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Sequence-Controlled Polymers Through Entropy-Driven Ring-Opening Metathesis Polymerization: Theory, Molecular Weight Control, and Monomer Design

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Abstract

The bulk properties of a copolymer are directly affected by monomer sequence, yet efficient, scalable, and controllable syntheses of sequenced copolymers remain a defining challenge in polymer science. We have previously demonstrated, using polymers prepared by a step-growth synthesis, that hydrolytic degradation of poly(lactic-*co*-glycolic acid)s is dramatically affected by sequence. While much was learned, the step-growth mechanism gave no molecular weight control, unpredictable yields, and meager scalability. Herein, we describe the synthesis of closely-related sequenced polyesters prepared by entropy-driven ring-opening metathesis polymerization (ED-ROMP) of strainless macromonomers with imbedded monomer sequences of lactic, glycolic, 6-hydroxy hexanoic, and syringic acids. The incorporation of ethylene glycol and metathesis linkers facilitated synthesis and provided the olefin functionality needed for ED-ROMP. Ring-closing to prepare the cyclic macromonomers was demonstrated using both ring-closing metathesis and macrolactonization reactions. Polymerization produced macromolecules with controlled molecular weights on a multigram scale. To further enhance molecular weight control, the macromonomers were prepared with *cis*-olefins in the metathesis-active segment. Under these selectivity-enhanced (SEED-ROMP) conditions, first-order kinetics and narrow dispersities were observed and the effect of catalyst initiation rate on the polymerization was investigated. Enhanced living character was further demonstrated through the preparation of block copolymers. Computational analysis suggested that the enhanced polymerization kinetics were due to the *cis*-macrocyclic olefin being less flexible and having a larger population of metathesis-reactive conformers. Although used for polyesters in this investigation, SEED-ROMP represents a general method for incorporation of sequenced segments into molecular weight-controlled polymers.

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Supporting Information. The Supporting Information including complete experimental details, spectral data, and supplementary figures is available free of charge on the ACS Publications Website.

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Graphical Abstract

Keywords

entropy-driven; sequenced-controlled polymers; polyester; selective metathesis

Introduction

There exists a clear demand for the development of general, scalable, and controllable synthetic methods for imbedding monomer sequence in synthetic copolymers both to enable the production of novel materials for applications and also to define more clearly how sequence controls properties. The effects of monomer sequence on polymer properties are well-established in Nature, wherein precise monomer order can be directly mapped to macromolecular function. In synthetic polymer chemistry, in contrast, sequence control and the derivation of structure/function relationships has been limited to a variety of more easily accessible motifs including alternating, gradient, or blocky structures.¹⁻³ The attainment of a more detailed understanding has been inhibited by the challenges inherent in preparing precisely sequenced copolymers with molecular weight control. In the few classes of materials for which data have been obtained, the majority of the work has focused on solution phase properties; the most studied systems include peptoids,^{4,5} foldamers,⁶⁻⁸ and systems exploiting molecular recognition moieties.⁹⁻¹¹ Bulk-phase structure/function studies, which typically require gram-scale quantities,¹² are extremely rare.^{4,13-23}

Our group has long been interested in understanding the connections between sequence and properties in non-biological polymers. In the course of that work we have developed synthetic routes to periodically sequenced poly(lactic-*co*-glycolic acid)s (PLGA)s and established that bulk phase behaviors during hydrolysis are extremely dependent on constituent monomer sequence.²⁴⁻²⁶ Our interest in this class of polymers and their behavior arose because of their potential for application as degradable sutures, drug delivery vehicles, and cell scaffolds.²⁷⁻³² The random version of this class of polymer, which is widely used and FDA-approved, exhibits a degradation profile that is controlled primarily by molecular weight and the ratio of lactic (L) to glycolic acid (G) monomers.²⁸ Our work on periodic PLGAs demonstrated that monomer sequence is an even more powerful tool for controlling

hydrolysis behavior, facilitating not only rate control, but also affecting the retention of mechanical properties, swelling, internal pH, lactic acid release, and release of guest molecules *in vitro*.²⁴⁻²⁶

Despite our successes in carrying out structure/function studies on sequenced PLGAs, the scalable and controllable syntheses of these polymers remained an ongoing challenge. Our original synthetic efforts relied on materials prepared by a step-growth synthesis in which hydroxy-acid sequenced oligomers (segmers) were first prepared and then polymerized. Using this segmer-assembly polymerization (SAP) method, we were able to prepare a large range of periodic copolymers. Although moderately scalable, SAP provided no molecular weight control, unpredictable yields, poor reproducibility, and relatively large dispersities ($D = 1.5 - 1.8$). These issues limited the accuracy and scope of structure/function studies for these materials.

To address these synthetic limitations, we sought to develop a general method of preparing sequence-controlled polymers that proceeded, at least initially, via a chain-mechanism pathway. Specifically, we targeted entropy-driven ring-opening metathesis polymerization (ED-ROMP), which has been used previously to polymerize cyclic macromonomers.³³⁻⁴² We sought to prepare sequenced macrocyclic oligomers (MCOs) containing LG segments, a “linker” monomer to connect these segments that would not dominate bulk properties,⁴³ and an olefin-metathesis site (Scheme 1). This new polymerization approach was expected to improve molecular weight control, decrease dispersity, and achieve higher molecular weights than those obtained by the step-growth SAP methodology. In previous communications,^{44,45} we reported our initial successes using this approach. Herein, we describe in detail how ED-ROMP functions in this system and how we were able to introduce living character into the polymerization through a simple *trans*- to *cis*-olefin substitution.

ED-ROMP has been used previously to polymerize strainless macrocycles, typically containing greater than 14 atoms, with entropy serving as the primary driving force (Figure 1).³³ High concentration is used to favor polymer chains over cyclic species. The molecular weight and dispersity observed at any timepoint depend on the rate of propagation (k_{pr}) relative to that of secondary metathesis (k_{sm}).⁴⁶⁻⁴⁸ It should be noted that termination reactions are not typically observed in these systems on the timescale of a normal polymerization. Final molecular weights and dispersities are a function of monomer-to-catalyst loading ($[M]/[cat]$), ring-chain equilibrium, and overall concentration.^{33, 49} ED-ROMP typically results in dispersities greater than 1.1 due to competing secondary metathesis/backbiting (chain-transfer) reactions.

Although unmodified ED-ROMP does allow for molecular weight control, the prevalence of secondary metatheses of the newly produced olefin backbone interferes with the ideal behavior that would be expected for a system in which k_{pr} is much greater than k_{sm} . Such a system would be expected to behave in a more “living” fashion, giving chains that grow steadily and similarly in molecular weight.

Knowing that these rates depend on the steric accessibility of the olefin,⁵⁰ we have chosen to investigate, both theoretically and experimentally, the effects of the olefin geometry on mechanism and living character. Our findings culminate in the description of a new variant that we term Selectivity-Enhanced ED-ROMP, or SEED-ROMP. The MCOs used in these studies consist of strainless rings equipped with metathesis-active olefin linkers (**M**). Each MCO also incorporates sequences of lactic (**L**), glycolic (**G**), 6-hydroxy hexanoic (**C**), and syringic (**Sy**) acids as well an ethylene glycol (**Eg**) unit for palindromic monomers (Scheme 2). For some MCOs the polymerizations are studied as a function the cis/trans content of the metathesis linker. The differential behavior of these species is also investigated computationally with the goal of understanding the increased living character observed for the cis SEED-ROMP monomers.

Results and Discussion

Synthesis

Detailed information on the synthesis of MCOs and polymerization conditions is provided in the Supporting Information. L-lactic acid is the isomer used throughout. The synthesis of MCOs in this work are derived from protocols previously reported by our group for the production of sequenced oligomers of α -hydroxy acids.^{24-26, 51} A typical synthesis begins with orthogonal protections of monomers—acid moieties with benzyl (Bn) groups and alcohols with tert-butyldiphenylsilyl (TBDPS) groups. Ester couplings with dicyclohexylcarbodiimide (DCC) and nucleophilic amine catalyst DPTS provide di-protected hydroxy-acids which are named from the acid-terminus to alcohol-terminus. Removal of the Bn group via hydrogenolysis provides free acids and removal of the TBDPS group with TBAF provides free alcohols that are then available for sequential cycles of couplings and deprotection. Under these reaction conditions little or no transesterification is observed.

MCOs *trans*-**Eg(LGLM)**₂ and *trans*-**Eg(LLCM)**₂ were prepared by symmetric coupling of trimeric sequences of the specified hydroxy-acids to **Eg**. The resulting diols were coupled to 4-pentenoic acid to give an acyclic diene. Ring-closing metathesis (RCM) with Grubbs second-generation catalyst (**G2**) provided the MCO products in excellent yield, with ~90% *trans*-selectivity. These oligomers may also be ring-closed in the presence of Grubbs Z-selective nitrato catalyst (**GN**)⁵² with the exact opposite selectivity. MCO *cis*-**Eg(LGLM)**₂ was obtained in this fashion. The RCM reactions are highly reproducible but require extremely low concentrations (0.001 M) and 10-15 mol% catalyst loadings.⁵³

Although the palindromic monomers were moderately scalable, we were able both to improve scalability and remove the **Eg** unit by de-symmetrizing the open-chain oligomer and incorporating the olefin as an intact unit. This method, which exploits macrolactonization⁵⁴ to achieve the ring-closing, benefits from eliminating the large ruthenium catalyst loadings required for RCM and can be used to obtain similar yields using 30x less solvent.

Additionally, the nonsymmetrical monomers provide linkage directionality more closely resembling virgin PLGAs with acid and alcohol termini. In this fashion, hydroxy-acid

sequenced precursors **LMLGLGLG** and **SyLMLGLGL** were ring-closed at low concentration at 60 °C in dichloroethane charged with DCC and DPTS. Slow addition of the oligomer over the course of 16 h provided the desired product in yields >85%.

Polymerization of these MCOs produced polymers with a conservation of order within each MCO-derived segment but head-tail disorder at the olefin connector as evidenced by ¹H NMR spectroscopy (Figure 2A).

All ED-ROMPs or SEED-ROMPs were performed at 0.7 M at RT with **G2** catalyst loadings ranging from 1-5 mol% resulting in 20 – 80 kDa across the four MCOs (Table 1).

As the residual ruthenium remaining in the polymers is a common problem for products of metathesis whose primary application is in the field of regenerative medicine, polymers were purified using a combination of reprecipitation and exposure to a thiol-functionalized resin.^{55, 56} Using this method, concentrations of ruthenium in the polymer matrix was reduced from 1200 ppm to 34 ppm, approaching acceptable levels for most biological applications.^{57, 58}

Syringic Acid Incorporation to Elevate T_g

As the ED-ROMP of **M**-containing MCOs produced polymers with T_gs of 18 - 20 °C, which are lower than that required for most biomedical applications (~ 40 - 50 °C), we exploited the modular nature of the synthetic method to incorporate a biocompatible monomer that would elevate T_g. We were inspired by the Miller group to include syringic acid (**Sy**), a phenolic acid antioxidant found in grains and plant cell walls.^{43, 59} Thus, the segment **SyLMLGLGL** was ring-closed via lactonization, and polymerized to prepare **Poly SyLMLGLGL** with M_ns up to 45 kDa. The polymer exhibited the targeted increase in T_g (50 °C).

Sequence Characterization

The exquisite sensitivity of the diastereotopic **G** methylene protons provide a fingerprint of sequence in ¹H NMR spectra (Figure 2). We have previously determined that in some instances the chemical shifts of these protons are affected by sequence differences up to six monomer units away, and as little as 2% sequence error can be easily resolved.⁶⁰ As such, we are able to clearly characterize and differentiate copolymers with a range of structures. In addition to general characterization and confirmation of sequence, this sensitivity also makes it possible to monitor quantitatively the progress of the polymerization reaction as a function of time and isomer.

The palindromic MCOs provided the simplest resonances. Pairs of doublets in which *cis*- and *trans*-olefin composition can be monitored allowed us to monitor RCM conformational selectivity and polymerization kinetics (Figure 3). The unsymmetric MCOs obtained via macrolactonization displayed more complex methylene resonances, yet ring-opened chains could still be fully distinguished from monomer (Figure 2).

Neither *trans*-**LM_{H-T}LGLGLG** nor *trans*-**SyLM_{H-T}LGLGL** polymerized regioselectively. Thus, a statistical distribution of head-tail connectivities of 50% head-tail M_{H-T}, 25% head-head M_{H-H}, and 25% tail-tail M_{T-T} resulted, as evidenced by the distinct ¹H NMR olefin

signals (Figure 2A). This analysis confirms that the selected **M** linker is long enough to eliminate local electronic effects of constituent monomers. We hypothesize that the selectivity could be improved by using a tri-substituted olefin although those experiments were not performed in this study.^{38, 61-65} The conversion of *trans*-**SyLMLGLGL** to **Poly SyLMLGLGL** can be similarly followed spectroscopically (Figure 2B).

MALDI-TOF Sequence Characterization

In addition to NMR spectroscopy as a sequence characterization tool, we have previously demonstrated the power of MALDI-TOF mass spectrometry to characterize the fidelity of sequenced polyesters, even in cases where only the most volatile shorter chains in the distribution are observed.⁶⁶ In the currently reported ED-ROMP copolymers, a similar analysis consists primarily of chains whose molecular weights are multiples of the MCO masses (Figure 2). Peaks for sequence errors that either add, omit or substitute monomers within repeat units are weak or not observed.

Cis- vs. trans-Olefin Studies

Our original MCO *trans*-**Eg(LGLM)₂**, prepared via RCM with **G2**, consisted of 85-90% *trans*-isomer. Polymerization kinetics were monitored by ¹H NMR spectroscopy. *Trans*-olefin content in the resulting polymer remained at ~85% at all times during the polymerization. Molecular weight stabilized in approximately 30 min with dispersities of ~1.3 – 1.5. Molecular weights were higher than those achieved using SAP (25-45 kDa vs. 10-20 kDa) and were reproducible. The degree of polymerization (DP) deviated, however, from that predicted from catalyst loading.

We sought to enhance the polymerization to approach a more living system by incorporating the less stable and potentially more reactive *cis*-olefin. The MCO, *cis*-**Eg(LGLM)₂**, was prepared by Z-selective metathesis.⁴⁵ The enhanced selectivity of the catalyst for *cis*-MCO leads to more living polymerization character, as highlighted in Figure 3. Dispersities of < 1.1 were maintained throughout the reaction and consistently higher molecular weights were obtained. Conversion of MCO followed linear first-order kinetics, assuming saturation conditions,⁶⁷ and >90% monomer conversion was attained in only 10 minutes. Over the course of the reaction, the 85-90% *cis*-olefin content was converted to ~85% *trans* in the resulting polymer. Moreover, we observed that the *cis*-olefin was consumed preferentially both under the polymerization conditions and in a control experiment in which equal mixtures of both isomers were exposed to catalyst (Figure 3B, Figure S1). Although the faster reaction of the *cis*-isomer was not surprising based on earlier reports of *cis/trans* metathesis behavior,⁶⁸ the true source of this preferential reactivity was not clear. To address this question, we examined the system computationally (*vide infra*).

Molecular weight control studies

Varying catalyst (**G2**) loading across all monomers resulted in at least a moderate degree of molecular weight control, yet the extent varied with olefin geometry and monomer composition (Figure 4). *Cis*-olefin-containing MCOs displayed a near-living linear molecular weight control ranging from 35 – 70 kDa while *trans*-MCOs tracked with [M]/[cat] at high catalyst loadings but saturated with low loadings. There was also some evidence

for composition dependence. **Trans-Eg(LLCM)₂** exhibited the least linear degree of control plateauing at ~55 kDa while the unsymmetric **trans-LMLGLGLG** displayed more control before saturation at 50 kDa. Although temperature would be expected to be an important factor in this entropy-controlled process,⁶⁹ cooling the reaction (0 °C) did appear to affect molecular weight or dispersity at the relatively high concentrations being used for these experiments.

The low dispersities observed for the *cis*-MCOs suggest that we have successfully favored propagation over secondary metathesis. We believe that the slightly larger *Đ*'s observed at high catalyst loadings of *cis*-MCOs are due primarily to incomplete initiation in these extremely concentrated, and therefore, rapid polymerization conditions.⁷⁰ Interestingly, dispersities remain below the theoretically predicted 2 for even *trans*-olefin MCOs which suggests that these systems are not achieving equilibrium before they are quenched. We are not certain why this is the case as reactions run at longer reaction times did not result in higher dispersities. Experimentally, it may be that the catalyst deteriorates before equilibrium is achieved or that the bond selectivity of the secondary metathesis reaction is not fully random. Functionally, the limit on dispersity may also be due to the relatively low *DP*s of these large MCOs.

Chain and Block Extensions via SEED-ROMP

As we were interested in determining the degree to which these polymerizations could be considered living, we carried out both chain extension experiments and block copolymerizations with **cis-Eg(LGLM)₂** (Figure 5). For chain extension, an initial polymerization with a fixed amount of MCO was followed by a second addition of the same MCO. To prepare block copolymers the initial polymerization of the MCO was followed by the addition of a traditional strained ROMP monomer, *cis*-cyclooctene (COE) or norbornene (NBE).⁷¹⁻⁷⁴

The chain extension produced a polymer of higher molecular weight which maintained a *Đ* <1.1. When the experiment was undertaken with NBE, block copolymers ranging from 60-90 kDa were obtained. These polymers, however, exhibited significant secondary metathesis resulting in unpredictable final molecular weights and considerable increases in dispersities (Figure 5B). Attempts to control the block formation by controlling reaction time and temperature resulted in only small improvements (Figure S3). Each time block addition was repeated, a clear increase in molecular weight with minimal increase in dispersity was observed within 1 min, but molecular weight quickly decreased and broadened in dispersity thereafter (Figure S4). However, ¹H NMR spectra remained clean in the **G** methylene region, indicating that the polyester block did not suffer from significant secondary metathesis and the uncontrollable molecular weights originated from the highly reactive norbornene block. It is worth mentioning that there exist copious variants of functionalized norbornenes that offer more control,⁷⁵⁻⁷⁷ but these derivatives were not investigated in this current proof of principle study. The significant increases in dispersities and small increases in molecular weights (Figure 5b, S3) could also indicate that a significant portion of chain ends were deactivated and unable to undergo block extension. To address this possibility, the experiment was repeated with COE. In this case, block addition

was achieved with no increase in dispersity and molecular weights of ~70 kDa (Figure 5C). This result is consistent with the conservation of a high percentage of active chain ends. As the process would not be expected to be purely living, however, some degree of chain transfer is expected.

Although not a subject of the current work, it is worth noting that ROMP monomers such as NBE and COE may be functionalized to potentially attach deliverable payloads into biological systems for controlled release, or initiate micellar self-assembly.^{72, 78, 79} SEED-ROMP is therefore an exceptionally powerful tool in engineering materials for controlled and precise function.

Catalyst Initiation Effects

Switching catalyst from the **G2** catalyst to the Grubbs 3rd generation catalyst (**G3**), which is known to exhibit faster initiation,⁸⁰⁻⁸² did not significantly affect the kinetics, molecular weights, or dispersities (Figure S2). Unsurprisingly, the more reactive catalyst proved less robust to contaminants.

Computational Studies

As the enhanced reactivity of the *cis*- relative to the *trans*-MCO is the basis for SEED-ROMP and key to the near-living nature of the polymerization, we performed computational studies to investigate the origins of the difference. A few factors are known to affect the kinetic reactivity of ROMP, including ring strain,⁸³⁻⁸⁶ sterics, and electronic effects.⁵⁰ However, both the *cis*- and *trans*-MCOs are expected to have very small ring strain energies, and, thus, the higher reactivity of the *cis*-isomer should not be promoted by ring-strain. In addition, the electronic and steric properties of *cis*- and *trans*-double bonds in the MCOs are similar. Based on the known kinetic reactivity of acyclic *cis*- and *trans*-olefins in cross-metathesis and ethenolysis,^{68, 87, 88} the local steric and electronic environments of the *cis*- and *trans*-MCOs are not expected to play a key role in differentiating their reactivities in ROMP. We surmised the conformational flexibility of the macrocyclic monomers might affect the ability of the C=C double bond to approach the Ru catalyst, which then could, in turn, affect the reactivity of *cis*- and *trans*-MCOs. Due to the flexible nature of the macrocyclic monomers, a large number of conformers exist in the resting state and many of these conformers are expected to provide access to the [2+2] cycloaddition transition state. Thus, a complete understanding of the relative reactivity of *cis*- versus *trans*-MCOs requires the evaluation of all ground state conformers and their relative reactivity in the [2+2] cycloaddition transition state.

As the large number of transition state conformers is highly challenging for conventional transition state calculations, we utilized a combined molecular dynamics/density functional theory (MD-DFT) approach to investigate the conformational flexibility of the monomers using MD trajectory simulations combined with evaluation of reactivities of each monomer conformation using DFT.

In our preliminary study,⁴⁵ we compared the conformations of the *cis*- and *trans*-MCOs from snapshots of MD simulations with the DFT-optimized transition state geometries that used

cis- and *trans*-3-hexene as model substrates.⁴⁵ It was revealed that the geometries of the *cis*-MCOs have smaller deviations from the reactive olefin conformations in the metathesis transition state, indicating greater populations of metathesis-active monomers, than the *trans*-MCO. To provide a more quantitative understanding of how monomer conformation affects the metathesis reactivity, we created a model that estimates the activation energy required for each MCO conformer to undergo [2+2] cycloaddition with the 14-electron Ru alkylidene complex derived from the **G2** catalyst.

First, we performed 20 ns MD simulations of *cis*- and *trans*-Eg(LGLM)₂ in a dichloroethane solvent box. Snapshots of the MD simulations revealed great levels of conformational flexibility in the two dihedral angles (θ and ϕ) for the C-C sigma bonds attached to the alkene. These conformers are expected to exhibit dramatically different reactivity in the [2+2] cycloaddition. Conformers with the two olefin substituents (highlighted in green in Figure 6A) placed on the same face of the double bond have exposed C=C double bonds (e.g. **cis-1**), which are expected to be more reactive with the catalyst. If one of the substituents is placed in the same plane of the double bond, an increased steric repulsion with the Ru catalyst is expected, and, thus, the reactivity should be lower (e.g. **cis-2**). For conformers with two substituents on opposite faces, the C=C double bond is blocked for both, which translates into the lowest reactivity with **G2** (e.g. **cis-3**). Similarly, the dihedral angles (θ and ϕ) are also expected to affect the reactivity of the conformers of the *trans*-MCO.

To quantify the effects of the two highlighted dihedral angles on the reactivity of the conformers, we established an activation energy contour by correlating the DFT-calculated activation energies of constrained [2+2] cycloaddition transition states ($E^\ddagger$) with two varying but fixed dihedral angles (θ and ϕ) for a model system of constrained *cis*- and *trans*-alkenes. The activation energy contours were calculated using a model system of constrained *cis*- and *trans*-3-hexenes and a 14-electron Ru alkylidene complex derived from the **G2** catalyst (Figure 6B). All constrained transition states were validated with the imaginary frequency vibration corresponding with the [2+2] cycloaddition process, and their activation energies were color-encoded where the blue indicated the more reactive region while the yellow to red indicated the less reactive region (Figure 6B, right).

We next mapped the *cis*- and *trans*-MCO conformers from the MD snapshots onto the generated activation energy contour. The activation energy for each conformer was thereby estimated with respect to two corresponding dihedral angles via linear interpolation from the contour. As shown in Figure 6B, the conformers of *cis*-MCO were primarily localized on the most reactive regions (blue, $E^\ddagger < 0$ kcal/mol) such as **cis-1** with the estimated activation energy of -2.7 kcal/mol. Only a relatively small fraction of *cis*-conformers were in the less reactive region, colored cyan-to-yellow. For example, **cis-2** with one dihedral angle close to 0, which makes the top face of the double bond more sterically hindered, has a higher activation energy of 1.5 kcal/mol.

In contrast, *trans*-MCOs were more flexible. The conformers are more dispersed throughout the contour map and have a greater distribution in less reactive regions than the *cis*-MCOs. For instance, **trans-1**, one of the most reactive conformers ($E^\ddagger = -2.3$ kcal/mol), has the

double bond fully exposed to Ru catalyst from the top face. **Trans-2** is less reactive ($E_{\ddagger} = 4.3$ kcal/mol), however, with two olefin substitutes placed on the plane of the double bond. This arrangement increases the steric repulsion between the substitutes and the Ru catalyst in their transition state.

Our computational studies suggest that the higher reactivity of *cis*-MCOs can be attributed to their more localized conformer ensembles with the majority being reactive conformations for [2+2] cycloaddition in ROMP. It is interesting to note that this finding is subtly different from the intuitive rationale that *cis*-olefins react faster because they are “more accessible” and can more easily approach the metal center. This reasoning is based on a model of the olefin in which the substituents present a singular steric profile rather than a population of profiles and does not take into account the fact that not all coordination modes lead to energetically favorable transition states.

Conclusions

We have successfully designed a general, modular, and scalable synthetic method for the preparation of copolymers in which sequence is imbedded into a large macrocycle which is then polymerized using ED-ROMP. SEED-ROMP, which exploits a more reactive *cis*-olefin metathesis handle, enhanced the living character of the polymerization, improving molecular weight control, decreasing dispersity, and facilitating the preparation of block copolymers. Computational analysis demonstrated that *cis*-olefin-containing monomers promote propagation over secondary metathesis by reacting faster than the *trans*-isomers and that the preference arises from the fact that conformationally rigid *cis*-macromonomers present a higher population of reaction-favoring binding modes.

We have also established that ring-closing via macrolactonization can be carried out at higher concentrations and less expensively than RCM-based ring-closure in the preparation of the cyclic MCOs. Moreover, the precursor oligomers required for this method do not require an additional linker group (**Eg**) in their construction, meaning that the final polymers more closely resemble the ideal structure for a poly(α -hydroxy acid).

The development of sequence-controlled polymer syntheses is an ongoing effort, in which homogeneity across samples in molecular weight and dispersity is critical. The current approach, which couples experimental intuition with theory, has expanded both the range of materials that can be prepared and our ability to control their characteristics. It seems likely, moreover, that this general approach will be applicable to polymers other than poly(α -hydroxy acid)s; all that is required is that target sequence be incorporated into an MCO with a metathesis-active olefin. With such methods it should be possible to better target sequenced materials for future applications.

Experimental Section

Materials.

All experiments were carried out in oven-dried glassware under an atmosphere of N₂ using standard Schlenk line techniques. N,N'-dicyclohexylcarbodiimide (DCC) was purchased from Oakwood Chemical and used without further purification. 1,2-dichloroethane (DCE)

was purchased from Fisher and used as is. Methylene chloride (CH_2Cl_2 , Fisher), ethyl acetate (EtOAc, Sigma-Aldrich) and tetrahydrofuran (THF, Fisher) were purified by passage over neutral activated alumina. The reagents 4-(dimethylamino)pyridinium 4-toluenesulfonate (DPTS), silyl (Si-R_3 , tert-butyldiphenylsilyl) and benzyl (Bn) protected monomers, unprotected monomers, and polymers were prepared according to previously published protocols.

^1H NMR Spectroscopy.

^1H (400 and 500 MHz) NMR spectra were recorded using Bruker spectrometers in CDCl_3 and calibrated to the solvent peak of δ 7.26 ppm.

Size Exclusion Chromatography.

Molecular weights and dispersities were obtained on: 1) Waters GPC (THF) with Jordi 500, 1000, and 10000 Å divinylbenzene columns, and refractive index detector (Waters) was calibrated to polystyrene standards 2) TOSOH HLC-8320GPC EcoSEC equipped with two columns (TSK-3000H, TSK-4000H). A mobile phase of THF inhibited with 0.025% butylated hydroxytoluene at 40 °C was used, reported molecular weights are relative to polystyrene standards.

Differential Scanning Calorimetry

Differential scanning calorimetry was performed with TA Instruments Q200. Each run was performed at 10 °C/min heating and cooling rates. T_g 's were recorded in the second heating cycle.

MALDI-of MS.

MALDI-TOF MS spectra were obtained on a Bruker ultrafleXtreme MALDI-ToF instrument. An accelerating voltage of 20 kV was applied, and spectra were obtained in reflection mode (500 shots). The polymers were dissolved in THF to yield a concentration of 1 mg/mL. Potassium trifluoro-acetate (KTFA) was used as the cationization agent and was dissolved in THF to form a 1 mg/mL solution. The matrix was DCTB in THF as a 40 mg/mL solution. The three solutions were combined in a ratio of 1:1:1.5 (polymer: DCTB: KTFA) and allowed to mix for 1 h. The solution was then drop cast onto a 100-well MALDI plate and allowed to dry before analysis. Spectra were obtained using Bruker flexAnalysis software package.

Trans-selective RCM of Eg(LGLM)_2 – Representative Procedure

A solution of **G2** (3.9 mg, 0.0046 mmol) in CH_2Cl_2 (1 mL) was added to a stirring solution of **Eg(LGLM) $_2$** (28.0 mg, 0.044 mmol) in CH_2Cl_2 (42 mL) and the reaction was stirred at rt overnight. The reaction was quenched by adding ethyl vinyl ether (1 mL), and the reaction was concentrated *in vacuo*. The crude material was purified by flash chromatography (SiO_2 , 20-30% EtOAc in hexanes) to provide products as colorless liquids with E:Z ratios near 85:15 (24.1 mg, 89.9%) ^1H NMR (400 MHz, CDCl_3) δ 5.44-5.23 (*trans*) and 5.39-5.38 (*cis*) (m, 2H), 5.23 (m, 4H), 4.79 (d, J = 16.0 Hz, 2H, *trans*), 4.77 (d, J = 16.0 Hz, 2H, *cis*), 4.38 (m, 4 H), 2.44 (m, 8H), 1.52 (d, J = 7.0 Hz, 6H), 1.50 (d, J = 7.0 Hz, 6H); ^{13}C NMR (100

MHz, CDCl₃) δ 172.27, 172.22, 170.19, 169.76, 166.61, 129.32 (*trans*), 129.12 (*cis*), 69.35, 68.19, 62.71, 60.86, 33.62, 27.56, 27.34, 22.75, 16.85, 16.69, 16.65; HRMS (ESI) calc. mass 603.1920, found 603.19028.

Cis-selective RCM of **Eg(LGLM)₂**

In the glovebox, a solution of **GN** (69 mg, 0.1094 mmol) in DCE (20 mL) was added to a stirring solution of **Eg(LGLM)₂** (690 mg, 1.094 mmol) in DCE (200 mL). The vessel was immediately removed from the glovebox and stirred at 60 °C under a constant low vacuum. After 26 h of stirring, the reaction solution was cooled to rt, ethyl vinyl ether (1 mL) was added, and the solution was concentrated. The crude product was purified by flash chromatography (SiO₂, 20-30% EtOAc in hexanes) to afford *cis*-**Eg(LGLM)₂** (0.578 g, 88% yield, 95% BRSM, 12:88 *E:Z*) as a colorless oil; ¹H NMR (500 MHz, CDCl₃) δ 5.51 (m, *trans*) and 5.43 (m, *cis*)(2H), 5.23 (m, 4H), 4.83 (d, *J* = 16.0 Hz, 2H, *trans*), 4.81 (d, *J* = 16.0 Hz, 2H, *cis*), 4.40 (m, 4 H), 2.48 (m, 8H), 1.55 (d, *J* = 7.0 Hz, 6H), 1.53 (d, *J* = 7.0 Hz, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 172.44, 170.32, 169.92, 166.77, 129.47 (*trans*), 129.12 (*cis*), 69.52, 68.34, 62.81, 60.96, 33.96, 22.91, 16.94, 16.80; HRMS (ESI) calc. mass 603.1920, found 603.1940.

Macrolactonization – Representative Procedure

Using a syringe pump, a solution of **SyLMLGLGL** (1.61 g, 2.25 mmol in 25 mL DCE) was injected into a 60 °C, 225 mL solution of DCE containing DCC (0.510 g, 2.47 mmol) and DPTS (0.132 g, 0.450 mmol) over the span of 16 h. The solution was stirred for an additional 8 h before cooling to rt, filtration, and concentrating *in vacuo* to obtain the crude product. The crude oil was purified by flash chromatography (SiO₂, 20-30% EtOAc in hexanes) to produce the product as a white solid. ; ¹H NMR (500 MHz, CDCl₃) δ 7.36 (s, 2H), 5.50 (q, *J* = 7.2 Hz, 1H), 5.46 (q, *J* = 7.2 Hz, 1H), 5.22 (q, *J* = 7.2 Hz, 1H), 5.03 (q, *J* = 7.2 Hz, 1H), 4.85 (d, *J* = 16 Hz, 1H), 4.72 (d, *J* = 16 Hz, 1H), 4.68 (d, *J* = 16 Hz, 1H), 4.63 (d, *J* = 16 Hz, 1H), 4.11 (t, *J* = 6.8 Hz, 2H), 3.85 (s, 6H), 3.16 (m, 2H), 2.35 (q, *J* = 6.8, 2H), 1.67 (m, 6H), 1.49 (d, *J* = 7.2 Hz, 3H), 1.36 (d, *J* = 7.2 Hz, 3H) ¹³C NMR (125 MHz, CDCl₃) δ 175.65, 170.06, 169.82, 169.25, 167.90, 166.30, 166.19, 165.08, 152.09, 132.38, 129.44, 127.73, 124.51, 106.65, 69.57, 69.33, 69.07, 68.50, 64.32, 60.97, 60.42, 56.43, 37.77, 31.69, 17.02, 16.91, 16.66, 16.58; HRMS (ESI) calc. mass 696.1880, found 696.18686.

Polymerization with **G2** – Representative Procedure **G2**

(10.5 mg, 0.012 mmol) was dissolved in CH₂Cl₂ (0.3 mL) and added via syringe to a stirring solution of *trans*-**Eg(LGLM)₂** (0.57 g, 0.94 mmol) in CH₂Cl₂ (1.0 mL). The reaction was stirred at rt for 4 h before ethyl vinyl ether was added (0.2 mL) to quench. The reaction mixture was dissolved in a minimal amount of CH₂Cl₂ and precipitated into 250 mL swirling MeOH. The off-white solid was isolated (74.8%). ¹H NMR (500 MHz, CDCl₃) δ 5.49 (m, 1.7 H; *trans*), 5.42 (m, 0.3H, *cis*), 5.20 (q, *J* = 7.5 Hz, 2H), 5.18 (q, *J* = 7.0 Hz, 2H), 4.89 (d, *J* = 16.5 Hz, 2H), 4.65 (d, *J* = 16.5 Hz, 2H), 4.41 (m, 4H), 2.50 (m, 4H), 2.35 (m, 4H), 1.56 (d, *J* = 7.0 Hz, 6H), 1.53 (d, *J* = 7.5 Hz, 6H); ¹³C NMR (125 MHz, CDCl₃) δ

172.39, 170.27, 169.75, 166.66, 129.35, 69.27, 68.17, 62.79, 60.69, 33.69, 27.58, 16.88, 16.74; DSC: $T_g = 18\text{ }^{\circ}\text{C}$.

Molecular weight control studies

To investigate the degree of molecular weight control, polymers were prepared using the same conditions as described above, with varying mol% catalyst loadings to achieve the desired M/cat ratio. Solutions (1 mg/mL) of unpurified polymers were utilized for SEC analysis.

Kinetics Study

To a stirring solution of *cis*-**Eg-(LGLM)₂** (72 mg, 120 μmol) and CH_2Cl_2 (121 μL) was added a solution of catalyst **G2** (1.3 mg, 1.5 μmol) in CH_2Cl_2 (50 μL). Aliquots were removed via pipette at the specified time points and added to a vial containing a solution of ethyl vinyl ether to quench the catalyst. The *cis*-to-*trans* ratio of monomer was approximated by comparing peak integrations at 4.85-4.75 ppm. Total conversion was approximated by comparing peak integrations at 4.9-4.75 ppm.

SEED-ROMP Block Extensions – Representative Procedure

A solution of **G2** (0.18 mg, 0.218 μmol) in CH_2Cl_2 (10 μL) was added to a pre-mixed solution of *cis*-**Eg(LGLM)₂** (10.5 mg, 17.4 μmol) in CH_2Cl_2 (15 μL) and allowed to shake for 10 min before a pre-mixed solution of NBE (41 mg, 435 μmol) in CH_2Cl_2 (25 μL) was added at room temperature. The vial was shaken for 1 min, and EVE was added and was shaken for an additional 10 min. The mixture was diluted to a pre-weighed vial after diluting with CH_2Cl_2 (0.3 mL) and concentrated *in vacuo* to provide crude **poly Eg(LGLM)₂-b-NBE** as a solid (18.5 mg, 93% SEED-ROMP monomer conversion, 64% *cis*-olefin incorporation in **poly(NBE)**; based on DP, composition is 46 mol% block A (**poly Eg(LGLM)₂**, DP 100) and 54 mol% block B (**polyNBE**, DP 117). To add clarity to integration numbers presented herein, a 50:50 ratio of A:B has been assigned, and a 50:50 *cis:trans* ratio has been assigned for the NBE block. ^1H NMR (500 MHz, CDCl_3) δ 5.51 (m) and 1.42 (m) (2H), 5.35 (br s, 1H) and 5.21 (m, 5H), 4.89 (d, $J = 16.0\text{ Hz}$, 2H), 4.65 (d, $J = 16.5\text{ Hz}$, 2H), 4.41 (m, 4H), 2.85 (br s, 2H), 2.50 (m, 5H), 2.33 (m, 4H), 1.87 (m, 1H), 1.80 (m, 2H) 1.55 (d, $J = 6.0\text{ Hz}$, 6H), 1.53 (d, $J = 7.0\text{ Hz}$, 6H) 1.35 (m, 2H), 1.09 (m, 1H); ^{13}C NMR (125 MHz, CDCl_3) δ 172.49, 170.38, 169.86, 166.78, 134.14, 134.07, 134.03, 133.99, 133.90, 133.28, 133.15, 1300, 129.49, 129.154, 129.04, 69.41, 68.36, 68.30, 62.92, 60.83, 43.56, 43.27, 42.90, 42.24, 38.81, 38.56, 33.84, 33.25, 33.07, 32.52, 32.36, 27.72, 17.02, 16.88; SEC (THF): $M_n = 71.4\text{ kDa}$, $M_w = 78.8\text{ kDa}$, $\text{Đ} = 1.10$.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Figure 1.

Living character as a function of the relative rates of propagation (k_{pr}) and secondary metathesis (k_{sm}) in entropy-driven ring-opening metathesis polymerization.

Figure 2.

^1H NMR spectroscopy and MALDI-TOF mass spectrometry characterization of sequence fidelity. (A) ^1H NMR spectra of olefin and methylene resonances of *trans*-LMLGLGLG (top) and Poly LMLGLGLG (bottom), displaying head-tail olefin resonances, and MALDI-TOF spectrum of low molecular weight cyclic Poly LMLGLGLG. (B) ^1H NMR methylene resonances of *trans*-SyLMLGLGL (top) and Poly SyLMLGLGL (bottom); and MALDI-TOF spectrum of low molecular weight cyclic Poly SyLMLGLGL.

Figure 3.

Comparison of *trans*- versus *cis*-Eg(LGLM)₂ polymerizations. (A) SEC traces and molecular weight data for polymerizations with identical catalyst loadings. (B) Catalyst competition experiment with 1:1 *trans:cis* olefin mixture upon exposure to Grubbs second generation catalyst over time, 0.04 M. (C) First-order kinetic fits of *trans*- (both low and high conversions) and *cis*-Eg(LGLM)₂ assuming saturation conditions.

Figure 4. Molecular weights as a function of catalyst loading from palindromic monomers *trans*-Eg(LLCM)₂, *cis*-Eg(LGLM)₂, and unsymmetric *trans*-LM_{H-T}LGLGLG.

Figure 5. (A) ¹H NMR spectra for *cis*-Eg(LGLM)₂, polymerization to 50 kDa Poly Eg(LGLM)₂, and Poly Eg(LGLM)₂-*b*-COE. (x denotes CH₂Cl₂) (B) SEC traces of control and block extension using NBE (unpurified), displaying large dispersity due to secondary metathesis. (C) SEC traces of control and block extension with COE (unpurified).

Figure 6. Molecular dynamics-density functional theory (MD-DFT) calculations. A) Example conformations of *cis*- and *trans*-Eg(LGLM)₂. B) Energy mapping of macrocycle conformations onto activation energy contour.

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Scheme 2.

Ring-closing and polymerization reactions of sequenced segmers for controlled molecular weight. (A) Grubbs second generation (**G2**) and nitrate Grubbs (**GN**) catalyst structures. (B) Ring-closing metathesis and ensuing polymerizations of **Eg(LLCM)₂** and **Eg(LGLM)₂** segmers. (C) Selectivity-enhanced entropy-driven ring-opening metathesis polymerization to prepare **poly (EgLGLM)₂-b-COE**. (D) Macrolactonization reactions of **LMLGLGLG** and **SyLMLGLGL** and subsequent polymerizations.

Table 1.

Polymer Thermal Data and Molecular Weights

Polymer	M/cat	T _g (°C) ^a	M _n (kDa)	M _w (kDa)	Đ
Poly Eg(LLCM) ₂	20	-11	24 ^b	32 ^b	1.3 ^b
	45		42 ^b	53 ^b	1.3 ^b
	75		48 ^b	63 ^b	1.3 ^b
	125		60 ^b	78 ^b	1.3 ^b
Poly Eg(LGLM) ₂ ^d	78	18	33 ^c	44 ^c	1.3 ^c
	33		36 ^c	41 ^c	1.1 ^c
	80		56 ^c	58 ^c	1.0 ^c
	90		66 ^c	68 ^c	1.0 ^c
	100		70 ^c	71 ^c	1.0 ^c
Poly LMLGLGLG	33	20	26 ^c	34 ^c	1.5 ^c
	50		30 ^c	42 ^c	1.4 ^c
	57		33 ^c	47 ^c	1.4 ^c
	67		33 ^c	45 ^c	1.4 ^c
	80		42 ^c	55 ^c	1.3 ^c
	100		43 ^c	55 ^c	1.3 ^c
Poly SyLMLGLGL	80	50	36 ^c	44 ^c	1.2 ^c
	100		44 ^c	51 ^c	1.2 ^c

^aSecond heating cycle at 10 °C/min.^bSEC in THF, absolute molecular weight data.^cSEC in THF, relative to PS standards.^dPrepared by SEED-ROMP from *Cis*-Eg(LGLM)₂