Published in final edited form as:

Neurotoxicol Teratol. 2019 ; 74: 106810. doi:10.1016/j.ntt.2019.106810.

Prenatal and Recent Methylmercury Exposure and Heart Rate Variability in Young Adults: the Seychelles Child Development Study

Wojciech Zareba¹, Sally W. Thurston², Grazyna Zareba³, Jean Philippe Couderc¹, Katie Evans², Jean Xia¹, Gene E Watson^{3,10}, JJ Strain⁴, Emeir McSorley⁴, Alison Yeates⁴, Maria Mulhern⁴, Conrad F Shamlaye⁵, Pascal Bovet⁶, Edwin van Wijngaarden^{3,7,8,10}, Philip W Davidson^{3,8}, and Gary J Myers^{8,3,9}

⁽¹⁾Heart Research, Cardiology Division; University of Rochester Medical Center, Rochester, NY

⁽²⁾Department of Biostatistics and Computational Biology; University of Rochester Medical Center, Rochester, NY

⁽³⁾Department of Environmental Medicine, University of Rochester Medical Center, Rochester, NY

⁽⁴⁾University of Ulster, Coleraine, Northern Ireland, UK

⁽⁵⁾Ministry of Health, Republic of Seychelles, Seychelles

⁽⁶⁾University Institute of Social and Preventive Medicine, Lausanne, Switzerland

⁽⁷⁾Department of Public Health Sciences; University of Rochester Medical Center, Rochester, NY

⁽⁸⁾Department of Pediatrics, University of Rochester Medical Center, Rochester, NY

⁽⁹⁾Department of Neurology, University of Rochester Medical Center, Rochester, NY

⁽¹⁰⁾Eastman Institute for Oral Health, University of Rochester, Rochester, NY

Abstract

Epidemiologic evidence of an adverse association between exposure to methylmercury (MeHg) from consuming fish and heart rate variability (HRV) is inconclusive. We aimed to evaluate MeHg exposure in relation to HRV parameters in a large cohort of young adults from a high fish consuming population in the Republic of Seychelles. Main Cohort participants in the Seychelles Child Development Study were evaluated at a mean age of 19 years. Prenatal MeHg exposure was determined in maternal hair growing during pregnancy and recent exposure in participant's hair taken at the evaluation. The evaluation consisted of short (~2 hours) and long (overnight) Holter recordings obtained in 514 and 203 participants, respectively. Multivariable analyses examined the association of prenatal and recent MeHg exposure (in separate models) with time-domain and frequency-domain HRV parameters in different physiologic circumstances: supine position, standing position, mental stress when undergoing a mathematics test, sleep, and long recording. Prenatal MeHg exposure was not associated with any of the 23 HRV parameters studied after adjustment for multiplicity. The recent MeHg showed a trend toward significance only for few variables in the primary model. However, after additional adjustment for activity levels,

Address for Correspondence: Wojciech Zareba, MD, PhD, Cardiovascular Clinical Research Center, Cardiology Division, University of Rochester Medical Center, 265 Crittenden Blvd., Rochester, NY 14642, wojciech_zareba@urmc.rochester.edu.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

The authors declare they have no actual or potential competing financial interests.

polyunsaturated fatty acids, and multiplicity none were significant after a Bonferroni adjustment. In conclusion, prenatal and recent MeHg exposure had no consistent pattern of associations to support the hypothesis that they are adversely associated with heart rate variability in this study population that consumes large amounts of fish.

Fish is an important source of nutrition worldwide with about 3.1 billion people depending daily on fish as their primary source of protein (FAO 2016). However, all fish also contain naturally occurring methylmercury (MeHg) which is a known neurotoxicant in high dosages. Maternal consumption during pregnancy of industrially polluted fish (after MeHg was discharged in waste water from a chemical plant and seed grain treated with a MeHg fungicide) have been associated with severe neurological deficits in children (WHO 2007). Since neurotoxicity was observed after large exposure to MeHg, several studies have evaluated heart rate variability (HRV) to investigate the association between prenatal or recent MeHg exposure (Sorensen et al. 1999; Oka et al. 2002; Grandjean et al. 2004; Murata et al. 2004; Choi et al. 2009; Lim, Chung, and Paek 2010).

Heart rate variability (HRV) is an accepted method to assess changes in the autonomic nervous system during parasympathetic and sympathetic modulation of the cardiovascular system (HRV 1996). Heart rate is under constant influence of an interplay between parasympathetic and sympathetic modulation leading to slower heart rate at rest or during sleep, and faster heart rate with exercise or stress. Beat-to-beat variation in heart rate, or in duration of beat-to-beat intervals, could be measured reflecting changes in the autonomic control of the heart (Kleiger, Stein, and Bigger 2005). In healthy subjects, it is expected that there is a physiologic variation of on NN intervals (defined as normal beats with sinus rhythm, while excluding premature and ectopic beats) influenced by respiration, baroreflex sensitivity, in addition to the above mentioned parasympathetic and sympathetic modulation (Cygankiewicz and Zareba 2013). HRV might be altered in patients after myocardial infarction, patients with heart failure, or those with diabetes, obesity, frequently with a decrease in parasympathetic modulation and an increased sympathetic modulation (HRV 1996; El Agaty, Kirmani, and Labban 2017; Tsuji et al. 1996). These changes in HRV have been found as predictors of outcome (mortality, heart failure, ventricular tachyarrhythmias) in various patient populations (Kleiger et al. 1987; Bigger et al. 1992; Stein et al. 1997; Kleiger, Stein, and Bigger 2005; Sherazi et al. 2015). There is profound evidence that in addition to environmental factors like exposure to air pollution or lead, pharmacological or electrical therapies might alter HRV parameters indicating changes in the autonomic control of the heart (Coumel et al. 1991; Malik et al. 1989; Simula et al. 2017; Jerling et al. 2018; Hoogerwaard et al. 2019; Petersen et al. 2018; Breitner et al. 2019; Gump et al. 2011). The guidelines regarding HRV analysis and its parameters recommended for research and clinical use were established in 1996 (HRV 1996). However, over the last 20 years there still have been inconsistencies regarding HRV methodology and interpretation (Shaffer and Ginsberg 2017).

Some epidemiologic studies have suggested that MeHg exposure may be associated with a decrease in HRV parameters but with general inconsistency of findings among studies, (Grandjean et al. 2004; Mozaffarian 2009; Oka et al. 2002; Sorensen et al. 1999) and some

were not able to confirm it (Valera, Dewailly, and Poirier 2008; Miller et al. 2018; Gump et al. 2017; Choi et al. 2009) These studies have generally been based on short term HRV recordings at rest, and some were done in populations with co-exposures. Varying duration of recordings, diverse ECG sampling rate, different management of ectopic beats may influence results of HRV analyses (Shaffer and Ginsberg 2017; Karita et al. 2018; Stapelberg et al. 2018; Melo et al. 2018). These factors limit our understanding of whether there are systematic and direct cardiovascular effects of MeHg exposure (Karita et al. 2018). As recently summarized in two extensive reviews on the topic (Gribble et al. 2015; Karita et al. 2018) it is still uncertain whether MeHg exposure from repeated consumption of fish with naturally occurring background levels has any adverse and meaningful effect on HRV.

In a prospective study of 19-year-old healthy subjects, we assessed whether the prenatal and recent MeHg exposure from consuming fish adversely affects HRV in comprehensive analyses of 24-hour Holter recordings and short-term 5-minute recordings reflecting different physiologic conditions associated with resting supine, standing, mental stress (mathematics test), and sleeping.

METHODS

Study Population and Design

The Republic of Seychelles is a 115-island archipelago in the middle of the Indian Ocean where citizens consume large amounts of ocean fish. The population is exposed to MeHg primarily from fish consumption and does not consume sea mammals (Cernichiari et al. 1995; Davidson et al. 1998). Fish consumed in Seychelles are contaminated only by natural background exposure to MeHg. This setting provides a unique opportunity to study the association of MeHg with health outcomes. In 1989-90, 779 mother-infant pairs were recruited as part of the Seychelles Child Development Study (SCDS) main cohort (Marsh et al. 1995). The demographics and earlier evaluations of this cohort have been reported previously (Davidson et al. 1998; Shamlaye et al. 1995; Myers et al. 2003; van Wijngaarden et al. 2013).

For this study, a total of 518 cohort participants returned for evaluations at the Study Centre at a mean $\pm$ SD of 19.0 ± 0.3 years of age. Data from four of these participants were not included due to technical errors in ECG data acquisition. The evaluation consisted of a questionnaires collecting demographic, socioeconomic, medical history, and lifestyle data, collection of biological samples (hair and blood samples), and Holter ECG recordings providing time- and frequency-domain HRV parameters. The study protocol was approved by the Institutional Review Boards of the Republic of Seychelles and the University of Rochester.

MeHg and Polyunsaturated Fatty Acids Measurements

Prenatal MeHg exposure was determined by measurement of total maternal Hg levels in hair in samples collected closed to the scalp after pregnancy. It reflected mercury concentrations during pregnancy assuming a hair growth rate of 1 cm per month. Recent MeHg exposure was available for 451 (87.7%) of the participants; 68% of those missing this variable were

males, many of whom had shaved heads. Recent MeHg was analyzed in the 1 cm hair samples collected closest to the scalp taken at the time of Holter testing (Myers et al. 2009). Hair analysis was performed using cold vapor atomic absorption spectroscopy (Magos and Clarkson 1972). The detailed description of hair sampling, analysis and quality control procedures was previously described (Cernichiari et al. 1995). It has been documented that mercury hair levels recapitulate mercury exposure since hair concentrations correlate well with blood mercury levels (Clarkson, Magos, and Myers 2003).

Recent status of n-3 and n-6 PUFA were determined from plasma samples taken at the time of evaluation. Samples were processed, aliquoted, and stored at -80°C in the Ministry of Health's Clinical Laboratories and transported to the nutrition biochemistry laboratory in Coleraine, Northern Ireland for fatty acid analysis. Plasma samples were subjected to lipid extraction using an adaptation of the method of Folch et al. (Folch, Lees, and Sloane Stanley 1957) and solid phase extraction columns (Merck Lichrolut, UK) were used to isolate phospholipids. Fatty acid methyl esters were prepared with boron trifluoride in methanol (Sigma-Aldrich Co. Ltd.) and analyzed using GC-MS technology (5975C series, Agilent Technologies Ltd., Stockport, UK). Fatty acids were identified by their retention time with reference to commercially obtained fatty acid standards and were quantified by use of an internal standard C17:0 (Sigma Aldrich Co. Ltd.). The GC column used was a BPX 70 capillary column (Phenomenex, Cheshire, UK) with a length of 100 m, internal diameter of 0.25 mm and a capillary thickness of 0.25 μm .

Holter ECG Recordings

All 514 participants underwent 12-lead high-resolution Holter ECG recordings which were initiated between 8-10 am using digital (1000 Hz) recorders (H-Scribe, Mortara Instruments Inc., Milwaukee, WI). ECG recordings started with a 10-minute supine position followed by a 10-minute standing and subsequently a 10-minute ECG while the participant was completing mathematics (math) test. The supine recording served to analyze HRV parameters in the resting non-active state (parasympathetic dominance), the standing recording provided insight into the response of the heart rate and HRV parameters to activation of the autonomic system by changing body position (sympathetic activation), and the recording during a mathematical test reflected the effect of mental stress on the studied ECG parameters (Tsuji et al. 1996; Umetani et al. 1998). The Holter recordings were finished after 1-2 hours in 312 subjects, but in the remaining 202 subjects recordings were continued to obtain long-term overnight data to further characterize the normal physiologic predominance of parasympathetic modulation of the autonomic nervous system during sleep and provide insight into the overall long-term values of time-domain HRV parameters.

All recordings were sent to and analyzed at the University of Rochester Medical Center's ECG Core Laboratory by technicians blinded to participants' demographic and methylmercury exposure data. Routine beat annotation was performed identifying normal sinus beats and abnormal beats (atrial and ventricular ectopic beats, noise) and only normal sinus beats were included in the analyses. In the case of ectopic beats, the sinus normal-to-normal beat (NN) interval following the ectopic beat was not included. All recordings were verified by a cardiologist (WZ).

Heart Rate Variability

From short-term recordings obtained in supine and standing positions and during the math test, 5-minute ECG recordings were extracted starting 3 minutes from the beginning of each segment to allow for adaptation to the changed conditions. A 5-minute segment from the overnight Holter recording was also analyzed at approximately 2 am to represent HRV during sleep.

Annotated NN intervals were used to compute HRV parameters for a given segment of data. The time-domain HRV parameters (Kleiger, Stein, and Bigger 2005) that were obtained from both the short (~5-minute) and long recordings included: SDNN (standard deviation of normal-to-normal sinus beat intervals) reflecting overall HRV and rMSSD intervals (root mean square of successive differences in NN intervals). We also computed: pNN50 (percentage of NN intervals differing more than 50 ms from the preceding beat) reflecting parasympathetic modulation, SDANN (standard deviation of averaged 5-min normal-to-normal RR intervals), and SDNIX (mean of the SDs of all normal-to-normal RR intervals) for all 5-min segments of the entire recording (SDNNIX). These reflect overall HRV and are similar to SDNN, which is why we elected to focus on SDNN and rMSSD as representative HRV parameters in time-domain analysis.

The frequency-domain HRV methods (Bigger et al. 1992) allow for identifying the relative contribution of specific frequency bands reflecting oscillatory behavior of the heart rate. The total power (TP) of the entire spectrum was measured from 0 to 0.40 Hz. Other components included high frequency (HF: 0.15-0.40 Hz), low frequency (LF: 0.04-0.15 Hz), and very low frequency (VLF: 0.003-0.04 Hz) bands. The HF power (expressed in normalized units as HF/[TP-VLF]) represents parasympathetic (respiratory) modulation of the heart rate whereas LF power (expressed in normalized units as LF/[TP-VLF]) reflects modulation of the heart rate by both sympathetic and parasympathetic tones, but with strong dominance of sympathetic influence and baroreflexes. We also evaluated LF/HF ratio but elected to focus on LF and HF separately to illustrate effects of MeHg on individual components of these variables.

Our primary statistical analysis focused on 23 of the most commonly used HRV parameters reflecting autonomic activity in different physiologic conditions: 5 in supine position, 5 in standing position, 5 in response to a mathematics test, 5 during sleep, and 3 for the entire recording. Five outcomes measured at baseline while supine at rest included: NN, SDNN, rMSSD, LF, and HF. We also analyzed the five outcomes measured at baseline minus their values while standing, taking a math test, and sleeping. Unless otherwise noted, response variables are defined as differences from baseline values. Three primary outcomes measured on the long recordings included: NN, SDNN, rMSSD. Additionally, we computed three more HRV parameters (pNN50, SDANN, and SDNNIX) for secondary analysis by MeHg categories.

Statistical Analyses

We used linear regression to examine the association of each exposure (prenatal and recent MeHg, in separate models) and each of the 23 HR variables. Models first examined the

association with a sex by exposure interaction term since significant differences in HRV are known to be present between males and females, and additionally, exposure differed by sex (Tsuji et al. 1996; Umetani et al. 1998). If the interaction was statistically significant ($p < 0.05$) then those results are reported. If the interaction was not significant ($p \geq 0.05$) then the analysis was rerun without the interaction term and those results are reported. Primary models were fit with and without covariate adjustment for sex, birth weight, and current BMI. We adjusted for birth weight due to its potential impact on childhood development and for BMI due to evidence of an association between ECG variables and obesity (Antelmi et al. 2004). For each analysis, we used a two-sided test with $\alpha = 0.05$ to assess significance. For the association of recent MeHg exposure with HRV we fit secondary models adjusting for recent n-3 and n-6 PUFA and for the participant's activity level. Physical activity level is a main predictor of HRV (Fisher, Young, and Fadel 2015). The Global Physical Activity Questionnaire (GPAQ) was developed by WHO for physical activity surveillance. The GPAQ standardizes level of total physical activity measured in MET-minutes/week categorizing the value <600 as low sedentary activity, $600-3000$ as moderate activity, and >3000 as high activity. The activity measurement was based on the participants self-report of their minutes of physical activity per week using three domains (work, walking, and leisure) and was treated as a categorical variable with three levels (<600 , $600-3,000$, and $>3,000$ minutes) following the WHO recommendation. We adjusted for n-3 and n-6 PUFA because of their reported cardiac effects (Christensen et al. 1999). Model assumptions were checked using standard methods, including checking for constant variance, nonlinearity, and normally distributed residuals (Weisberg 2005). In some cases, there was evidence of assumption violations which necessitated a log transformation of the outcome. For variables that required a logarithmic transformation, differences from baseline values were calculated after the log transform of both variables (e.g. the outcome is the logarithm of the ratio of values, rather than the logarithm of the difference from baseline). After transformations when required, there was no evidence of extreme outliers or highly influential observations. We report the individual associations and corresponding betas and p values from the primary analysis. Additionally, we applied the Bonferroni correction to account for multiple comparisons. Since we focused on 23 primary HRV parameters measured (5 in supine position, 5 in standing position, 5 in response to math test, 5 during sleep and 3 for the entire recording), we considered a p value of $0.05/23 = 0.0022$ as significant after this correction.

We also performed a univariate comparison of studied HRV parameters by MeHg levels categorized as 0 to <5 ppm, 5 to <10 ppm, and ≥ 10 ppm in prenatal and recent exposure. Median value of MeHg level in hair of US females 16-49 years of age in the NHANES 1999-2000 was $0.19 \mu\text{g/g}$ with 95th percentile of $1.73 \mu\text{g/g}$, reaching $2.75 \mu\text{g/g}$ in individuals eating fish 3 times in the past 30 days (McDowell et al. 2004). Prenatal assessments of women from the Seychelles high fish consumption population reported mean $\pm$ SD total hair MeHg of 6.85 ± 4.5 ppm and a median value of 5.94 ppm (Myers et al. 2003). Therefore, the cutoff of 0-5ppm was chosen as reflecting low levels of MeHg. The median total hair Hg level of women in the Faroes birth cohort study was $4.5 \mu\text{g/g}$; 12% had levels $>10 \mu\text{g/g}$, considered as elevated (Grandjean et al. 2004). Therefore, we proposed the above arbitrary

cutoffs reflecting low, medium, and high levels of MeHg in the Seychelles fish eating populations.

RESULTS

Descriptive Analysis: Characteristics of Participants

There were 514 eligible participants (273 females and 241 males) who had short Holter recordings and 203 participants (117 women and 86 men) who had long Holter recordings. Table 1 shows the participants clinical characteristics, MeHg levels, and n-3 and n-6 PUFA levels compared by sex. The mean prenatal MeHg exposures were similar in males and females (6.77 vs 7.06 ppm, respectively), but recent MeHg exposures at 19 years of age were significantly higher in males than females (12.16 vs. 8.67 ppm, respectively). Males were physically more active and had significantly lower BMI values than females. The n-3 and n-6 PUFA levels did not differ by sex. Correlation between prenatal Hg and recent Hg (obtained at the age of 19 years) was weak: the correlation coefficient was 0.13.

As expected, time-domain SDNN and rMSSD parameters from long-term recordings and short-term supine recordings were significantly ($p < 0.0022$) different between males and females (Supplementary Table S1). In supine resting position, males had significantly longer NN interval (lower heart rate) than females. Males also had significantly higher levels of long-term and baseline supine time-domain SDNN and rMSSD parameters, but response to standing, math test, and sleep did not significantly differ by sex. Differences in frequency-domain HRV parameters between males and females were less pronounced and did not reach $p < 0.0022$ level.

Heart Rate Variability Parameters by Categorized MeHg Exposure Levels

Table 2 shows a univariate comparison of studied HRV parameters by MeHg levels categorized as 0 to <5 ppm (low), 5 to <10 ppm (medium), and 10 ppm (high) in prenatal and recent exposure (Table 2). For prenatal MeHg exposure this comparison did not show any significant associations indicating level-related changes in HRV parameters in long term recordings, baseline supine or response to standing, math and sleep. For recent MeHg categories there were few statistically borderline associations (p value between 0.0022 and 0.05) which were driven by lower values in medium than low and high MeHg exposure: NN in response to standing and math test, LF in response to sleep, and HF at baseline, in response to math and sleeps. Among these associations the direction was inconsistent and there was no evidence of a dose response. Since we observed sex-related differences in HRV parameters we reanalyzed 24-hour HRV data for males and females separately using the primary time domain parameters (NN, SDNN, rMSSD) and 3 additional ones (pNN50, SDANN, & SDNNIX). No significant associations were present in these HRV parameters among subject with low, medium, and high MeHg exposure (Supplementary Table S2).

Regression Analysis: Prenatal MeHg Exposure and Heart Rate Variability

The sex by prenatal exposure interaction was significant for the NN interval long recording and the NN in response to sleep (Table 3). After adjusting for sex, birth weight, and BMI, there were two borderline significant associations at p value between 0.0022 and 0.05. Both

were in males; and one was beneficial and the other adverse. The NN interval in the long recording was predicted to increase in males by 5.5 ms per 1 ppm increase in MeHg ($p=0.006$). This beneficial association reflected a 0.5 beat per minute decrease in heart rate per 1 ppm increase in MeHg, or a 3.4 beat per minute decrease per IQR increase in MeHg. The NN interval in response to sleep decreased only in males by 10.49 ms per 1 ppm increase in MeHg ($p = 0.0055$), reflecting about 0.5 a beat per minute increase in heart rate, physiologically an adverse association. When the Bonferroni correction was applied, none of the associations reached statistical significance ($p<0.0022$). There were no other significant interactions or associations of prenatal MeHg with either the time- or frequency-domain HRV parameters computed from the long or short recordings.

Regression Analysis: Recent MeHg Exposure at 19 years and Heart Rate Variability

Recent MeHg exposure was associated with four outcomes in models that adjusted for sex, BMI, and birth weight (Supplementary Table S3). Only one sex by MeHg interaction was borderline significant at p between 0.0022 and 0.05, the NN interval in response to standing ($p = 0.015$). That association was only in females and was adverse ($p = 0.0172$). For the NN interval in response to standing, females had a 3 ms increase in the difference in the NN interval per 1 ppm increase of MeHg ($p = 0.017$), or a 20.8 ms increase in the difference in the NN interval for an interquartile range (IQR). The associations with the two most classical time- domain parameters: SDNN baseline supine and on the long recording were not significant ($p=0.600$ and 0.128 respectively). Similarly, the associations with rMSSD resting supine and on the long recording were not significant ($p=0.197$ and 0.489 , respectively). For the difference in log (rMSSD) in response to a math test, there was a 0.009 ms increase for each 1 ppm increase in MeHg, a beneficial response. Stated differently, a 1 ppm increase in exposure was associated with a 1% increase in the ratio of rMSSD at baseline to the rMSSD during the math test, or an IQR increase in exposure was associated with a 6.4% increase in this ratio. For HF in response to standing both sexes together had a 0.40% increase in difference from baseline ($p=0.029$), and for HF in response to the math test, they had a 0.375% increase in difference from baseline ($p=0.032$), indicating lower parasympathetic parameters during these tests. These associations were inconsistent in direction, of small magnitude, nonsignificant after Bonferroni correction, and of uncertain meaning.

After additional adjustment for activity levels and n-3 and n-6 PUFA, recent MeHg exposure had borderline significant sex by recent MeHg interactions in two models: the NN interval in response to standing and the LF in response to the math test (Table 4). The association between MeHg and NN interval in response to standing was borderline at $p=0.0499$ only in females who had on average a 2.6 ms increase per 1 ppm increase in recent MeHg. There was also a borderline ($p=0.0210$) association between MeHg and LF in response to the math test for males only. Each 1 ppm increase in exposure was associated with a 0.56 ms decrease in the difference in LF (a beneficial effect).

Of the six significant borderline significant associations (at P between 0.0022 and 0.05) associations with the combined sexes, three were also present in the primary analysis (the difference in rMSSD on the logarithmic scale in response to the math test and the difference

in HF in response to standing and to the math test). The association with the difference in rMSSD in response to the math test was virtually unchanged, and the associations with HF responses to standing and the math test were both slightly attenuated. The three additional outcomes with borderline significant MeHg associations were the NN interval for the long recording, the baseline supine NN, and the LF in response to sleep. For the baseline supine NN, each 1 ppm increase in recent MeHg was associated with on average a decrease of 3.05 ms, an adverse association. For the LF in response to sleep, each 1 ppm increase in recent MeHg was associated with a 0.69 decrease (beneficial effect). The inconsistency of these associations and their small magnitude (milliseconds) make them challenging to interpret. After applying the Bonferroni correction for multiplicity, there were no statistically significant MeHg associations (all p values >0.0022).

DISCUSSION

We studied whether HRV parameters are associated with prenatal or recent MeHg exposure. We measured a comprehensive set of HRV parameters obtained from short-term (5 minutes) and long-term (24 hours) Holter recordings in a cohort of young adults exposed to MeHg from frequent consumption of open ocean fish. Their prenatal MeHg exposures were approximately 10 times more than those present in the United States. We focused on 23 measured HRV variables and found none of them significant at prespecified after Bonferroni correction with $p < 0.0022$. There were two borderline significant (at p between 0.0022 and 0.05) associations with prenatal and eight borderline significant associations with recent MeHg exposure in secondary models. The two prenatal associations were present only in males and were in opposite directions suggesting random variation. The eight recent MeHg exposure significant associations were all of small magnitude, and there was no consistent pattern that would suggest they are related to exposure. Following Bonferroni correction for multiplicity, there were no statistically significant (at $p < 0.0022$) associations with either prenatal or recent MeHg exposure.

We observed the expected sex differences in HRV parameters that have been previously reported in the literature. Males showed significantly higher values of HRV parameters than females (Agelink et al. 2001; Umetani et al. 1998), and the responses to standing and other conditions were as expected (Srinivasan, Sucharita, and Vaz 2002). These findings in a large cohort provide reassurance that there was sufficient statistical power to detect meaningful associations of exposures and HRV if they were present. The borderline significant findings were of very small magnitude without clear physiological or clinical meaning. Previously, we measured beat to beat blood pressure in 95 participants from this cohort, using the Finapres to record the baroreflex, another measure of autonomic nervous system integrity (Periard et al. 2015). That study found no association of blood pressure variability with either prenatal or recent MeHg levels. These data do not support the hypothesis that prenatal or recent exposure to MeHg from fish consumption influences HRV parameters.

The categorical analyses (Table 2 and Supplemental Table S2), although not adjusted for covariates, further corroborate the absence of associations between prenatal and recent MeHg exposure and the NN interval and other HRV parameters among the three exposure groups. If there was an association, one would expect some MeHg level-dependent changes

in HRV parameters, but this categorical approach showed no trend toward any association, i.e. no decrease or increase in studied parameters across incremental levels of MeHg. At the same time, we were able to detect significant differences as we documented by detecting expected sex-related differences in HRV parameters (positive control test).

Prenatal MeHg exposure was associated with about a 0.5 beat per minute slower heart rate in males per 1 ppm increase in MeHg. This change does not have clinical meaning considering that exposure to MeHg in Western societies is generally less than 1 ppm. Even in high fish-consuming societies like Japan, MeHg exposure is generally less than 2 ppm which would be associated with a potential variation in heart rate of approximately 1 beat per minute. Neither time-domain nor frequency-domain HRV parameters were associated with prenatal MeHg levels. Prenatal MeHg does not appear to be influencing parameters or autonomic nervous system activity reflected by HRV at the age of 19 years in this cohort.

Several HRV parameters showed borderline significant associations after adjustment for birth weight, BMI, activity level, and n-3 and n-6 PUFA levels. These associations were all of small magnitude, inconsistent across parameters, and there was no indication of a dose response curve. No associations were significant after Bonferroni adjustment. Additionally, it seems most likely that if recent exposure to low levels of MeHg does impact the nervous system, it would be associated with prolonged cumulative dosage rather than the one month value that our measure determined. Considering all of these factors, we do not believe these data provide support for the hypothesis that HRV parameters are associated with recent MeHg exposure.

In analyses adjusted for PUFA and activity level, heart rate assessed during supine, standing, and math test periods increased with increasing MeHg levels with borderline significance. A 3-millisecond decrease in NN interval translates to about 0.2- 0.3 beats per minute increase in heart rate, and has no clinical relevance. Similarly rMSSD and HF decreased with increasing recent MeHg during the math test and LF/HF with standing. These changes were very small and not significant clinically or statistically after Bonferroni correction. In response to a math test, there was a 0.010 ms decrease in rMSSD per 1 ppm increase in MeHg, which despite borderline $p=0.006$ (after Bonferroni correction) is of no clinical or physiologic meaning.

Roman and colleagues reviewed the literature on associations of prenatal and recent MeHg with cardiovascular outcomes (Roman et al. 2011). They noted that evidence of impact at dose levels of interest was lacking and that the "...specific measures of HRV in each study vary." However, based on the consistency of the epidemiological literature, they classified the evidence supporting the association of HRV with MeHg exposure as "strong" compared to other cardiovascular measures. This reasoning is tenuous when one considers the known bias against publishing negative studies. The European Food Safety Authority ((CONTAM) 2012) reviewed reports examining MeHg and HRV in 2012 with subsequent update in 2018 and the panel concluded that, although some studies suggest an autonomic effect of MeHg, the results were inconsistent among studies and the implication for health were unclear.

Recently, Japanese researchers (Karita et al. 2018) reviewed 13 studies examining the association between MeHg and HRV parameters and 8 of them showed the significant association with MeHg and 5 failed to demonstrate any significant association. Earlier studies that reported an association between prenatal MeHg exposure and HRV parameters varied from the current study in a number of ways. They were conducted in a variety of populations and measured HRV variables using different methods and recording times. Also, some populations studied had significant exposures to other toxicants that might also influence HRV.

Discussing studies supporting the MeHg and HRV associations, the Faroe Islands investigators reported a decrease in the coefficient of variation of RR (a HRV parameter similar to SDNN) and in LF and HF levels as prenatal MeHg increased (Sorensen et al. 1999; Grandjean et al. 2004). Their studies differed in that Hg exposure was primarily from whale consumption and were thus co-exposed to polychlorinated biphenyls. In addition, the ECG recordings were short. The largest cohort studied to date evaluated 1,589 Koreans living near an industrial complex. Participants ranged in age from 5 to 83 years with 389 below age 20 years, had mean hair Hg levels that were low (0.83 ug/gm), and had 5-minute ECG recordings obtained in sitting position, which affect results (Lim, Chung, and Paek 2010). When categorized by decades of life, the 104 subjects below age 10 years had an 8.4% decrease in the HF parameter with each 1 ppm increase in hair mercury concentration (95% confidence interval: 2.2–15.1). In a study of French Polynesians aged 12–17 years (Valera et al. 2011), significant differences were observed in LF/HF ratio, LF, and HF between the second (7.9–10.0 g/L) and third (11.0–26.0 g/L) tertiles of blood mercury concentration. That study reported no associations between HRV parameters and MeHg exposures in the 180 adults studied. In the only intervention study (Yaginuma-Sakurai et al. 2010), a significant difference in LF was observed between the experimental group (mean mercury levels of 8.76 µg/g in hair and 26.9 µg/L in blood) and control group, but one cannot exclude the effect of concomitant changes in levels of n-3 PUFA influenced these results.

Discussing studies with no clear evidence of the MeHg-HRV association, analysis of Faroese whaling men showed that blood mercury level was associated with increased coefficient of variation of RR intervals, and coefficient of variation of LF, but latent mercury level, estimated from mercury levels in blood, toe nail, and hair was not significantly associated with any HRV parameter (Choi et al. 2009). In Inuit adults (Valera, Dewailly, and Poirier 2008), blood mercury was significantly correlated with coefficient of variation of RR interval and LF in univariate analyses, but these associations were not significant after adjusting for confounders. In fish consumers from Long Island (Miller et al. 2018), HRV parameter in a multiple regression analysis were not significant after adjustment for serum docosahexaenoic acid and eicosapentaenoic acid levels. In the study of 203 children aged 9–11 years (Gump et al. 2017), LF, HF, and LF/HF ratio at rest and during stress were not significantly associated with blood mercury levels.

As summarized above the effect of MeHg on HRV parameters does not seem consistent and remains uncertain. Some studies indicate changes in the sympathetic (LF) component, some in parasympathetic (HF, rMSSD), and some in both. Recording time and conditions varied

among the studies as did MeHg assessments. Competing environmental pollutions such as persistent organic pollutants from consuming ocean mammals (Sorensen et al. 1999; Valera et al. 2011), and underlying comorbidities such as in the Korean study (Lim, Chung, and Paek 2010), might also have influenced the findings. If there is a causal association of prenatal or recent MeHg exposure with changes in the autonomic nervous system, one would expect consistency in the directionality of the associations for both classical time-domain and frequency-domain HRV parameters. No study has demonstrated this. Our study with a large cohort and a wide range of prenatal and recent MeHg exposures does not confirm the earlier reports.

We want to stress that none of the studies published to date in the literature on MeHg association with HRV parameters has used 24-hour recording, the most standard approach to evaluate HRV in cardiovascular literature. The strengths of this study consist of having accurate prenatal and recent MeHg exposure measures at 19 years of age, a large cohort whose MeHg exposure is significantly higher than other reported cohorts, a comprehensive battery of HRV parameters completed under controlled conditions over a prolonged time period, blinding of clinicians to exposure, and *a priori* specified analysis plans. Additionally, the analyses identified known associations of HRV parameters indicating that if MeHg did influence autonomic control of HRV the study could have detected it. As with all epidemiological studies, there are also limitations. Complete data were not available on all subjects, conditions during the overnight recordings were not monitored, and we were not able to complete long recordings on all participants. Furthermore, our measure of recent exposure was limited to the month prior to evaluation.

In conclusion, in a large cohort of 19 year old participants with MeHg exposures significantly above those in previous reports, we found variations in HRV parameters related to sex that have been previously reported, but found no association of them with prenatal MeHg exposure. We found some trends toward associations of recent (at the age of 19 years) MeHg exposure with heart rate and HRV parameters when subjects were challenged with math test that could suggest that concurrent MeHg exposure might influence autonomic response, but the magnitude of these findings is so miniscule that it could not be considered meaningful from a clinical standpoint. In addition, we are cautious with this interpretation because of the multiple comparisons and small magnitude of the associations. We do not believe that these data support the hypothesis that either prenatal or recent MeHg exposure from consuming naturally contaminated ocean fish adversely influences heart rate variability in young adults.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

Study was funded by grants RO1-ES008442, P30-ES01247, and T32-ES007271 from the National Institute of Environmental Health Sciences to University of Rochester Medical Center and by the Government of the Republic of Seychelles.

References

- (CONTAM), EFSA Panel on Contaminants in the Food Chain. 2012 'Scientific Opinion on the risk for public health related to the presence of mercury and methylmercury in food. EFSA', EFSA Journal, 10: 241.
- Agelink MW, Malessa R, Baumann B, Majewski T, Akila F, Zeit T, and Ziegler D. 2001 'Standardized tests of heart rate variability: normal ranges obtained from 309 healthy humans, and effects of age, gender, and heart rate', Clin Auton Res, 11: 99–108. [PubMed: 11570610]
- Antelmi I, de Paula RS, Shinzato AR, Peres CA, Mansur AJ, and Grupi CJ. 2004 'Influence of age, gender, body mass index, and functional capacity on heart rate variability in a cohort of subjects without heart disease', Am J Cardiol, 93: 381–5. [PubMed: 14759400]
- Bigger JT Jr., Fleiss JL, Steinman RC, Rolnitzky LM, Kleiger RE, and Rottman JN. 1992 'Frequency domain measures of heart period variability and mortality after myocardial infarction', Circulation, 85: 164–71. [PubMed: 1728446]
- Breitner S, Peters A, Zareba W, Hampel R, Oakes D, Wiltshire J, Frampton MW, Hopke PK, Cyrus J, Utell MJ, Kane C, Schneider A, and Rich DQ. 2019 'Ambient and controlled exposures to particulate air pollution and acute changes in heart rate variability and repolarization', Sci Rep, 9: 1946. [PubMed: 30760868]
- Cernichiari E, Toribara TY, Liang L, Marsh DO, Berlin MW, Myers GJ, Cox C, Shamlaye CF, Choisy O, Davidson P, and et al. 1995 'The biological monitoring of mercury in the Seychelles study', Neurotoxicology, 16: 613–28. [PubMed: 8714867]
- Choi AL, Weihe P, Budtz-Jorgensen E, Jorgensen PJ, Salonen JT, Tuomainen TP, Murata K, Nielsen HP, Petersen MS, Askham J, and Grandjean P. 2009 'Methylmercury exposure and adverse cardiovascular effects in Faroese whaling men', Environ Health Perspect, 117: 367–72. [PubMed: 19337510]
- Christensen JH, Christensen MS, Dyerberg J, and Schmidt EB. 1999 'Heart rate variability and fatty acid content of blood cell membranes: a dose-response study with n-3 fatty acids', Am J Clin Nutr, 70: 331–7. [PubMed: 10479194]
- Clarkson TW, Magos L, and Myers GJ. 2003 'The toxicology of mercury--current exposures and clinical manifestations', N Engl J Med, 349: 1731–7. [PubMed: 14585942]
- Coumel P, Hermida JS, Wennerblom B, Leenhardt A, Maison-Blanche P, and Cauchemez B. 1991 'Heart rate variability in left ventricular hypertrophy and heart failure, and the effects of beta-blockade. A non-spectral analysis of heart rate variability in the frequency domain and in the time domain', Eur Heart J, 12: 412–22. [PubMed: 1828232]
- Cygankiewicz I, and Zareba W. 2013 'Heart rate variability', Handb Clin Neurol, 117: 379–93. [PubMed: 24095141]
- Davidson PW, Myers GJ, Cox C, Axtell C, Shamlaye C, Sloane-Reeves J, Cernichiari E, Needham L, Choi A, Wang Y, Berlin M, and Clarkson TW. 1998 'Effects of prenatal and postnatal methylmercury exposure from fish consumption on neurodevelopment: outcomes at 66 months of age in the Seychelles Child Development Study', JAMA, 280: 701–7. [PubMed: 9728641]
- El Agaty SM, Kirmani A, and Labban E. 2017 'Heart rate variability analysis during immediate recovery from exercise in overweight/obese healthy young adult females', Ann Noninvasive Electrocardiol, 22.
- FAO. 2016 'The State of World Fisheries and Aquaculture 2016. Contributing to food security and nutrition for all. .' In, 1–200. Rome.
- Fisher JP, Young CN, and Fadel PJ. 2015 'Autonomic adjustments to exercise in humans', Compr Physiol, 5: 475–512. [PubMed: 25880502]
- Folch J, Lees M, and Sloane Stanley GH. 1957 'A simple method for the isolation and purification of total lipides from animal tissues', J Biol Chem, 226: 497–509. [PubMed: 13428781]
- Grandjean P, Murata K, Budtz-Jorgensen E, and Weihe P. 2004 'Cardiac autonomic activity in methylmercury neurotoxicity: 14-year follow-up of a Faroese birth cohort', J Pediatr, 144: 169–76. [PubMed: 14760255]
- Gribble MO, Cheng A, Berger RD, Rosman L, and Guallar E. 2015 'Mercury Exposure and Heart Rate Variability: a Systematic Review', Curr Environ Health Rep, 2: 304–14. [PubMed: 26231507]

- Gump BB, Dykas MJ, MacKenzie JA, Dumas AK, Hruska B, Ewart CK, Parsons PJ, Palmer CD, and Bendinskas K. 2017 'Background lead and mercury exposures: Psychological and behavioral problems in children', *Environ Res*, 158: 576–82. [PubMed: 28715786]
- Gump BB, Mackenzie JA, Bendinskas K, Morgan R, Dumas AK, Palmer CD, and Parsons PJ. 2011 'Low-level Pb and cardiovascular responses to acute stress in children: the role of cardiac autonomic regulation', *Neurotoxicol Teratol*, 33: 212–9. [PubMed: 20934510]
- Hoogerwaard AF, de Jong MR, Adiyaman A, Smit JJJ, Pphm Delnoy JE Heeg Baam van Hasselt, A. R. Ramdat Misier, M. Rienstra, I. C. van Gelder, and A. Elvan. 2019 'Renal sympathetic denervation induces changes in heart rate variability and is associated with a lower sympathetic tone', *Clin Res Cardiol*, 108: 22–30. [PubMed: 29943270]
- HRV. 1996 'Heart rate variability. Standards of measurement, physiological interpretation, and clinical use. Task Force of the European Society of Cardiology and the North American Society of Pacing and Electrophysiology', *Eur Heart J*, 17: 354–81. [PubMed: 8737210]
- Jerling M, Cygankiewicz I, Al-Tawil N, Darpo B, Ljungstrom A, and Zareba W. 2018 'Effects of intranasal kinetic oscillation stimulation on heart rate variability', *Ann Noninvasive Electrocardiol*, 23.
- Karita K, Iwata T, Maeda E, Sakamoto M, and Murata K. 2018 'Assessment of Cardiac Autonomic Function in Relation to Methylmercury Neurotoxicity', *Toxics*, 6.
- Kleiger RE, Miller JP, Bigger JT Jr., and Moss AJ. 1987 'Decreased heart rate variability and its association with increased mortality after acute myocardial infarction', *Am J Cardiol*, 59: 256–62. [PubMed: 3812275]
- Kleiger RE, Stein PK, and Bigger JT Jr. 2005 'Heart rate variability: measurement and clinical utility', *Ann Noninvasive Electrocardiol*, 10: 88–101. [PubMed: 15649244]
- Lim S, Chung HU, and Paek D. 2010 'Low dose mercury and heart rate variability among community residents nearby to an industrial complex in Korea', *Neurotoxicology*, 31: 10–6. [PubMed: 19833149]
- Magos L, and Clarkson TW. 1972 'Atomic absorption determination of total, inorganic, and organic mercury in blood', *J Assoc Off Anal Chem*, 55: 966–71. [PubMed: 5071876]
- Malik M, Farrell T, Cripps T, and Camm AJ. 1989 'Heart rate variability in relation to prognosis after myocardial infarction: selection of optimal processing techniques', *Eur Heart J*, 10: 1060–74. [PubMed: 2606116]
- Marsh DO, Clarkson TW, Myers GJ, Davidson PW, Cox C, Cernichiari E, Tanner MA, Lednar W, Shamlaye C, Choisy O, and et al. 1995 'The Seychelles study of fetal methylmercury exposure and child development: introduction', *Neurotoxicology*, 16: 583–96. [PubMed: 8714865]
- Melo HM, Martins TC, Nascimento LM, Hoeller AA, Walz R, and Takase E. 2018 'Ultra-short heart rate variability recording reliability: The effect of controlled paced breathing', *Ann Noninvasive Electrocardiol*, 23: e12565. [PubMed: 29863781]
- Miller C, Karimi R, Silbernagel S, Kostrubiak D, Schiavone F, Zhang Q, Yang J, Rashba E, and Meliker JR. 2018 'Mercury, omega-3 fatty acids, and seafood intake are not associated with heart rate variability or QT interval', *Arch Environ Occup Health*, 73: 251–57. [PubMed: 28379067]
- Mozaffarian D 2009 'Fish, mercury, selenium and cardiovascular risk: current evidence and unanswered questions', *Int J Environ Res Public Health*, 6: 1894–916. [PubMed: 19578467]
- Murata K, Weihe P, Budtz-Jorgensen E, Jorgensen PJ, and Grandjean P. 2004 'Delayed brainstem auditory evoked potential latencies in 14-year-old children exposed to methylmercury', *J Pediatr*, 144: 177–83. [PubMed: 14760257]
- Myers GJ, Davidson PW, Cox C, Shamlaye CF, Palumbo D, Cernichiari E, Sloane-Reeves J, Wilding GE, Kost J, Huang LS, and Clarkson TW. 2003 'Prenatal methylmercury exposure from ocean fish consumption in the Seychelles child development study', *Lancet*, 361: 1686–92. [PubMed: 12767734]
- Myers GJ, Thurston SW, Pearson AT, Davidson PW, Cox C, Shamlaye CF, Cernichiari E, and Clarkson TW. 2009 'Postnatal exposure to methyl mercury from fish consumption: a review and new data from the Seychelles Child Development Study', *Neurotoxicology*, 30: 338–49. [PubMed: 19442817]

- Oka T, Matsukura M, Okamoto M, Harada N, Kitano T, Miike T, and Futatsuka M. 2002 'Autonomic nervous functions in fetal type Minamata disease patients: assessment of heart rate variability', *Tohoku J Exp Med*, 198: 215–21. [PubMed: 12630553]
- Periard D, Beqiraj B, Hayoz D, Viswanathan B, Evans K, Thurston SW, Davidson PW, Myers GJ, and Bovet P. 2015 'Associations of baroreflex sensitivity, heart rate variability, and initial orthostatic hypotension with prenatal and recent postnatal methylmercury exposure in the Seychelles Child Development Study at age 19 years', *Int J Environ Res Public Health*, 12: 3395–405. [PubMed: 25807149]
- Petersen KK, Andersen HH, Tsukamoto M, Tracy L, Koenig J, and Arendt-Nielsen L. 2018 'The effects of propranolol on heart rate variability and quantitative, mechanistic, pain profiling: a randomized placebo-controlled crossover study', *Scand J Pain*, 18: 479–89. [PubMed: 29858911]
- Roman HA, Walsh TL, Coull BA, Dewailly E, Guallar E, Hattis D, Marien K, Schwartz J, Stern AH, Virtanen JK, and Rice G. 2011 'Evaluation of the cardiovascular effects of methylmercury exposures: current evidence supports development of a dose-response function for regulatory benefits analysis', *Environ Health Perspect*, 119: 607–14. [PubMed: 21220222]
- Shaffer F, and Ginsberg JP. 2017 'An Overview of Heart Rate Variability Metrics and Norms', *Front Public Health*, 5: 258. [PubMed: 29034226]
- Shamlaye CF, Marsh DO, Myers GJ, Cox C, Davidson PW, Choisy O, Cernichiari E, Choi A, Tanner MA, and Clarkson TW. 1995 'The Seychelles child development study on neurodevelopmental outcomes in children following in utero exposure to methylmercury from a maternal fish diet: background and demographics', *Neurotoxicology*, 16: 597–612. [PubMed: 8714866]
- Sherazi S, Kutiyfa V, McNitt S, Aktas MK, Couderc JP, Peterson B, Bloch Thomsen PE, Kautzner J, Moss AJ, and Zareba W. 2015 'Prognostic Significance of Heart Rate Variability Among Patients Treated With Cardiac Resynchronization Therapy: MADIT-CRT (Multicenter Automatic Defibrillator Implantation Trial-Cardiac Resynchronization Therapy)', *JACC Clin Electrophysiol*, 1: 74–80. [PubMed: 29759342]
- Simula S, Laitinen TP, Laitinen TM, Hartikainen P, and Hartikainen JEK. 2017 'Sequence of cardiovascular autonomic alterations after fingolimod initiation', *Ann Noninvasive Electrocardiol*, 22.
- Sorensen N, Murata K, Budtz-Jorgensen E, Weihe P, and Grandjean P. 1999 'Prenatal methylmercury exposure as a cardiovascular risk factor at seven years of age', *Epidemiology*, 10: 370–5. [PubMed: 10401870]
- Srinivasan K, Sucharita S, and Vaz M. 2002 'Effect of standing on short term heart rate variability across age', *Clin Physiol Funct Imaging*, 22: 404–8. [PubMed: 12464145]
- Stapelberg NJC, Neumann DL, Shum DHK, McConnell H, and Hamilton-Craig I. 2018 'The sensitivity of 38 heart rate variability measures to the addition of artifact in human and artificial 24-hr cardiac recordings', *Ann Noninvasive Electrocardiol*, 23.
- Stein PK, Freedland KE, Skala JA, Carney RM, Davila-Roman V, Rich MW, and Kleiger RE. 1997 'Heart rate variability is independent of age, gender, and race in congestive heart failure with a recent acute exacerbation', *Am J Cardiol*, 79: 511–2. [PubMed: 9052363]
- Tsuji H, Larson MG, Venditti FJ Jr., Manders ES, Evans JC, Feldman CL, and Levy D. 1996 'Impact of reduced heart rate variability on risk for cardiac events. The Framingham Heart Study', *Circulation*, 94: 2850–5. [PubMed: 8941112]
- Umetani K, Singer DH, McCraty R, and Atkinson M. 1998 'Twenty-four hour time domain heart rate variability and heart rate: relations to age and gender over nine decades', *J Am Coll Cardiol*, 31: 593–601. [PubMed: 9502641]
- Valera B, Dewailly E, and Poirier P. 2008 'Cardiac autonomic activity and blood pressure among Nunavik Inuit adults exposed to environmental mercury: a cross-sectional study', *Environ Health*, 7: 29. [PubMed: 18538022]
- Valera B, Dewailly E, Poirier P, Counil E, and Suhas E. 2011 'Influence of mercury exposure on blood pressure, resting heart rate and heart rate variability in French Polynesians: a cross-sectional study', *Environ Health*, 10: 99. [PubMed: 22078280]
- van Wijngaarden E, Thurston SW, Myers GJ, Strain JJ, Weiss B, Zarcone T, Watson GE, Zareba G, McSorley EM, Mulhern MS, Yeates AJ, Henderson J, Gedeon J, Shamlaye CF, and Davidson PW.

2013 'Prenatal methyl mercury exposure in relation to neurodevelopment and behavior at 19 years of age in the Seychelles Child Development Study', *Neurotoxicol Teratol*, 39: 19–25. [PubMed: 23770126]

Weisberg S 2005 *Applied Linear Regression*. (John Wiley & Sons, Inc.: Hoboken, NJ).

WHO. 2007 "Exposure to mercury: A major public health concern ." In *Preventing disease through healthy environments*. Geneva, Switzerland: Public Health and Environment, World Health Organization.

Yaginuma-Sakurai K, Murata K, Shimada M, Nakai K, Kurokawa N, Kameo S, and Satoh H. 2010 'Intervention study on cardiac autonomic nervous effects of methylmercury from seafood', *Neurotoxicol Teratol*, 32: 240–5. [PubMed: 19732823]

Highlights

- High fish consumption might be associated with exposure to methylmercury.
- The effect of methylmercury on heart rate variability remains controversial.
- This is the most comprehensive assessment of HRV parameters to our knowledge among published studies focused on methylmercury toxicity
- In this study, methylmercury exposure from open ocean fish consumption did not adversely impact heart rate variability in young adults.

Table 1:

Characteristics of 19-year old subjects with comparison of males and females.

	All		Females		Males		P value
	N	Mean ± SD	N	Mean ± SD	N	Mean ± SD	
BMI (kg/m ²)	507	22.25 ± 4.83	270	22.93 ± 5.60	237	21.47 ± 3.6	0.06
Child's Birth Weight (Kg)	514	3.18 ± 0.5	273	3.12 ± 0.48	241	3.25 ± 0.51	0.01
Prenatal MeHg (ppm)	514	6.92 ± 4.54	273	7.06 ± 4.68	241	6.77 ± 4.37	0.65
Recent MeHg (ppm)	451	10.21 ± 5.79	253	8.67 ± 4.76	198	12.16 ± 6.37	<0.01
n-3 LCPUFA (mg/ml)	482	0.05 ± 0.02	256	0.05 ± 0.02	226	0.04 ± 0.02	0.48
n-6 LCPUFA (mg/ml)	482	0.15 ± 0.04	256	0.15 ± 0.04	226	0.15 ± 0.05	0.85
Activity Level (total MET):	N (%)	N (%)	N (%)				
Sedentary (< 600)	171 (33.6%)	137 (50.6%)	34 (14.3%)	<0.01			
Moderate (600 - < 3,000)	139 (27.3%)	87 (32.1%)	52 (21.9%)	0.01			
High (3,000+)	199 (39.1%)	47 (17.3%)	152 (63.9%)	<0.01			

Activity level: categorized from participants reports of their weekly walking, working, and recreational activities, in minutes estimated time per week (MET).

P values are from the nonparametric Wilcoxon rank-sum test

n-3 = sum of DHA, EPA, and ALA. n-6 = sum of AA and LA

Table 2.

Average heart rate variability measurements by categories of prenatal and recent (at 19 years of age) MeHg (ppm) exposure^a

Outcome	Prenatal MeHg			P value	Recent MeHg			P value [*]
	0<5 ppm (n=211)	5<10 ppm (n=184)	10+ ppm (n=119)		0<5 ppm (n=63)	5<10 ppm (n=201)	10+ ppm (n=187)	
	Mean±SD	Mean±SD	Mean±SD		mean±SD	mean±SD	mean±SD	
NN (ms)								
Long-term Recording	794±84	795±95	825±98	0.19	768±75	806±90	806±102	0.12
Baseline Supine	928±167	915±162	922±154	0.76	890±156	907±160	936±168	0.08
Response to Standing	157±99	149±94	160±92	0.43	152±99	141±93	168±96	0.02
Response to Math Test	65±72	67±80	81±87	0.28	78±84	59±71	80±87	0.04
Response to Sleep	-101±153	-110±146	-140±114	0.44	-112±130	-109±150	-118±142	0.98
SDNN (ms)								
Long-term Recording	181±40	181±421	182±39	0.97	173±44	180±41	184±40	0.31
Baseline Supine	83±39	80±37	82±36	0.74	83±39	78±34	86±41	0.39
Response to Standing	5±33	8±32	3±27	0.73	12±32	3±27	8±37	0.36
Response to Math Test	9±24	8±27	6±28	0.74	10±28	5±23	11±28	0.20
Response to Sleep	-11±40	-17±43	-17±38	0.64	-4±34	-12±38	-18±46	0.29
rMSSD (ms)								
Long-term Recording	66±28	63±33	64±24	0.52	64±25	63±25	69±35	0.88
Baseline Supine	77±42	73±44	74±42	0.46	76±44	71±41	79±46	0.37
Response to Standing	31±32	31±35	31±32	0.70	35±38	27±29	35±35	0.13
Response to Math Test	11±27	12±30	11±34	0.49	14±35	8±24	15±36	0.25
Response to Sleep	-10±36	-11±40	-18±32	0.60	-8±36	-15±36	-9±41	0.44
LF								
Baseline Supine	39±16	41±17	40±16	0.72	35±15	42±17	39±15	0.03
Response to Standing	-15±25	-15±23	-13±26	0.72	-17±24	-12±25	-17±24	0.23
Response to Math Test	-7±19	-4±22	-6±20	0.51	-8±19	-3±22	-8±18	0.23
Response to Sleep	3±22	8±21	8±22	0.33	0±18	10±22	3±23	0.03
HF								

Outcome	Prenatal MeHg			Recent MeHg			P value	P value *
	0-<5 ppm (n=211)	5-<10 ppm (n=184)	10+ ppm (n=119)	0-<5 ppm (n=63)	5-<10 ppm (n=201)	10+ ppm (n=187)		
	Mean±SD	Mean±SD	Mean±SD	mean±SD	mean±SD	mean±SD		
Baseline Supine	50±19	49±20	40±16	55±18	47±19	51±19		0.01
Response to Standing	29±21	26±21	-13±26	30±24	23±21	30±20		0.01
Response to Math Test	7±19	7±22	-6±20	10±20	4±20	10±20		0.10
Response to Sleep	-4±26	-3±27	8±22	0±27	-10±27	1±24		0.02

^ P = value is from 2-df nonparametric Kruskal Wallis ANOVA test

= Difference between baseline values and values obtained at standing, math test, or sleeping, respectively

* Please note that significant differences were observed when comparing middle MeHg dose group with lower and higher dose groups without dose-dependent effect.

Table 3.

Prenatal MeHg (ppm) exposure association with heart rate variability measurements, adjusting for sex, birth weight and BMI

Outcome	N	P value for sex interaction	β	P value	95% CI
NN (ms)					
Long-term Recording	202	0.0230	1.883	0.1160	(-0.4710, 4.2360)
female			-0.1729	0.9073	(-3.0974, 2.7516)
male			5.494	0.0058	(1.6105, 9.3775)
Baseline Supine	507	0.3050	-1.303	0.3470	(-4.0210, 1.4150)
Response to Standing	497	0.2717	-0.185	0.8420	(-2.0120, 1.6410)
Response to Math Test	442	0.5480	1.534	0.0620	(-0.0760, 3.1440)
Response to Sleep	202	0.0120	-2.915	0.2000	(-7.3900, 1.5600)
female			1.3949	0.6203	(-4.1492, 6.9390)
male			-10.4873	0.0055	(-17.8493, -3.1253)
SDNN (ms)					
Long-term Recording ⁺	200	0.1820	-0.001	0.7160	(-0.0080, 0.0050)
Baseline Supine ⁺	507	0.4077	-0.004	0.4120	(-0.0120, 0.0050)
Response to Standing [*]	497	0.8190	-0.001	0.7080	(-0.0090, 0.0060)
Response to Math Test [*]	442	0.9039	0.000	0.9360	(-0.0070, 0.0060)
Response to Sleep [*]	202	0.7592	-0.003	0.7310	(-0.0180, 0.0130)
rMSSD (ms)					
Long-term Recording ⁺	202	0.5522	-0.006	0.3470	(-0.0180, 0.0060)
Baseline Supine ⁺	499	0.2371	-0.008	0.1730	(-0.0190, 0.0030)
Response to Standing [*]	490	0.3928	0.002	0.6740	(-0.0070, 0.0110)
Response to Math Test [*]	434	0.3294	0.003	0.3760	(-0.0040, 0.0110)
Response to Sleep [*]	199	0.6891	-0.001	0.9150	(-0.0170, 0.0150)
LF					
Baseline Supine	505	0.9939	0.07	0.6610	(-0.2440, 0.3850)
Response to Standing	496	0.3730	0.037	0.8800	(-0.4390, 0.5120)
Response to Math Test	441	0.7007	-0.05	0.8130	(-0.4620, 0.3630)
Response to Sleep	201	0.6549	0.246	0.4730	(-0.4280, 0.9200)
HF					
Baseline Supine	505	0.4400	-0.064	0.7310	(-0.4300, 0.3020)
Response to Standing	496	0.3806	-0.12	0.5650	(-0.5310, 0.2910)
Response to Math Test	441	0.2875	0.131	0.5230	(-0.2700, 0.5320)
Response to Sleep	201	0.1498	-0.107	0.8010	(-0.9380, 0.7250)

⁺ = Model uses the natural logarithm of the outcome, and slopes are multiplicative effects

= Difference between baseline values and values obtained at standing, math test, or sleeping respectively

* = Difference in natural logarithms of baseline values and values obtained at standing, math test, or sleeping, respectively

If Male*MeHg interaction is NOT significant ($p > .05$), mercury coefficient reported for entire sample

If Male*MeHg interaction IS significant ($p \leq .05$), mercury coefficient reported by sex

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

Table 4.

Recent MeHg (ppm) exposure and n-3 and n-6 PUFA status association with heart rate variability measurements, adjusting for birth weight, BMI, and activity level

Outcome	N	P value for sex interaction	β	P value	95% CI
NN (ms)					
Long-term Recording	167	0.3404	-2.755	0.0270	(-5.191, -0.319)
Baseline Supine	418	0.0717	-3.048	0.0160	(-5.525, -0.571)
Response to Standing	411	0.0185	0.167	0.8420	(-1.485, 1.82)
female			2.617	0.0499	(0.0011, 5.2323)
male			-1.426	0.1847	(-3.5366, 0.6838)
Response to Math Test	365	0.1291	0.578	0.4420	(-0.9, 2.056)
Response to Sleep	167	0.4733	-0.095	0.9680	(-4.77, 4.581)
SDNN (ms)					
Long-term Recording ⁺	165	0.4254	-0.006	0.0950	(-0.012, 0.001)
Baseline Supine ⁺	418	0.6280	-0.005	0.1870	(-0.013, 0.003)
Response to Standing *	411	0.5215	0	0.9400	(-0.007, 0.007)
Response to Math Test *	365	0.7105	0.004	0.1480	(-0.002, 0.01)
Response to Sleep *	167	0.4633	-0.010	0.2050	(-0.026, 0.006)
rMSSD (ms)					
Long-term Recording ⁺	167	0.1793	-0.010	0.1300	(-0.022, 0.003)
Baseline Supine ⁺	411	0.5038	-0.008	0.1280	(-0.018, 0.002)
Response to Standing *	405	0.1855	0.004	0.3360	(-0.004, 0.012)
Response to Math Test *	358	0.4383	0.010	0.0060	(0.003, 0.016)
Response to Sleep *	165	0.4679	-0.007	0.4200	(-0.024, 0.01)
LF					
Baseline Supine	416	0.3200	-0.128	0.3860	(-0.417, 0.161)
Response to Standing	410	0.2926	-0.361	0.1060	(-0.798, 0.077)
Response to Math Test	364	0.0425	-0.245	0.1910	(-0.612, 0.122)
female			0.209	0.4720	(-0.3615, 0.7791)
male			-0.563	0.0210	(-1.0401, -0.0855)
Response to Sleep	166	0.2318	-0.688	0.0500	(-1.377, 0.001)
HF					
Baseline Supine	416	0.6151	0.226	0.1890	(-0.112, 0.564)
Response to Standing	410	0.8996	0.490	0.0110	(0.114, 0.866)
Response to Math Test	364	0.7939	0.544	0.0030	(0.188, 0.901)
Response to Sleep	166	0.8533	0.814	0.0550	(-0.018, 1.646)

⁺ = Model uses the natural logarithm of the outcome and slopes assume multiplicative effects

= Difference between baseline values and values obtained at standing, math test, or sleeping respectively

* = Difference in natural logarithms of baseline values and values obtained at standing, math test, or sleeping respectively

If Male*MeHg interaction is not significant ($p > .05$), mercury coefficient reported for entire sample

If Male*MeHg interaction is significant ($p \leq .05$), the mercury coefficient is reported by sex

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript