNPs regardless of charges aggregated within several hours when incubated with simulated groundwater. We also observed that compared to polyelectrolyte-wrapped AuNPs, AuNPs wrapped with relatively labile capping agents were more susceptible to irreversible aggregation in simulated groundwater.⁶⁹

We have evaluated NP toxicity and the interaction of NPs with diverse bacteria. In one study, we chose a panel of bacteria present in diverse habitats.⁷⁰ Each bacterium possesses a unique cell wall structure made up of either smooth or rough lipopolysaccharides (LPS) with different phosphate unites and charges. We observed that bacteria with smooth LPS bound more AuNPs than those with rough LPS.⁷⁰ Toxicity to bacteria was observed, even though NPs did not enter the bacteria; again, compromised cell membranes appear to be one of the root causes.⁶⁹ In another study, we utilized metagenomics and found that AuNSs and AuNRs, at low doses for 56 days, altered the composition of microbial communities and increased levels of antibiotic resistance genes (ARGs).⁷¹ This result is consistent with a study which shows 40 ARGs were significantly enriched in the presence of rare earth oxide nanoparticles.⁷²

As gold is of low natural abundance in the environment, and inductively-coupled plasma mass spectrometry (ICP-MS) detection limits are at the part-per-trillion level, we have

measured the distribution of both initially cationic and anionic AuNRs in model estuarine mesocosms, at doses that correspond to that of a viral “infection” in an ecosystem^{73,74} After two weeks of exposure, the microbial biofilms were found to be the main sink in the living food web, and the initial surface chemistry of the nanorods did dictate ultimate environmental distributions.^{73,74} On top of these observed changes in microbial community composition as a function of AuNP exposure, recent work has shown that bacterial biofilms associated with an aquatic plant can spontaneously produce sufficient cyanide to oxidize gold according to:⁷⁵

Thus, another mode of biological damage that involves bacteria is the unexpected production of more inherently toxic gold ions, compared to elemental gold, by microbial biofilms under certain conditions.

10. Conclusions and future directions

This Account has surveyed a great deal of ground: synthesis, surface chemistry, biomolecule-surface interactions, cellular interactions, and environmental fate and impacts of AuNPs. Much has been done, and much remains to do. What are the most impactful future directions?

Patchy particles: measurement and impact.

Just as virus particles display ordered yet varied surfaces, colloidal nanocrystals could in principle display similar functionalities, as opposed to “monoculture” surfaces. The synthesis of nanocrystals bearing multiple functional groups is reasonably straightforward; but the measurement of individual particle “surface identities” is not at all straightforward. With the advent of improving single-particle, single-molecule, and cryo electron microscopy methods, progress is likely to be rapid in this area. In principle, one could design artificial nanocrystals that have both the symmetry and variety of natural viral particles. But even for simple “monoculture surface” systems, nanocrystal diameter affects molecular display.⁴⁶

Microbiomes plus nanoparticles.

Of all the biological systems that could interact with nanocrystals, we believe bacteria are the most important. Microbiomes are abundant in the open environment, within organisms and might be the first point of contact between an engineered nanomaterial and anything – soil, GI tract, etc. We speculate that one reason many nanoparticle-enabled drug delivery experiments fail in an organism is due to not taking into account the relevant microbiomes.⁷⁶ Detailed understanding and prediction of how nanocrystals are processed by microbiomes and impact their resident bacterial populations are key research areas for future study.

Plasmonic biology.

Just as optogenetics has opened new doors in understanding and controlling, especially, the brain, can “plasmonogenetics” be a new field of scientific inquiry, to understand and control

cellular and organismal functions? As organisms regulate temperature far more closely than they regulate light absorption, this is a fanciful suggestion. But as more data accumulates about cellular responses to nanoparticle exposure that include turning on “unfolded protein response” genes,⁶⁷ one can imagine that the fundamental disordering of protein structure due to nanocrystal binding could have cascading impacts on biological function at higher levels.

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Biography

Catherine J. Murphy holds the Larry R. Faulkner Endowed Chair in Chemistry at the University of Illinois at Urbana-Champaign (UIUC). She earned two B.S. degrees from UIUC in 1986, one in chemistry and one in biochemistry. Her Ph.D. (1990) is from the University of Wisconsin, Madison, with A. B. Ellis. After postdoctoral fellowships at the California Institute of Technology with J. K. Barton (1990-1993), she joined the faculty of chemistry and biochemistry at the University of South Carolina. After rising through the faculty ranks at South Carolina (1993-2009), she was recruited back to UIUC in 2009.

Huei-Huei Chang is from Taipei, Taiwan and received a B.A. degree in Chemistry from Iowa State University. She is currently a Ph.D. candidate at UIUC in the Murphy group. Her research interests focus on how the size and surface chemistry of gold nanoparticles affect cellular responses.

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Xi Zhang received her B.S. degree in Clinical Pharmacy at Fujian Medical University with an internship at the Second Hospital of Fuzhou in China, 2012. She received her Master of Medicine degree in Pharmaceutical Analysis at Fujian Medical University in 2015. She joined the Murphy research group in 2015 to pursue a PhD degree in chemistry. Her research interests include the interaction and impact of gold nanoparticles on biological systems.

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Figure 1.

a) Schematic of the localized surface plasmon resonances of gold nanorods. (Reprinted with permission from Ref. 14. Copyright 2016 American Chemical Society) b) Normalized Vis-NIR extinction spectra and c) TEM images (scale bars on images) of gold nanorods with varying aspect ratios (black AR = 2.4, red AR = 2.7, blue AR = 3.6, pink AR = 4.4, green AR = 6.1). (*Faraday Discuss.*, **2016**, 191, 9-33. Reproduced by permission of The Royal Society of Chemistry).¹⁵

Figure 2. Darkfield optical micrograph of human dermal fibroblast cells incubated with polyallylamine-coated gold nanospheres that are 90 nm in diameter; all bright spots are due to scattered light from the gold nanospheres, which have been ingested by the unstained cells. Scale bar = 20 microns. (Reprinted with permission from Ref 17. Copyright 2013 American Chemical Society).

Figure 3:

A representative transmission electron micrograph for each synthesis method referred to in the text. The scale bars are 100 nm.

Figure 4:

Gold nanorods (AuNRs) with porous shells a) Transmission electron micrographs of silica-coated AuNRs with varying shell thicknesses. Scale bar 50 nm. b) CTAB concentration in growth solution versus silica shell thickness. (Reprinted with permission from Ref. 32. Copyright 2014 American Chemical Society) c) Transmission electron micrographs of HKUST-1 coated gold nanorods. Scale bars 50 nm. d) HKUST-1 shell thickness related to number of layers deposited. (Reprinted with permission from Ref. 29. Copyright 2018 American Chemical Society)

Figure 5.

LbL surface coating of AuNRs with PAA and PAH. a) Visible spectra of AuNRs as a function of numbers of alternating PE coatings: curve 0= as-prepared GNRs, curves 1,3,5= PAA, 2,4,6= PAH. b) Longitudinal plasmon peak maxima for AuNRs as a function of PE layers c). Zeta potential data for AuNRs as a function of PE coating d) Effective hydrodynamic diameters (d_h) of AuNRs with PE coatings measured by DLS, and equivalent thickness (h) of PE layers calculated by equation 1-5; alterations in PE layer thickness are thought to result from different degrees of hydration within the layers. (Reprinted with permission from Ref. 42. Copyright 2013 American Chemical Society)

Figure 6.

Graphs of (a) the chemical shift of MTAB headgroup protons, (b) MTAB ligand density from two independent methods, (c) T_2 (orange line) and T_2^* (blue line) relaxation times, and (d) MTAB conformation and packing, as a function of AuNS diameter. (Reprinted with permission from Ref 46. Copyright 2019 American Chemical Society)

Figure 7:

$C_{\text{bound}}/C_{\text{total}}$ percentage, after pull-down separation of AuNP pellets, based on LC-MS/MS quantification. The lipids are a) DOPC, b) 9:1 DOPC/DOPS, c) 9:1 DOPC/DOPG, d) 9:1 DOPC/DOPE. Bar heights represent mean values; error bars correspond to one standard deviation for triplicate runs from the same batches of nanoparticle and lipid vesicle solutions. Asterisks correspond to the following p values using ANOVA followed by post hoc multiple comparisons test: * $p < 0.05$; ** $p < 0.01$, *** $p < 0.001$. (Reprinted with permission from Ref. 57. Copyright 2018 American Chemical Society).

Figure 8:

Schematic representation of the photothermal heating process in gold nanoparticles. (1) Photoexcitation of free electrons in the plasmon band. (2) Relaxation via electron-electron scattering leading to a rapid increase in the nanoparticle surface temperature. (3) Heat transfer from the electrons to the gold lattice phonons via electron-phonon coupling. (4) Heat dissipation from the gold lattice to the medium surrounding the plasmonic nanostructure through phonon-phonon interactions.

Figure 9:

Cartoon showing the behavior of proteins on nanoparticle surfaces given the thermodynamics of each system. The behavior is tested by measuring the protein concentration in the supernatant following NIR laser irradiation. Endothermic reactions favor the complex upon addition of heat, producing less protein in the supernatant, while exothermic reactions favor free reactants, leading to higher protein concentration in the supernatant. (Reprinted with permission from Ref. 61. Copyright 2017 American Chemical Society)

Figure 10:

Selective photothermal killing of pathogenic bacteria using AuNRs. The left panel shows the viability of *Pseudomonas aeruginosa* cells with attached AuNRs following exposure to NIR light: a) Control cells without nanorods or NIR exposure; b) Cells without nanorods exposed to NIR; c) Cells with nanorods and no NIR exposure; and d) Cells with nanorods and exposed to NIR for 10 min. Green fluorescent cells are live while red fluorescent cells are dead or compromised. e) TEM image (30000×) of *P. aeruginosa* cells with attached antibody-conjugated nanorods following 10 min exposure to NIR. The arrows indicate irreparable damage in the bacterial cell surface. (Adapted with permission from Ref. 62. Copyright 2008 American Chemical Society)

Table 1:

List of AuNR properties for four different synthetic methods in the literature (which we name mini, Murphy, Zubarev and Murray).^{21,23-25} Each synthetic method produces tunable aspect ratio nanorods. SPR = surface plasmon resonance. Molar extinction coefficients are reported in water on a per-particle basis.

AuNR Type	Dimension Range (nm) L × W	Aspect Ratio Range	Reported SPR Range	Scattering % of Total Extinction		Molar Extinction Coefficients (M ⁻¹ cm ⁻¹)		Ref.
				Experimental	Calculated			
Mini	L: 20-100 nm W: 5-10 nm	≈2-11	Longitudinal: 600-1300 nm Transverse: 504-523 nm	N/A	0.5 – 2.5 %	AR ≈ 2.2 (λ = 607)	3×10 ⁸	23
						AR ≈ 3.8 (λ = 793)	2×10 ⁸	
						AR ≈ 8.2 (λ = 1040)	9×10 ⁸	26
Murphy	L: 20-100 nm W: 10-20 nm	≈2-5	Longitudinal: 650-950 nm Transverse: 504-523 nm	20 – 30 %	7 – 10%	AR ≈ 2 (λ = 605)	2.5×10 ⁹	21
						AR ≈ 3 (λ = 728)	3.9×10 ⁹	27
						AR ≈ 3.5 (λ = 785)	4.6×10 ⁹	13
						AR ≈ 4.5 (λ = 845)	5.5×10 ⁹	28
Zubarev	L: 50-100+ nm W: 10-30 nm	≈ 2-7	Longitudinal: 770-1160 nm Transverse: 504-523 nm	25 – 40 %	20 – 40 %	AR ≈ 4 (λ = 800)	1.6×10 ¹⁰	24
								27
								13
								29
Murray	L: 70-190 nm W: 15-75 nm	≈ 2-7	Longitudinal: 650-1150 nm Transverse: 504-540 nm	30 – 50 %	30 – 70 %	AR ≈ 2.9 (λ = 787)	*7.9×10 ¹⁰	25
						AR ≈ 3.6 (λ = 794)	*4.6×10 ¹⁰	30
								13
								31

* Extinction coefficients reported are for AuNRs with silica coating.

Table 2:

Summary of ligand density of representative SAMs in the literature.

Ligand type	Gold surface	Ligand density / molecule per Nm^2 (method)	Ref.
CH_3SH	Flat Au(111) surface	4.65 (XPS)	47
3-Mercaptopropionic acid (MPA)	AuNSs (13.2 ± 1.4 nm to 26.2 ± 4.4 nm)	6.3 ± 0.6 (ICP-MS)	48
6-Mercaptohexanoic acid (6-MHA)	AuNSs (13.0 ± 1.3 nm)	4.6 ± 0.1 (XPS)	49
8-Mercaptooctanoic acid (MOA)	AuNSs (13 ± 1 nm and 31 ± 1 nm)	4.0 ± 0.1 (qNMR after aqua regia digestion)	50
11-Mercaptoundecanoic acid (MUA)	AuNSs (13 ± 1 nm and 31 ± 1 nm)	4.4 ± 0.1 (qNMR after aqua regia digestion)	50
	AuNSs (13.2 ± 1.4 nm to 26.2 ± 4.4 nm)	5.7 ± 0.1 (ICP-MS)	48
	AuNSs (12.6 ± 1.2 nm)	5.0 ± 0.1 (XPS)	49
Dodecanethiol	AuNSs (2.4 ± 0.1 nm)	9.30 (SAXS and elemental analysis)	51
16-Mercaptohexadecanoic acid (16-MHA)	AuNSs (13.2 ± 1.4 nm to 26.2 ± 4.4 nm)	5.3 ± 0.2 (ICP-MS)	48
(16-mercaptohexadecyl)trimethylammonium bromide (MTAB)	AuNSs (4.8 ± 1.1 nm to 25.0 ± 4.4 nm)	3.0 ± 0.8 to 6.5 ± 1.8 (in situ qNMR) 1.8 ± 0.4 to 4.8 ± 1.1 (qNMR after I_2 digestion)	46
SH-PEG ₄ -COOH	AuNSs (13.2 ± 1.4 nm to 26.2 ± 4.4 nm)	5.0 ± 0.3 (ICP-MS)	48
SH-PEG ₇ -COOH	AuNSs (13.2 ± 1.4 nm to 26.2 ± 4.4 nm)	4.3 ± 0.4 (ICP-MS)	48
mPEG-SH (average Mw=1000 Da)	AuNSs (13 ± 1 nm and 31 ± 1 nm)	2.2 ± 0.1 (qNMR after aquaregia digestion)	50
H ₂ N-PEG-SH (average Mw=5000 Da)	PVP-AuNCs (30 nm in edge length)	0.61 (Fluorescamine-based assay, total PEGs) 0.19 (FITC-labeling assay, active PEGs)	52
	PVP-AuNCs (50 nm in edge length)	0.85 (Fluorescamine-based assay, total PEGs)	52
	PVP-AuNCs (60 nm in edge length)	1.36 (Fluorescamine-based assay, total PEGs)	52

Ligand type	Gold surface	Ligand density / molecule per Nm^2 (method)	Ref.
	CTAC-AuNSs (40 nm)	0.41 (Fluorescamine-based assay, total PEGs) 0.22 (FITC-labeling assay, active PEGs)	52
	Cit-AuNSs (42 nm)	1.63 (Fluorescamine-based assay, total PEGs) 1.01 (FITC-labeling assay, active PEGs)	52
	CTAB-AuNRs (80 nm $\times$ 22 nm)	0.052 (Fluorescamine-based assay, total PEGs) 0.029 (FITC-labeling assay, active PEGs)	52
Thioctic acid	AuNSs (12.3 $\pm$ 1.2 nm)	2.2 $\pm$ 0.1 (XPS)	49
Mercaptobenzimidazole	AuNSs (13 nm)	3.4 (Ratiometric SERS)	53
Oligonucleotides-SH	AuNSs (10 nm to 200 nm)	0.06 to 0.21 (Oligreen assay after CN^- digestion)	54