Published in final edited form as:

Environ Monit Assess. ; 191(8): 500. doi:10.1007/s10661-019-7638-2.

Assessment of Placental Metal Levels in a South African Cohort

Laetitia Meyrueix¹, Linda Adair¹, Shane A Norris², Folami Ideraabdullah^{1,2,3,4,*}

¹Nutrition Department, University of North Carolina-Chapel Hill, Chapel Hill, North Carolina 27599, USA

²MRC Developmental Health Pathways for Health Research Unit, University of Witwatersrand, Johannesburg 2000, SA

³Nutrition Research Institute, University of North Carolina-Chapel Hill, Chapel Hill, North Carolina 27599, USA

⁴Genetics Department, University of North Carolina-Chapel Hill, Chapel Hill, North Carolina 27599, USA

Abstract

The placenta plays an important role in mediating the effect of maternal metals exposure on fetal development, acting as both barrier and transporter. Term-placenta metals levels serve as an informative snapshot of maternal/fetal exposure during pregnancy and could be used to predict offspring short- and long-term health outcomes. Here, we measured term-placenta metal levels of 11 metals in 42 placentas from the Soweto First 1000 days cohort (S1000, Soweto-Johannesburg, SA). We compared these placental metals concentrations to previously reported global cohort measurements to determine whether this cohort is at increased risk of exposure. Placental metals were tested for correlations to understand potential interactions between metals. Since these samples are from a birth cohort study, we also performed exploratory analyses to determine whether metals levels were associated with placenta and birth outcomes. Most S1000 placental metals levels were similar to other cohorts, however, cadmium (Cd) levels up to 50-fold lower, and essential elements nickel (Ni) and chromium (Cr) levels up to 6- and 16-fold lower, respectively. Cd, Se, and Ni were associated with placenta and birth outcomes. Studies are ongoing to examine underlying mechanisms and how these developmental differences affect long-term health.

*Corresponding Author: Folami Y. Ideraabdullah, Ph.D., folami@email.unc.edu, +1 919 445 9047; University of North Carolina at Chapel Hill, 120 Mason Farm Rd, CB# 7264, Chapel Hill, NC 27599, USA.

Author contributions: We declare that this study was conceptualized and designed by FI with input from LA and LM. Cohort recruitment and management, sample collection and selection supervised by SN. Sample preparation for metals measurements supervised by FI. Data QC and statistical analyses were performed by LM under FI supervision with input from LA. Manuscript was drafted and revised by LM and FI with input from LA, and SN. All authors have seen and approved the final version of the manuscript.

Compliance with Ethical Standards

Ethics approval: The work reported here was conducted in accordance with The Code of Ethics of the World Medical Association, approved by the Human Research Ethics Committee (Medical) at the University of the Witwatersrand, and reviewed and cleared by the Office of Human Research Ethics at the University of North Carolina at Chapel Hill.

Informed consent: Informed consent was obtained from all individual participants included in the study.

Availability of data and material: The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Declarations

Declaration of interest: none

Conflict of Interest: The authors declare that they have no conflict of interest.

Keywords

metals; placenta; pregnancy; birth outcomes; South Africa

Introduction

Metals naturally occur in the environment but can act as toxicants with adverse developmental effects, particularly neurodevelopmental and birthweight outcomes (Lawn et al. 2014; Wright and Baccarelli 2007; Parajuli et al. 2013; Luo et al. 2017; Thomas et al. 2015). Vanadium, cadmium, and lead were shown to be negatively associated with birthweight, while mercury and arsenic were associated with an increased risk for small for gestational age (SGA) (Bloom et al. 2014; Shirai et al. 2010; Hu et al. 2017; Sun et al. 2014; Xie et al. 2013). For some of these metals and others such as manganese, zinc, chromium, copper, and nickel, global industrialization in the last century has created an added burden on the environment (Tchounwou et al. 2012; Singh et al. 2011; He, Yang, and Stoffella 2005). However, even low exposure to metals is linked to adverse effects. For example, arsenic, lead, cadmium, and mercury can be hazardous at low exposure levels particularly during sensitive developmental windows or due to bioaccumulation (Tchounwou et al. 2012; Jaishankar et al. 2014; Wirth and Mijal 2010). Therefore, the role of metals as pollutants and their impact on public health is a growing concern.

Metals exposure is of particular concern during pregnancy since maternal exposure to harmful levels is linked to adverse fetal/birth outcomes (Luo et al. 2017; Shirai et al. 2010; Arbuckle et al. 2016). On the other hand, maternal deficiency of essential metals (e.g. zinc) has also been linked to adverse pregnancy outcomes and prenatal development resulting in SGA babies, intra-uterine growth retardation or reduced birth weight (Shah and Sachdev 2006; Keen et al. 1998; Mariath et al. 2011). However, a systematic review of the literature reported conflicting results between studies and further analyses are required to elucidate the full impact on human development (Chaffee and King 2012). The placenta likely plays an important role in mediating the effects of metals on fetal development since it serves as the interface between mother and child, supplying essential elements and acting as a barrier to harmful elements. Differences in harmful or essential metals levels could alter the capacity of the placenta to be an effective transporter and barrier through changes in placental formation, function, and pathology (Donnelly and Campling 2016; Saenz et al. 2013; Vaughan et al. 2011; Cooke 2014; Mattison 2010). Furthermore, differences in transplacental transport of metals to expose the fetus may directly impact fetal development. However, this remains poorly characterized for most metals (Myllynen, Pasanen, and Pelkonen 2005).

This is the first study to measure placental metal levels in a South African population. Our samples are from a cohort located in a region of South Africa with a history of mining for gold, uranium, manganese, platinum, and copper (Statistics South Africa 2017). To characterize the placental metal levels in these samples, we investigated the risk of metals exposure and interactions between metals, by comparing placental metal levels to previous placental measurements and testing for correlations between the metals. We assessed eleven

metals having differing levels of transport through the placenta – cadmium (Cd), which can accumulate in the placenta (Esteban-Vasallo et al. 2012), while arsenic (As), chromium (Cr), copper (Cu), lead (Pb), manganese (Mn), nickel (Ni), selenium (Se), and zinc (Zn) are transferred to the fetus (Punshon et al. 2015; Ziaee et al. 2007; Hardman et al. 2007; Takser et al. 2004; Gundacker and Hengstschläger 2012; Odland et al. 1997; Shennan 1988; Ford 2004). We also assessed palladium (Pd) and platinum (Pt) levels for which placental transport has not been determined. Previous work in this population measured metal levels in maternal and fetal blood. Lead, arsenic, and selenium levels were found to be correlated between maternal and fetal blood samples, suggesting that maternal exposure is reflective of fetal exposure for certain metals (Rudge et al. 2009). Due to the potential for metals exposure to affect fetal development, we also assessed relationships between the metal levels and placenta and birth outcomes.

Although not the direct topic of this study, it is important to note that S1000 samples assayed here are enriched for pregnancies with gestational diabetes mellitus (GDM), a metabolic disorder diagnosed during pregnancy. GDM is linked to changes in placental physiology such as altered maturity and branching of placental villi, which could explain the altered nutrient transport seen in GDM placentas (Taricco et al. 2009; Daskalakis et al. 2008; Cvitic, Desoye, and Hiden 2014; Schäfer-Graf et al. 1998; Gauster et al. 2012; Castillo-Castrejon and Powell 2017). GDM is also linked to increased placenta size likely caused by longitudinal vascular growth and enhanced branching angiogenesis (Taricco et al. 2009; Gauster et al. 2012; Edu et al. 2016; Daskalakis et al. 2008), possibly in response to the increased oxygen needs of the fetus in a hyperglycemic environment (Babawale et al. 2000; Jirkovská et al. 2002; Leach and Mayhew 2005).

Methods

Sample population

Forty-two de-identified placenta samples and matched maternal, placental and infant phenotype data were obtained from the Soweto First 1000 Days Pregnancy Cohort (S1000, MRC Developmental Pathways for Health Research Unit at the University of Witwatersrand). S1000 is a pregnancy cohort consisting of women of African descent located in Soweto, South Africa. Soweto is a highly transitioned poor urban area south-west of Johannesburg and is enclosed by dormant gold mine dumps. S1000 participants with gestational diabetes mellitus (GDM) had access to treatment at a tertiary hospital. The 42 samples were selected to be HIV negative, half GDM (n=21) and half nonGDM (n=21), and equal proportions of non-obese and obese in each disease group. For our sample group, 2 participants reported smoking before or during pregnancy and 2 reported alcohol consumption during pregnancy.

Placenta sampling

Placentas were weighed at delivery and samples collected within 1 hour of delivery. Tissue punches selected from this study were taken from the fetal side of the placenta disc, avoiding the umbilical cord and at least 3cm from the edge of the placenta. Blood was not removed from the tissue, but care was taken to avoid any visible lesions as well as areas that look

distinctly abnormal. Tissue punches were stored at -80°C until use (The INTERBIO-21st Consortium 2012).

Trace metals measurement

Placental trace metal analysis was performed by the University of North Carolina Biomarker Mass Spectrometry Core Facility. In brief, placental samples were digested with 70% nitric acid at room temperature for five hours before incubation at 85 degrees overnight. Samples were cooled to room temperature and 30% hydrogen peroxide added followed by an additional incubation at 85 degrees for 24hrs. Samples were then diluted to 4mL with deionized water and total concentrations of As, Cd, Cr, Cu, Pb, Mn, Ni, Pd, Pt, Se, and Zn were measured using Agilent Technologies 7500cx inductively coupled plasma mass spectrometer (ICP-MS), (Santa Clara, CA USA). External calibration and quality control standards were prepared from National Institute Standards Technology (NIST) traceable solutions (High Purity Standards, Charleston, SC. USA) (Laine et al. 2015).

For placental metals levels comparisons to previous cohorts, concentrations from previous studies that were reported relative to dry weight of placenta were converted to approximate wet weights by dividing dry weight by a conversion factor of 6.285 representing the average ratio of dry/wet weights, as previously reported (Iyengar and Rapp 2001).

Data transformations and variable score calculations

To normalize distributions, Cd, Cr, Cu, Pb, Mn, Ni, Pd, and Pt were log10 transformed before analyses. As, Se and Zn were normally distributed and therefore not log transformed. Placenta weight Z-scores were calculated using means and standard deviations of placenta weight from pregnancies matched for gestational age (GA) and offspring sex, as previously described (Almog et al. 2011). Birth weight and length Z-scores were calculated using the Intergrowth 21st Neonatal Size Calculator for newborn infants between 24 and 42 weeks' gestation (Intergrowth 21st 2017). The ratio of birthweight to placenta weight [birthweight (g)/placenta weight (g)] was calculated as an indicator of placenta efficiency (Hayward et al. 2016). Ponderal index [weight (g) $\times$ 100/[height (cm)]³] was calculated as an indicator of newborn adiposity (Armangil et al. 2011). Socio-economic status (SES) scores were created using a principal component analysis on household access to electricity, ownership of a television, refrigerator, personal computer, bicycle, vehicle, and the number of household rooms following the International Wealth Index (Smits and Steendijk 2015). From the principal component analysis, the first component explained the most variance in the sample population and was chosen as the SES indicator (Mean $\pm$ STD: $-1.17 \times 10^{-8} \pm 1.7$, Median: 0.168, Range: $-7.35 - 1.86$).

Statistical analyses

A two-tailed t-test or chi-square tests were used to test for differences between GDM status for each outcome and covariate. Multivariate regression was used to test for associations between GDM status and metal levels. Spearman's correlation was used to assess correlations between untransformed placental metal levels.

Linear regression models were run using STATA 15 (StataCorp, TX), to assess the association between placental metal levels and pregnancy outcomes: placenta weight Z-score, placenta efficiency, birthweight Z-score, ponderal index, and birth length Z-score. All models were adjusted for GDM, maternal age, maternal BMI, gestational age, parity, offspring sex, and socio-economic status (SES). A significance threshold of $p < 0.05$ was used for all models

Sensitivity analyses were conducted to identify major outliers in outcome variables, defined as individual data points that alone substantially influenced the significance of the results. As a result, we removed one outlier from the ponderal index dataset reducing sample size for ponderal index to 41. Sensitivity analyses were also conducted for maternal smoking status and showed no differential effect on the associations.

Results

Maternal and placenta clinical characteristics and birth outcomes

Placentas from forty-two pregnancies were selected from the S1000 cohort. Table 1 describes the maternal, placental, and birth outcome characteristics from these pregnancies. S1000 was enriched for gestational diabetes mellitus (GDM) pregnancies, therefore, we selected an equal number of GDM and nonGDM samples. GDM samples were on average from pregnancies with slightly higher maternal age ($p = 0.002$), maternal parity ($p = 0.003$), and proportion of male offspring ($p = 0.0001$) but did not significantly differ for other maternal, placental, or birth outcomes.

Placental levels of Cd, Cr, and Ni are lower in S1000 compared to other cohorts

To infer potential risk of exposure to metals during pregnancy in the S1000 pregnancy cohort, we compared S1000 placental levels of eleven metals (As, Cd, Cr, Cu, Pb, Mn, Ni, Pd, Pt, Se, and Zn) to previously described cohorts from other geographical locations (Table 2). This is necessary since reference doses and/or recommended exposure limits for placental metal levels have not yet been determined since the direct health risk associated with different levels of placental metals is unclear. Most of the metals exhibited similar or lower levels in S1000 compared to other populations. Cd, Cr, and Ni levels in S1000 were substantially lower compared to other reports (Table 2). For similarly measured wet weight concentrations, S1000 Cd levels were ~50-fold lower than a Chinese cohort (Guo et al. 2010); Cr levels were ~8- and 16-fold lower than a Turkish and Chinese cohort, respectively (Guo et al. 2010; Arica et al. 2013); and Ni levels were ~6- and 2-fold lower than a Turkish and Chinese cohort, respectively (Guo et al. 2010; Arica et al. 2013) (Table 2). S1000 As, Cu, Pb, Mn, Se, and Zn levels were mostly similar (less than 2-fold different) to other populations, while no reported data could be found for Pd and Pt (Table 2). The directionality of these comparative results remained the same even when GDM samples were excluded from the S1000 dataset.

S1000 placental metal levels show several strong positive correlations between metals

To better understand the potential interactions between metals, we assessed correlations between metals levels. We detected strong positive correlations ($r > 0.70$) between Pd & Pt

and Se & Zn ($r = 0.836$ and 0.908 , respectively) (Figure 1). Moderate correlations ($0.50 < r < 0.70$) were detected between Cd & Zn, Cr & Ni, Cu & Pb, Cu & Mn, Pb & Pd, Mn & Pd, and Mn & Se ($r = 0.579, 0.652, 0.630, 0.672, 0.551, 0.575$ and 0.509 , respectively) (Figure 1). All other significant correlations found were considered weakly correlated ($r < 0.5$). No significant negative correlations were found.

S1000 placental levels of Se, Cd, and Ni are associated with placental and/or birth outcomes

To determine whether placental levels of the eleven metals assessed in S1000 are associated with offspring developmental outcomes we used measurements of placenta weight Z-score and placenta efficiency (defined as the ratio of fetal to placental weight) as placental outcomes; and birth weight Z-score, birth length Z-score, and ponderal index as birth outcomes. For placental outcomes, Se was significantly negatively associated with placenta weight Z-score (Table 3) such every 1-unit decrease of Se was associated with a 7.74×10^3 unit increase in placenta weight Z-score. Cd was significantly positively associated with placenta efficiency (Table 3) such that every log transformed-unit increase of Cd was associated with a 1.06 increase in placenta efficiency (Table 3).

For birth outcomes, Ni placental levels were negatively associated with ponderal index (Table 4) such that every log transformed-unit decrease in Ni was associated with a 0.0995 unit ($\text{g} \times 100/\text{cm}^3$) increase in ponderal index (Table 4). None of the metal's levels were associated with birth weight or birth length Z-score after adjustments for covariates (Tables 4).

All regression models were adjusted for GDM, which would remove any effect of GDM on birth outcomes. To test separately whether there could be a causal effect of metals exposure on GDM, we used the placental metals levels as a proxy for maternal exposure and tested the association between metal's levels as a predictor and GDM status as an outcome. No significant associations were found.

Discussion

We have assessed the levels of eleven metals in a subset of the S1000 pregnancy cohort of Soweto-Johannesburg, South Africa, compared the levels to previous populations, and determined relationships with birth and placental outcomes. To the best of our knowledge, none of the previous populations used for comparison were specifically reported to have known/suspected increased risk of exposure or deficiency. Although we propose that the placental metals levels reflect differences in environmental exposure levels, unrelated differences in intrinsic features of the population such as placental metabolism or transport of metals, or technical differences in how the metals were measured may also play a role. Nonetheless, this comparison remains a valuable assessment to infer high vs. low risk of metals exposure vs. deficiency where limited data are available.

Cd is the only metal we measured known to accumulate in the placenta (Esteban-Vasallo et al. 2012). Our samples had substantially lower levels of Cd in comparison to previous populations and were moderately correlated with Zn levels ($r = 0.579$; Figure 1). This

relationship between Cd and Zn in the placenta was previously reported in a Ukrainian population, although it was a weak correlation ($r = 0.26$) (Zadorozhnaja et al. 2000). Cd is known to interact with essential metals, like zinc, by competitively binding to metal-binding proteins named metallothioneins (Brzóska and Moniuszko-Jakoniuk 2001). Cd has been shown to accumulate in liver and kidneys leading to increased retention of Zn in those same organs (Brzóska and Moniuszko-Jakoniuk 2001). The positive correlation between Cd and Zn in the placenta may be due to this interaction previously seen in other organs and is particular cause for concern during pregnancy. Zn supplementation in mice was shown to cause a >30% reduction of kidney and liver Cd levels (Pabis et al. 2018), perhaps Cd exposure during pregnancy could be mitigated by Zn supplementation. Interestingly, S1000 samples exhibited a positive association with placenta efficiency. This may be the result of more efficient placenta having higher barrier function such that more efficient placentas accumulate higher levels of Cd with increased environmental exposure.

We found lower S1000 placental levels of essential elements Cr, Ni and Zn compared to other reports, which may indicate that this population is at risk for deficiency. Maternal deficiency of Se and Zn were previously shown to be associated with increased risk for preterm birth and SGA (Iyengar, Kollmer, and Bowen. 1978; Ward, Machanon, and Mason 1987). We could not test for these outcomes specifically here but did show that Se was negatively associated with placenta weight Z-score. Se and Cd were both associated with placenta outcomes and significantly correlated (Figure 1), which may be reflective of the retention effects of Cd on essential nutrients. Despite a strong correlation between Se and Zn levels (Figure 1), Zn was not associated with any outcomes. Ni levels were positively correlated with Cr levels ($r = 0.652$; Figure 1). No current studies have shown this interaction between Ni and Cr in the placenta, however, Cr was recently shown to be positively correlated to the essential element Mn (Freire et al. 2019) but this correlation was not significant in our study. In the case that our findings reflect maternal deficiency in both Cr and Ni, this population should be studied further as Ni deficiency in animal models has also been shown to affect development including reduced birth weight, decreased weight gain and increased risk of preweaning mortality (Anke et al. 1978). In S1000, Ni was negatively associated with ponderal index. The relationships found here may reflect adverse effects of inadequate levels of these metals on development, however, further studies are required to fully elucidate these effects.

Conclusions

This study provides an important examination of placental metals levels in a previously unassessed South African cohort and gives some preliminary evidence suggesting a link with birth outcomes. Comparisons to findings in other cohorts suggest S1000 may be potentially deficient for Cr, Ni, and Zn. Further studies should investigate the roles of these important metals in maternal and child health.

Acknowledgments:

We thank Catherine Zimmer (UNC Odum Institute) for assistance with the statistical analyses, Michelle Mendez (Environmental and Occupational Health Department, University of Pittsburgh) for assistance with statistical

analysis and revisions of paper, and Peter Cable & Wanda Bodnar (UNC Biomarker and Mass Spectrometry Core) for assistance with measuring metals levels.

Funding: National Institute of Environmental Health Sciences (NIEHS)-P30ES010126 to the UNC Biomarker and Mass Spectrometry Core for placental metals measurements; NIEHS Transition to Independent Environmental Health Research Career Development Award [KES023849A] to FI for study design, data analyses, data interpretation & manuscript writing; the UK Medical Research Council (MRC) DFID African Research Leader Scheme Award to SN for sample and data collection, data interpretation & manuscript writing; and the T32 predoctoral fellowship from the National Heart, Lung, and Blood Institute (NHLBI) funded Global Cardiometabolic Disease Training Grant [5T32HL129969] to LM for data analyses, data interpretation & manuscript writing. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

REFERENCES

- Almog B, Shehata F, Aljabri S, Levin I, Shalom-Paz E, and Shrim A. 2011 “Placenta Weight Percentile Curves for Singleton and Twins Deliveries.” *Placenta* 32 (1): 58–62. doi:10.1016/j.placenta.2010.10.008. [PubMed: 21036395]
- Anke M, Partscheffeld M, Grün M, and Groppe B. 1978 “Nickel — an Essential Element.” *Archiv Für Tierernährung* 28 (2): 83–90. doi:10.1080/17450397809426782.
- Arbuckle, Tye E, Liang Chun Lei, Morisset Anne-Sophie, Fisher Mandy, Weiler Hope, Cirtiu Ciprian Mihai, Legrand Melissa, et al. 2016 “Maternal and Fetal Exposure to Cadmium, Lead, Manganese and Mercury: The MIREC Study.” *Chemosphere* 163 (November): 270–282. doi:10.1016/j.chemosphere.2016.08.023. [PubMed: 27540762]
- Arica Enes, Kayaalti Zeliha, Akyuzlu Dilek Kaya, and Soylemezoglu Tulin. 2013 “Assessment of Chromium and Nickel Levels in Maternal Blood, Placenta and Cord Blood by Graphite Furnace Atomic Absorption Spectrometry.” *Toxicology Letters* 221 (August): S248. doi:10.1016/j.toxlet.2013.05.616.
- Armangil Didem, Yurdakök Murat, Korkmaz Ay e, Yi it Sule, and Tekinalp Gülsevin. 2011 “Ponderal Index of Large-for-gestational Age Infants: Comparison Between Infants of Diabetic and Non-diabetic Mothers.” *The Turkish Journal of Pediatrics* 53 (2): 169–172. [PubMed: 21853654]
- Babawale MO, Lovat S, Mayhew TM, Lammiman MJ, James DK, and Leach L. 2000 “Effects of Gestational Diabetes on Junctional Adhesion Molecules in Human Term Placental Vasculature.” *Diabetologia* 43 (9): 1185–1196. doi:10.1007/s001250051511. [PubMed: 11043866]
- Bloom Michael S, Surdu Simona, Neamtui Iulia A, and Gurzau Eugen S. 2014 “Maternal Arsenic Exposure and Birth Outcomes: a Comprehensive Review of the Epidemiologic Literature Focused on Drinking Water.” *International Journal of Hygiene and Environmental Health* 217 (7): 709–719. doi:10.1016/j.ijheh.2014.03.004. [PubMed: 24713268]
- Brzóska MM, and Moniuszko-Jakoniuk J. 2001 “Interactions Between Cadmium and Zinc in the Organism.” *Food and Chemical Toxicology* 39 (10): 967–980. doi:10.1016/S0278-6915(01)00048-5. [PubMed: 11524135]
- Castillo-Castrejon Marisol, and Powell Theresa L. 2017 “Placental Nutrient Transport in Gestational Diabetic Pregnancies.” *Frontiers in Endocrinology* 8 (November): 306. doi:10.3389/fendo.2017.00306. [PubMed: 29163373]
- Chaffee Benjamin W, and King Janet C. 2012 “Effect of Zinc Supplementation on Pregnancy and Infant Outcomes: a Systematic Review.” *Paediatric and Perinatal Epidemiology* 26 Suppl 1 (July): 118–137. doi:10.1111/j.1365-3016.2012.01289.x. [PubMed: 22742606]
- Cooke Gerard M. 2014 “Biomonitoring of Human Fetal Exposure to Environmental Chemicals in Early Pregnancy.” *Journal of Toxicology and Environmental Health. Part B, Critical Reviews* 17 (4): 205–224. doi:10.1080/10937404.2014.898167.
- Cvitc Silvija, Desoye Gernot, and Hiden Ursula. 2014 “Glucose, Insulin, and Oxygen Interplay in Placental Hypervascularisation in Diabetes Mellitus.” *BioMed Research International* 2014 (September): 145846. doi:10.1155/2014/145846. [PubMed: 25258707]
- Daskalakis George, Marinopoulos Spyros, Krielesi Vasiliki, Papapanagiotou Angeliki, Papantoniou Nikolaos, Mesogitis Spyros, and Antsaklis Aris. 2008 “Placental Pathology in Women with

- Gestational Diabetes.” *Acta Obstetrica et Gynecologica Scandinavica* 87 (4): 403–407. doi: 10.1080/00016340801908783. [PubMed: 18382864]
- Donnelly Leo, and Campling Gillian. 2016 “Functions of the Placenta.” *Anaesthesia & Intensive Care Medicine* 17 (7): 349–353. doi:10.1016/j.mpaic.2016.04.004.
- Edu Antoine, Teodorescu Cristina, Dobjanschi Carmen Gabriela, Socol Zi a Zsuzsana, Teodorescu Valeriu, Matei Alexandru, Albu Dinu Florin, and Radulian Gabriela. 2016 “Placenta Changes in Pregnancy with Gestational Diabetes.” *Romanian Journal of Morphology and Embryology* 57 (2): 507–512. [PubMed: 27516026]
- Esteban-Vasallo María D, Aragonés Nuria, Pollan Marina, López-Abente Gonzalo, and Perez-Gomez Beatriz. 2012 “Mercury, Cadmium, and Lead Levels in Human Placenta: a Systematic Review.” *Environmental Health Perspectives* 120 (10): 1369–1377. doi:10.1289/ehp.1204952. [PubMed: 22591711]
- Ford Dianne. 2004 “Intestinal and Placental Zinc Transport Pathways.” *The Proceedings of the Nutrition Society* 63 (1): 21–29. doi:10.1079/PNS2003320. [PubMed: 15070437]
- Freire Carmen, Amaya Esperanza, Gil Fernando, Murcia Mario, Sabrina Llop Maribel Casas, Vrijheid Martine, et al. 2019 “Placental Metal Concentrations and Birth Outcomes: The Environment and Childhood (INMA) Project.” *International Journal of Hygiene and Environmental Health* 222 (3): 468–478. doi:10.1016/j.ijheh.2018.12.014. [PubMed: 30638867]
- Gauster M, Desoye G, Tötsch M, and Hiden U. 2012 “The Placenta and Gestational Diabetes Mellitus.” *Current Diabetes Reports* 12 (1): 16–23. doi:10.1007/s11892-011-0244-5. [PubMed: 22102097]
- Gundacker Claudia, and Hengstschläger Markus. 2012 “The Role of the Placenta in Fetal Exposure to Heavy Metals.” *Wiener Medizinische Wochenschrift* (1946) 162 (9–10): 201–206. doi:10.1007/s10354-012-0074-3. [PubMed: 22717874]
- Guo Yongyong, Huo Xia, Li Yan, Wu Kusheng, Liu Junxiao, Huang Jingrong, Zheng Guina, et al. 2010 “Monitoring of Lead, Cadmium, Chromium and Nickel in Placenta from an E-waste Recycling Town in China.” *The Science of the Total Environment* 408 (16): 3113–3117. doi: 10.1016/j.scitotenv.2010.04.018. [PubMed: 20451954]
- Hardman Belinda, Michalczyk Agnes, Greenough Mark, Camakaris James, Mercer J Julian, and Ackland Leigh. 2007 “Distinct Functional Roles for the Menkes and Wilson Copper Translocating P-type ATPases in Human Placental Cells.” *Cellular Physiology and Biochemistry* 20 (6): 1073–1084. doi:10.1159/000110718. [PubMed: 17975309]
- Hayward Christina E, Lean Samantha, Sibley Colin P, Jones Rebecca L, Wareing Mark, Greenwood Susan L, and Dilworth Mark R. 2016 “Placental Adaptation: What Can We Learn from Birthweight:Placental Weight Ratio?” *Frontiers in Physiology* 7 (February): 28. doi:10.3389/fphys.2016.00028. [PubMed: 26903878]
- He Zhenli L, Yang Xiaoe E, and Stoffella Peter J. 2005 “Trace Elements in Agroecosystems and Impacts on the Environment.” *Journal of Trace Elements in Medicine and Biology* 19 (2–3): 125–140. doi:10.1016/j.jtemb.2005.02.010. [PubMed: 16325528]
- Hu Jie, Xia Wei, Pan Xinyun, Zheng Tongzhang, Zhang Bin, Zhou Aifen, Buka Stephen L, et al. 2017 “Association of Adverse Birth Outcomes with Prenatal Exposure to Vanadium: a Population-based Cohort Study.” *The Lancet Planetary Health* 1 (6): e230–e241. doi:10.1016/S2542-5196(17)30094-3. [PubMed: 29851608]
- Intergrowth 21st. 2017 “The International Fetal and Newborn Growth Consortium for the 21st Century” <https://intergrowth21.tghn.org/>.
- Iyengar GV, Kollmer WE, and Bowen HJM. 1978 *The Elemental Composition of Human Tissues and Body Fluids* New York: Verlag Chemie.
- Iyengar GV, and Rapp A. 2001 “Human Placenta as a ‘Dual’ Biomarker for Monitoring Fetal and Maternal Environment with Special Reference to Potentially Toxic Trace Elements. Part 1: Physiology, Function and Sampling of Placenta for Elemental Characterisation.” *The Science of the Total Environment* 280 (1–3): 195–206. [PubMed: 11763267]
- Jaishankar Monisha, Tseten Tenzin, Anbalagan Naresh, Mathew Blessy B, and Beeregowda Krishnamurthy N. 2014 “Toxicity, Mechanism and Health Effects of Some Heavy Metals.” *Interdisciplinary Toxicology* 7 (2): 60–72. doi:10.2478/intox-2014-0009. [PubMed: 26109881]

- Jirkovská Marie, Lucie Kubínová Jirí Janáček, Milena Moravcová Vratislav Krejčí, and Karen Petr. 2002 “Topological Properties and Spatial Organization of Villous Capillaries in Normal and Diabetic Placentas.” *Journal of Vascular Research* 39 (3): 268–278. doi:10.1159/000063692. [PubMed: 12097825]
- Keen CL, Uriu-Hare JY, Hawk SN, Jankowski MA, Daston GP, Kwik-Urbe CL, and Rucker RB. 1998 “Effect of Copper Deficiency on Prenatal Development and Pregnancy Outcome.” *The American Journal of Clinical Nutrition* 67 (5 Suppl): 1003S–1011S. doi:10.1093/ajcn/67.5.1003S. [PubMed: 9587143]
- Laine Jessica E, Ray Paul, Bodnar Wanda, Cable Peter H, Boggess Kim, Offenbacher Steven, and Fry Rebecca C. 2015 “Placental Cadmium Levels Are Associated with Increased Preeclampsia Risk.” *Plos One* 10 (9): e0139341. doi:10.1371/journal.pone.0139341. [PubMed: 26422011]
- Lawn Joy E, Blencowe Hannah, Oza Shefali, You Danzhen, Lee Anne C C, Waiswa Peter, Lalli Marek, et al. 2014 “Every Newborn: Progress, Priorities, and Potential Beyond Survival.” *The Lancet* 384 (9938): 189–205. doi:10.1016/S0140-6736(14)60496-7.
- Leach Lopa, and Mayhew Terry M. 2005 “Vasculogenesis and Angiogenesis in the Diabetic Placenta.” *Diabetology of Pregnancy* 17: 110–126.
- Luo Yiwen, McCullough Lauren E, Tzeng Jung-Ying, Darrah Thomas, Vengosh Avner, Maguire Rachel L, Maity Arnab, et al. 2017 “Maternal Blood Cadmium, Lead and Arsenic Levels, Nutrient Combinations, and Offspring Birthweight.” *BMC Public Health* 17 (1): 354. doi:10.1186/s12889-017-4225-8. [PubMed: 28438148]
- Mariath Aline B, Bergamaschi Denise P, Rondó Patrícia H C, Tanaka Ana C D'A, Hinnig Patrícia de Fragas, Abbade Joécio F, and Diniz Simone G. 2011 “The Possible Role of Selenium Status in Adverse Pregnancy Outcomes.” *The British Journal of Nutrition* 105 (10): 1418–1428. doi: 10.1017/S0007114510005866. [PubMed: 21338537]
- Mattison Donald R. 2010 “Environmental Exposures and Development.” *Current Opinion in Pediatrics* 22 (2): 208–218. doi:10.1097/MOP.0b013e32833779bf. [PubMed: 20216314]
- Myllynen P, Pasanen M, and Pelkonen O. 2005 “Human Placenta: a Human Organ for Developmental Toxicology Research and Biomonitoring.” *Placenta* 26 (5): 361–371. doi:10.1016/j.placenta.2004.09.006. [PubMed: 15850640]
- Odland JO, Romanova N, Sand G, Thomassen Y, Salbu B, Lund Eiliv, and Nieboer E. 1997 “Cadmium, Lead, Mercury, Nickel, and Cesium-137 Concentrations in Blood, Urine, or Placenta from Mothers and Newborns Living in Arctic Areas of Russia and Norway.” In *Environmental Biomonitoring: Exposure Assessment and Specimen Banking*, edited by Subramanian KS and Iyengar GV, 654:135–150. ACS Symposium Series. Washington, DC: American Chemical Society. doi:10.1021/bk-1997-0654.ch013.
- Pabis Kamil, Gundacker Claudia, Giacconi Robertina, Basso Andrea, Costarelli Laura, Piacenza Francesco, Strizzi Sergio, Provinciali Mauro, and Malavolta Marco. 2018 “Zinc Supplementation Can Reduce Accumulation of Cadmium in Aged Metallothionein Transgenic Mice.” *Chemosphere* 211 (November): 855–860. doi:10.1016/j.chemosphere.2018.08.017. [PubMed: 30103140]
- Parajuli Rajendra Prasad, Fujiwara Takeo, Umezaki Masahiro, and Watanabe Chiho. 2013 “Association of Cord Blood Levels of Lead, Arsenic, and Zinc with Neurodevelopmental Indicators in Newborns: a Birth Cohort Study in Chitwan Valley, Nepal.” *Environmental Research* 121 (February): 45–51. doi:10.1016/j.envres.2012.10.010. [PubMed: 23164520]
- Punshon Tracy, Davis Matthew A, Marsit Carmen J, Theiler Shaleen K, Baker Emily R, Jackson Brian P, Conway David C, and Karagas Margaret R. 2015 “Placental Arsenic Concentrations in Relation to Both Maternal and Infant Biomarkers of Exposure in a US Cohort.” *Journal of Exposure Science & Environmental Epidemiology* 25 (6): 599–603. doi:10.1038/jes.2015.16. [PubMed: 25805251]
- Rudge Cibele V, Röllin Halina B, Nogueira Claudina M, Thomassen Yngvar, Rudge Marilza C, and Odland Jon Ø. 2009 “The Placenta as a Barrier for Toxic and Essential Elements in Paired Maternal and Cord Blood Samples of South African Delivering Women.” *Journal of Environmental Monitoring* 11 (7): 1322–1330. doi:10.1039/b903805a. [PubMed: 20449220]
- Saenz JM, Fernandez MF, Olea N, and Inc. Nova Science Publishers. 2013 “Exposure to Endocrine Disruptors During Pregnancy and Their Potential Health Effects on Newborns.” In *Placenta:*

- Development, Function and Diseases, edited by Nicholson Richard, 167–186. Nova Science Publishers, Incorporated.
- Schäfer-Graf UM, Dupak J, Vogel M, Dudenhausen JW, Kjos SL, Buchanan TA, and Vetter K. 1998 “Hyperinsulinism, Neonatal Obesity and Placental Immaturity in Infants Born to Women with One Abnormal Glucose Tolerance Test Value.” *Journal of Perinatal Medicine* 26 (1): 27–36. [PubMed: 9595364]
- Shah Dheeraj, and Sachdev HPS. 2006 “Zinc Deficiency in Pregnancy and Fetal Outcome.” *Nutrition Reviews* 64 (1): 15–30. doi:10.1301/nr.2006.jan.15-30. [PubMed: 16491666]
- Shennan DB. 1988 “Selenium (selenate) Transport by Human Placental Brush Border Membrane Vesicles.” *The British Journal of Nutrition* 59 (1): 13–19. doi:10.1079/BJN19880005. [PubMed: 3345300]
- Shirai Sayaka, Suzuki Yayoi, Yoshinaga Jun, and Mizumoto Yoshifumi. 2010 “Maternal Exposure to Low-level Heavy Metals During Pregnancy and Birth Size.” *Journal of Environmental Science and Health. Part A, Toxic/Hazardous Substances & Environmental Engineering* 45 (11): 1468–1474. doi:10.1080/10934529.2010.500942.
- Singh Reena, Gautam Neetu, Mishra Anurag, and Gupta Rajiv. 2011 “Heavy Metals and Living Systems: An Overview.” *Indian Journal of Pharmacology* 43 (3): 246–253. doi: 10.4103/0253-7613.81505. [PubMed: 21713085]
- Smits Jeroen, and Steendijk Roel. 2015 “The International Wealth Index (IWI).” *Social Indicators Research* 122 (1): 65–85. doi:10.1007/s11205-014-0683-x.
- Statistics South Africa. 2017 Mining Industry, 2015 Vol. 02.
- Sun Hong, Chen Wen, Wang Dongyue, Jin Yinlong, Chen Xiaodong, and Xu Yan. 2014 “The Effects of Prenatal Exposure to Low-level Cadmium, Lead and Selenium on Birth Outcomes.” *Chemosphere* 108 (August): 33–39. doi:10.1016/j.chemosphere.2014.02.080. [PubMed: 24875909]
- Takser Larissa, Lafond Julie, Bouchard Maryse, St-Amour Genevieve, and Mergler Donna. 2004 “Manganese Levels During Pregnancy and at Birth: Relation to Environmental Factors and Smoking in a Southwest Quebec Population.” *Environmental Research* 95 (2): 119–125. doi: 10.1016/j.envres.2003.11.002. [PubMed: 15147916]
- Taricco E, Radaelli T, Rossi G, de Santis M S Nobile, Bulfamante GP, Avagliano L, and Cetin I. 2009 “Effects of Gestational Diabetes on Fetal Oxygen and Glucose Levels in Vivo.” *BJOG: An International Journal of Obstetrics and Gynaecology* 116 (13): 1729–1735. doi:10.1111/j.1471-0528.2009.02341.x. [PubMed: 19832834]
- Tchounwou Paul B, Yedjou Clement G, Patlolla Anita K, and Sutton Dwayne J. 2012 “Heavy Metal Toxicity and the Environment.” *Exs* 101: 133–164. doi:10.1007/978-3-7643-8340-4_6. [PubMed: 22945569]
- The INTERBIO-21st Consortium. 2012 The Functional Classification of Abnormal Fetal and Neonatal Growth Phenotypes: Biological Sample Collection Operations Manual University of Oxford.
- Thomas Shari, Arbuckle Tye E, Fisher Mandy, Fraser William D, Ettinger Adrienne, and King Will. 2015 “Metals Exposure and Risk of Small-for-gestational Age Birth in a Canadian Birth Cohort: The MIREC Study.” *Environmental Research* 140 (July): 430–439. doi:10.1016/j.envres.2015.04.018. [PubMed: 25967284]
- Vaughan OR, Sferruzzi-Perri AN, Coan PM, and Fowden AL. 2011 “Environmental Regulation of Placental Phenotype: Implications for Fetal Growth.” *Reproduction, Fertility, and Development* 24 (1): 80–96. doi:10.1071/RD11909.
- Ward NI, Machanon TD, and Mason JA. 1987 “Elemental Analysis of Human Placenta by Neutron Irradiation and Gamma-ray Spectrometry” 113 (2): 501–514.
- Wirth Julia J, and Mijal Renée S. 2010 “Adverse Effects of Low Level Heavy Metal Exposure on Male Reproductive Function.” *Systems Biology in Reproductive Medicine* 56 (2): 147–167. doi: 10.3109/19396360903582216. [PubMed: 20377313]
- Wright Robert O, and Baccarelli Andrea. 2007 “Metals and Neurotoxicology.” *The Journal of Nutrition* 137 (12): 2809–2813. doi:10.1093/jn/137.12.2809. [PubMed: 18029504]

- Xie Xin, Ding Guodong, Cui Chang, Chen Limei, Gao Yu, Zhou Yijun, Shi Rong, and Tian Ying. 2013 "The Effects of Low-level Prenatal Lead Exposure on Birth Outcomes." *Environmental Pollution* 175 (April): 30–34. doi:10.1016/j.envpol.2012.12.013. [PubMed: 23321271]
- Zadorozhnaja TD, Little RE, Miller RK, Mendel NA, Taylor RJ, Presley BJ, and Gladen BC. 2000 "Concentrations of Arsenic, Cadmium, Copper, Lead, Mercury, and Zinc in Human Placentas from Two Cities in Ukraine." *Journal of Toxicology and Environmental Health. Part A* 61 (4): 255–263. doi:10.1080/00984100050136571. [PubMed: 11071319]
- Ziaee H, Daniel J, Datta AK, Blunt S, and McMinn DJW. 2007 "Transplacental Transfer of Cobalt and Chromium in Patients with Metal-on-metal Hip Arthroplasty: a Controlled Study." *The Journal of Bone and Joint Surgery. British Volume* 89 (3): 301–305. doi:10.1302/0301-620X.89B3.18520. [PubMed: 17356138]

Figure 1.

Correlations between placental metal levels. Spearman correlation was used to assess the correlation between untransformed placental metal levels. R-values are shown in the top-right portion of the graph. N=38–42. Only the statistically significant correlations are denoted in the lower-left portion of the graph (* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$). All r-values with light blue colors had weak correlations ($-0.2 < r < 0.2$). The color scheme denotes the strength of the r-values, with positive correlations in blue and negative correlations in red.

Table 1.

Characteristics of the 42 mother-infant dyads from the S1000 Days cohort.

Maternal Characteristics	Mean $\pm$ SD/ (%) of total	Range
Maternal BMI (1 st trimester, kg/m ²)	29.4 $\pm$ 6.8	18.2 – 48
Maternal Age (years)	30.2 $\pm$ 5.6	19 – 43
Gestational age (weeks)	38.4 $\pm$ 2.0	33 – 41
Parity (# of full-term pregnancies)	1.2 $\pm$ 0.9	0 – 4
Mode of Delivery (% cesarean)	59%	n.a.
Placenta Outcomes		
	n = 33	
Placenta weight (g)	478.7 $\pm$ 89.7	290 – 628
Placenta efficiency (grams fetus/gram placenta)	6.5 $\pm$ 1.0	4.6 – 8.9
Birth Outcomes		
Birth weight (kg)	3.1 $\pm$ 0.5	1.9 – 4.1
Birth length (cm)	48.4 $\pm$ 3.3	40.7 – 55.2
Ponderal index (g x 100/cm ³)	2.7 $\pm$ 0.3	1.8 – 4.1
Sex (% female)	38%	n.a.

n.a., not applicable

Table 2.

Placental metal levels for 42 placental samples from S1000 Days South African cohort compared to previous studies.

Trace Metal	N	Arithmetic Mean $\pm$ SD	Reports from other cohorts (approximate values)			Reference
			Location	Mean	Range	
Arsenic (ppt)	42	1,685.7 $\pm$ 661.9	USA Mexico Multiple	NR 2,600.0 6,000.0	NR 500.0 – 6,000.0 3,000.0 – 12,000.0	(Punshon et al. 2015) ¹ (Diaz-Barriga et al. 1995) ¹ (Iyengar and Rapp 2001) ²
Cadmium (ppb)	38	2.5 $\pm$ 1.5	Sweden China Italy	NR NR NR	NR 1.1 – 19.1 2.3 – 393.5 2.1 – 28.6	(Osman et al. 2000) ¹ (Guo et al. 2010) ¹ (Rovero et al. 2015) ²
Chromium (ppb)	41	26 $\pm$ 36.2	Turkey China Italy	220.7 NR NR	NR NR 83.5 – 6,638.9 2.4 – 300.7	(Arica et al. 2013) ¹ (Guo et al. 2010) ¹ (Rovero et al. 2015) ²
Copper (ppb)	42	1,318.1 $\pm$ 809.1	USA Sweden Italy	1,598.0 NR NR	NR NR 635.5 – 1,270.9 588.7 – 4,932.4	(Karp and Robertson 1977) ¹ (Osman et al. 2000) ¹ (Rovero et al. 2015) ²
Lead (ppb)	42	21.5 $\pm$ 17.9	Sweden China Multiple	NR NR 34.0	NR NR 0 – 130.5 4.5 – 3,176.1 5 – 60	(Osman et al. 2000) ¹ (Guo et al. 2010) ¹ (Iyengar and Rapp 2001) ²
Manganese (ppb)	42	110.6 $\pm$ 68.1	USA Sweden Italy	115.0 NR NR	NR NR 35.7 – 280.2 11.8 – 795.5	(Karp and Robertson 1977) ¹ (Osman et al. 2000) ¹ (Rovero et al. 2015) ²
Nickel (ppb)	40	20.3 $\pm$ 50.9	Turkey China Multiple	124.2 NR 36.0	NR NR 1.76 – 593.7 9 – 62	(Arica et al. 2013) ¹ (Guo et al. 2010) ¹ (Iyengar and Rapp 2001) ²
Palladium (ppb)	42	21.5 $\pm$ 11.9	NR	NR	NR	NR
Platinum (ppt)	42	754.5 $\pm$ 743.2	NR	NR	NR	NR
Selenium (ppb)	42	147.7 $\pm$ 37.3	Sweden Croatia Italy	NR NR 100.2	NR 157.9 – 260.6 100.0 – 240.0 55.7 – 151.2	(Osman et al. 2000) ¹ (Klapek et al. 2008) ¹ (Rovero et al. 2015) ²
Zinc (ppm)	42	7.8 $\pm$ 2.0	USA Sweden Italy	10.2 NR 8.1	NR NR 7.9 – 18.3 1.4 – 15.9	(Karp and Robertson 1977) ¹ (Osman et al. 2000) ¹ (Rovero et al. 2015) ²

Italics indicates lower levels and bold indicates higher levels reported for other cohorts compared to S1000. Concentration relative to wet weight¹ or dry weight converted to wet weight² using conversion factor of 6.285 as previously described (Iyengar and Rapp, 2001). NR, not reported.

Table 3.

Association between placental metal levels and placenta outcomes (n = 33).

Metal Measured in Placenta	Placenta weight Z-score		Placenta efficiency	
	(unadjusted) β (p-value)	(adjusted) β (p-value)	(unadjusted) β (p-value)	(adjusted) β (p-value)
Arsenic	-0.000219 (0.199)	-0.000142 (0.475)	0.000168 (0.528)	0.000062 (0.846)
Cadmium ^I	-0.328 (0.143)	-0.464 (0.105)	0.842 (0.011*)	1.06 (0.011*)
Chromium ^I	0.0886 (0.602)	-0.0757 (0.707)	0.125 (0.634)	0.234 (0.461)
Copper ^I	-0.0773 (0.753)	-0.182 (0.519)	-0.0437 (0.908)	-0.0755 (0.867)
Lead ^I	0.0702 (0.703)	0.109 (0.634)	-0.315 (0.264)	-0.569 (0.109)
Manganese ^I	-0.152 (0.612)	-0.0138 (0.968)	-0.0774 (0.868)	-0.38 (0.484)
Nickel ^I	-0.0205 (0.898)	-0.125 (0.474)	0.206 (0.416)	0.289 (0.334)
Palladium ^I	-0.264 (0.194)	-0.254 (0.259)	0.446 (0.154)	0.349 (0.331)
Platinum ^I	-0.353 (0.040*)	-0.306 (0.108)	0.489 (0.067)	0.376 (0.219)
Selenium	-0.00712 (0.023*)	-0.00774 (0.044*)	0.00749 (0.129)	0.00838 (0.181)
Zinc	-0.000116 (0.051*)	-0.000108 (0.125)	0.000115 (0.220)	0.000104 (0.364)

^I log-transformed

* statistically significant

Adjusted for GDM, maternal age, gestational age, maternal BMI, parity, offspring sex, and SES

Table 4.

Association between placental metal levels and birth outcomes (n = 42).

Metal Measured in Placenta	Birth weight Z-score		Birth length Z-score		Ponderal Index (n = 41)	
	(unadjusted) β (p-value)	(adjusted) β (p-value)	(unadjusted) β (p-value)	(adjusted) β (p-value)	(unadjusted) β (p-value)	(adjusted) β (p-value)
Arsenic	-0.000289 (0.182)	-0.000116 (0.569)	-0.000018 (0.964)	0.00028 (0.521)	-0.0000487 (0.471)	-0.0000759 (0.326)
Cadmium ^I	-0.397 (0.110)	-0.189 (0.459)	-0.734 (0.126)	-0.588 (0.286)	0.0493 (0.554)	0.035 (0.720)
Chromium ^I	0.2151 (0.176)	0.166 (0.228)	0.458 (0.113)	0.397 (0.184)	-0.106 (0.025*)	-0.098 (0.056)
Copper ^I	-0.229 (0.452)	-0.484 (0.081)	-0.561 (0.311)	-0.873 (0.146)	0.091 (0.314)	0.0905 (0.383)
Lead ^I	-0.193 (0.385)	-0.234 (0.234)	-0.161 (0.692)	-0.265 (0.535)	-0.0371 (0.577)	-0.0278 (0.703)
Manganese ^I	-0.155 (0.639)	-0.196 (0.516)	-0.230 (0.702)	-0.411 (0.527)	0.0286 (0.771)	0.0409 (0.713)
Nickel ^I	0.203 (0.131)	0.196 (0.071)	0.456 (0.059)	0.423 (0.082)	-0.114 (0.004*)	-0.0995 (0.016*)
Palladium ^I	-0.0196 (0.943)	-0.0914 (0.703)	-0.182 (0.717)	-0.300 (0.561)	-0.0835 (0.332)	-0.0865 (0.351)
Platinum ^I	-0.159 (0.481)	-0.148 (0.456)	-0.349 (0.397)	-0.388 (0.363)	-0.0381 (0.589)	-0.0506 (0.517)
Selenium	-0.00728 (0.055)	-0.00549 (0.156)	-0.0152 (0.026*)	-0.0145 (0.080)	0.00165 (0.152)	0.0012 (0.412)
Zinc	-0.000147 (0.039*)	-0.0000754 (0.290)	-0.000309 (0.016*)	-0.000272 (0.072)	0.0000323 (0.140)	0.0000276 (0.313)

^I log-transformed

* statistically significant

Adjusted for GDM, maternal age, gestational age, maternal BMI, parity, offspring sex, and SES