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Targeted Therapy in Metastatic Colorectal Cancer: Current Standards and Novel Agents in Review

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Abstract

Purpose of Review—Treatment options for patients with metastatic colorectal cancer continue to advance as the therapeutic implications of the molecular subtypes of this disease are becoming better understood. DNA sequencing and mismatch repair assessment are now standard of care analyses for patients with metastatic colorectal cancer. This review describes important aspects of the biology of the clinically relevant molecular subtypes of colorectal cancer based on the current standard of care testing. In addition, the clinical treatment strategies available now and potentially in the future for these colorectal cancer subtypes are discussed.

Recent Findings—Currently for metastatic colorectal cancer, standard of care molecular testing is done for mutations in exons 2, 3, and 4 of *KRAS* and *NRAS*, and *BRAF* V600E. Testing for mismatch repair (MMR) deficiency/microsatellite instability (MSI) status is also done. These aberrations are well known to change the clinical prognosis and guide patients' treatment strategies. Additionally, three new subtypes have emerged: *PIK3CA* mut, *HER2* amplified, and *NTRK* fusions. With the addition of these emerging subtypes, tumor heterogeneity further validates the need to examine mCRC as a heterogeneous disease. Here we present recent exciting data from translational research and clinical trials exhibiting possible distinct treatment strategies for these different subtypes.

Summary—Altogether these data show promising treatment strategies for many of these well-known and emerging subtypes of mCRC. In addition, these also give better clinical prognostic and predictive information. We believe that as molecular testing expands *PIK3CA* mutation, *HER2* amplification, and *NTRK* fusion molecular testing will be included in standard of care analyses.

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Conflict of Interest

Rebecca A. DeStefanis declares that she has no conflict of interest.

Jeremy D. Kratz declares that he has no conflict of interest.

Philip B. Emmerich declares that he has no conflict of interest.

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Human and Animal Rights and Informed Consent

This article does not contain any studies with human or animal subjects performed by any of the authors.

This incorporation of testing in clinical practice will generate further information regarding prognostic and therapeutic options for these and other CRC subtypes in the future.

Keywords

colon cancer; rectal cancer; targeted therapy; molecular subtypes; emerging subtypes chemotherapy

INTRODUCTION

Colorectal cancer (CRC) is the third-leading cause of cancer-related deaths in the United States with ~50,000 deaths each year [1]. Standard cytotoxic chemotherapy remains the mainstay of treatment for metastatic CRC, however a growing appreciation for the molecular subtypes of CRC has led to recent advancements in the manner in which this disease is treated [2]. FOLFOX (leucovorin, 5-FU, and oxaliplatin) or FOLFIRI (leucovorin, 5-FU, and irinotecan) chemotherapy regimens are largely utilized in the first-line setting, commonly in conjunction with bevacizumab, an anti-vascular endothelial growth factor antibody, regardless of molecular subtype [3–4]. Bevacizumab is a humanized monoclonal antibody that targets the vascular endothelial growth factor. Initial investigations of bevacizumab showed evidence of clinical benefit when added to bolus irinotecan, 5-FU and leucovorin (IFL) including improvements in median overall survival (OS) and progression free survival (PFS)[5]. This was extrapolated to current use in combination with FOLFOX chemotherapy [6]. Pooled analysis of phase II and III trials has shown improvement for the addition of bevacizumab to chemotherapy with improvement in OS 18.7 versus (vs) 16.1 months ($p=0.0003$), PFS 8.8 vs 6.4 months ($p<0.0001$), and response rate (RR) 39% vs 33% [7].

The current standard of care molecular testing for metastatic colorectal cancer is to examine the mutational status of multiple genes. This includes exons 2, 3, and 4 of *KRAS* and *NRAS*. In addition, sequencing for the *BRAF*V600E mutation is needed. Beyond *RAS* and *RAF* testing it is also important to test for mismatch repair (MMR) deficiency/microsatellite instability (MSI). This is now most commonly done using immunohistochemistry for MLH1, MSH2, PMS2 and MSH6. Next-generation sequencing approaches are also commonly being utilized and the previously widely utilized PCR-based analyses are now done less frequently. *RAS/RAF* testing and MMR/MSI testing can be done as stand-alone tests, though these are now being more commonly performed as part of larger next-generation sequencing panels, allowing for the examination of other alterations of potential interest.

The standardized practice of analyzing CRCs for these alterations has identified subtypes of CRC with distinct treatment strategies (Table 1). These subtypes include *RAS/RAF* wild-type (wt) microsatellite stable (MSS), *KRAS/NRAS* mutant (mt) MSS, microsatellite instability-high (MSI-H)/MMR deficient, and *BRAF*V600E mt. Beyond standard of care testing there are three emerging populations of interest in metastatic CRC including *HER2*-amplified, *PIK3CA* mt, and NTRK fusions.

COLORECTAL CANCER MOLECULAR SUBTYPES

RAS/RAF Wild-type, Microsatellite Stable

The *RAS/RAF* wt MSS subtype represents about 30–40% of all CRC [8]. Cytotoxic chemotherapy remains the primary treatment backbone though this subtype is targetable with agents against the epidermal growth factor receptor (EGFR). EGFR is a transmembrane glycoprotein with extracellular ligand-binding domain and intracellular tyrosine kinase domain that mediates downstream signaling[9]. Upon ligand binding, EGFR functions as a homodimer with downstream activation of the RAS/RAF/MEK/ERK and PI3K/AKT/mTOR pathways resulting in cell growth, proliferation, and regulation of other critical cellular functions [10]. Pre-clinical models have demonstrated the therapeutic benefit of these agents, including the dependence of models with loss of APC requiring signaling through EGFR [11]. Two monoclonal antibodies targeting EGFR are approved for use in metastatic CRC including chimeric human-murine cetuximab and fully humanized panitumumab [12–13].

EGFR inhibitors, such as cetuximab and panitumumab, have undergone extensive clinical investigation and demonstrated benefit in the first-line in combination with chemotherapy and later-lines across multiple investigations [14–17]. The phase III multicenter prospective CALGB/SWOG 80405 clinical trial examined first-line chemotherapy in combination with either bevacizumab or cetuximab [14]. These regimens resulted in a median OS of 31.2 months in those patients with extended spectrum RAS/BRAF wt cancers. This study failed to show a significant difference between OS when comparing bevacizumab to cetuximab in this setting [18]. Despite selection for extended spectrum RAS, EGFR inhibitor resistance remains common at up to 60% of patients for which an improve understanding of mechanistic resistance and novel therapeutic strategies are needed [19].

An additional analysis of CALGB/SWOG80405 stratified clinical outcomes by primary disease sidedness. Right-sided disease was defined as proximal to the splenic flexure and left-sided distal to the spleen flexure [20]. In the extended-spectrum RAS/RAF wt population, right-sided primary tumors were found to have inferior clinical outcomes with the use of anti-EGFR therapy in the front-line setting. This was consistent with the population-based cohort from Canada showing inferior median OS for the use of EGFR inhibitors in the right-sided primary cohort at 30.5 vs 39.3 months [21]. Additionally, retrospective analysis of the FIRE-3 and CRYSTAL studies revealed improvement for left-sided tumors with treated with FOLFIRI in combination with EGFR inhibition and improved OS for right-sided tumors when starting in combination with bevacizumab[22]. The mechanistic basis for these differential clinical outcomes remains of great clinical interest at this time for which an improved mechanistic understanding is needed and may be accounted for due to differences in molecular profiling [23].

Despite the clinical utility of EGFR inhibition in the first-line setting, the molecular profiling might not be available at the time of treatment initiation. The first-line use of EGFR inhibitors also prolongs exposure and toxicities for patients including acneiform rash. The timing of incorporating EGFR inhibition into the treatment paradigm remains controversial among oncologists[18,24–28].

We recently reviewed the impact of the metastatic disease bulk in predicting the late-line utility of EGFRi[29]. Using a retrospective cohort, disease bulk was found to be an independent marker of therapeutic resistance to EGFR inhibition [29]. This study demonstrated consistent results with prior studies that also revealed delayed presentations for right-sided tumors with resultant increased rate of both bulky disease and multi-site metastases at presentation [30–32]. In addition to bulk harboring complex intratumor heterogeneity, alternative considerations for bulk resulting in therapeutic resistance include reduced antibody drug delivery [33], increased hydrostatic pressure [34], and the complexities of local blood supply [35]. This work requires validation in prospective trial design to tune the durability of EGFR inhibition in the non-bulky cohort.

KRAS/NRAS and *BRAF* mutations lead to primary resistance to EGFR inhibition and multiple other alterations are also implicated in this resistance, including amplifications in *KRAS* (<2%), *ERBB2* (5%), or *MET* (2%) [36–39]. The effect of alterations in the PI3K/AKT/PTEN pathway in predicting response to EGFR inhibition remains unclear due to limited prospective data and the high rate of concomitant mutations in both *KRAS* and *BRAF*[40]. Mutations in *EGFR* including S492R at the extracellular domain inhibit the binding of cetuximab as an acquired or secondary mechanism of resistance [41, 42]. Mechanisms of resistance are generally established, however the rarity of individual mutations provides a formidable challenge for prospective trial design to overcome these mechanisms. Limitations of traditional profiling with core biopsy may not capture the molecular profile of subclonal populations, which may be overcome by enhanced sensitivity in sequencing techniques [43, 44]. While intratumor heterogeneity leads to acquired resistance, further innovation is needed to improve the reliable characterization of these subclones.

Tracking the clonal dynamics over the course of therapy with circulating tumor DNA (ctDNA) may provide additional insights into the acquisition of different mechanisms of resistance for individual patients. Interestingly, the depth of the initial response and the time since progression from prior EGFR inhibition predicts future success with EGFR inhibition retreatment [45]. ctDNA data indicates selection for RAS mutations in response to EGFR inhibition through therapeutic pressure driving this mechanism of secondary resistance [31]. The mutant alleles responsible for secondary resistance undergo exponential decay following discontinuation of EGFR inhibition [47]. These techniques using ctDNA, help to explain the clonal basis for both secondary resistance and successful of retreatment strategies. These should be prospectively investigated as a powerful clinical tool to optimize EGFR inhibition retreatment strategies in the *RAS/RAF*wt MSS subtype.

KRAS/NRAS Mutant, Microsatellite Stable

KRAS and *NRAS* are members of the family of Ras oncogenes, small GTPases important in RAS/MAPK signaling. Activation of this pathway leads to cell growth, cell cycle progression, migration, and cell survival. Mutations in *KRAS* are found in approximately 40% of all CRC cases and are most often found at exons 2 (codon 12, 13), and less commonly at 2–5% in exons 3 (codon 61), and 4 (codon 146). Mutations in *NRAS* are found in only 2–5% of all CRC cases and are mutually exclusive with mutations in *KRAS*.

Mutations in either *KRAS* or *NRAS* result in constitutive activation of the RAS/MAPK pathway signaling and lead to resistance upstream EGFR targeting [48–53]. Some clinical investigations have even demonstrated worse outcomes when EGFR inhibitors are used in the setting of activating RAS alterations [19, 51, 54].

Targeted therapies aimed at the RAS oncoproteins or the downstream signaling cascades have been ineffective and strategies specifically benefitting this subtype of CRC remain elusive. Recently there was excitement in this area with early investigations of the combination of cobimetinib, a MEK inhibitor, and atezolizumab, an anti-PD-L1 agent, showing promise in treatment refractory CRC [55]. Excitingly, in the phase I trial increased activity was seen specifically in those cancers with enhanced RAS/RAF/MEK/ERK signaling [36]. The confirmatory phase III clinical trial, IMblaze370, examined the combination of cobimetinib and atezolizumab in chemotherapy-refractory metastatic CRC compared to atezolizumab or regorafenib, a multi-targeted receptor tyrosine kinase inhibitor, alone. Over half of the patients in this study were *KRAS* mt. Unfortunately, this study did not meet its primary endpoint of an improvement in OS and the RR to this regimen was only 3% [56]. Studies using other similar regimens are on-going with the benefit of this approach specifically for the *KRAS* mt MSS yet to be determined. Additional therapies of interest for targeting this subtype of CRC include agents that target CEA expression, often found in *KRAS* mt cancers. This includes agents such as a cibisatuzumab which is a CEA CD3 bispecific antibody that has demonstrated interesting activity in an early phase clinical trial [57].

Microsatellite Instability-High/Mismatch Repair Deficient

Microsatellites are short, tandemly-repeated sequences of DNA throughout the genome and are commonly shortened in the setting of deficient mismatch repair (dMMR) protein activity [58]. Metastatic MSI-high/dMMR patients represent 3–4% of all metastatic CRC [59]. The most commonly altered DNA MMR genes are MLH1, MSH2, MSH6, and PMS2 with greater than 90% of those dMMR cancers having alterations in MLH1 and MSH2. The genes can undergo mutation or epigenetic regulation resulting in loss of the functional protein. MMR deficiency can be detected either on a protein, mutation status, or analyses of microsatellite status [60].

MMR deficient cancers have been an area of active interest for the development of immunotherapeutic strategies. Secondary to the dMMR status these tumors develop possessing 100s to 1000s of mutations. These mutations result in an enhanced neoantigen load leading to the potential for enhanced immune recognition [61]. These cancers also tend to have lower rates of alterations in WNT signaling compared to MSS CRCs, which has been implicated in immunotherapy resistance [62, 63]. Recent investigations have examined the use of anti-PD1 therapies for patients with MSI-H/dMMR metastatic CRCs in the treatment refractory setting [64]. Pembrolizumab demonstrated a RR of 40% and the progression free survival (PFS) at 20 weeks was 78% [65]. Similarly, nivolumab demonstrated a RR of 31% and 69% had a PFS of 12 weeks or greater [66]. Of note, durable responses are seen with these patients, especially in those who develop a clinical response.

This has led to the FDA approval of both pembrolizumab and nivolumab for patients with treatment refractory MSI-H/dMMR metastatic CRCs.

Dual immune checkpoint has also recently been reported examining the combination of nivolumab with the CTLA4 antibody ipilimumab as part of the CheckMate 142 clinical trial. This combination resulted in a RR of 31% and a 12-month PFS of 71% [67]. Additionally, this regimen was also recently investigated in the first-line metastatic setting in CRC and demonstrated a RR of 60% and a 12-month PFS of 77% [68, 69]. Given this success, other immuno-oncology treatment strategies targeting anti-PD1 therapies in combination with agents against other immune targets, such as LAG3 and TIM3, are being actively pursued.

BRAF Mutant

BRAF is a serine/threonine kinase found downstream of RAS in the RAS/RAF/MEK/ERK pathway. Mutations in *BRAF* are found in about 10% of CRC patients with the majority of them being V600E (80% of all *BRAF* mutations). This mutation leads to constitutive activation of BRAF and the downstream signaling pathway. *BRAF*^{V600} mt CRCs have a worse prognosis, relative chemotherapy resistance, and upon progression these patients clinically deteriorate quickly resulting in many patients being unable to receive second-line therapy [2, 70]. In addition, secondary to the effects of mutant BRAF downstream from EGFR signaling, anti-EGFR therapies have not shown consistent benefit for patients. Given this lack of benefit, patients with *BRAF* mt CRC are not treated with anti-EGFR therapies as single-agents [71–74]. Since the *BRAF*^{V600} mutation activates the BRAF kinase, agents targeting BRAF have known clinical utility across multiple cancers including melanoma. However, *BRAF*^{V600} mt CRC is not nearly as sensitive to single agent BRAF inhibitors as compared to *BRAF* mt melanoma. Combination strategies have emerged to overcome resistance mechanisms, including signaling through EGFR. The recent SWOG1406 phase III clinical trial examined *BRAF* mt CRC patients treated with the combination of irinotecan (topoisomerase I inhibitor), cetuximab (anti-EGFR), and vemurafenib (BRAF inhibitor) compared to irinotecan and cetuximab. This study demonstrated an increase in median PFS (4.4 vs 2 months, $p < 0.001$) and increased RR (16% vs 4%, $p = 0.09$) for the triplet combination compared to the standard of care arm, respectively [75]. Though the additional benefit of this regimen is modest, for this particular subtype of CRC these results are important and have led to this becoming a standard treatment option for these patients.

Additionally, the combination of encorafenib (Raf kinase inhibitor), cetuximab (anti-EGFR) and binimetinib (MEK inhibitor) was examined in *BRAF*^{V600E} mt CRC patients in a phase I/II and in an on-going phase III clinical trial. In the phase II, 30 patients were enrolled. A RR of 40% and a median PFS of over 8 months were observed. Given the results of the phase II study the outcomes of the phase III clinical trial are expected to be practice changing and a first-line study is expected to start enrolling soon [76].

It is important to note that *BRAF*^{non-V600} mutations largely possess a different biology than *BRAF*^{V600} mutations. *BRAF*^{non-V600} mutations occur in 2% of all CRC cases and many of the common *BRAF*^{non-V600} mutations actually inactivate the BRAF kinase [54]. *BRAF*^{non-V600} mt CRCs have a better prognosis than *BRAF*^{V600} mt. *BRAF*^{non-V600} mt CRC patients have a longer OS compared to those *BRAF*^{V600} (60.7 vs. 11.4 months,

respectively). Additionally, *BRAF^{non-V600}* CRCs are more likely to co-occur with *RAS* mutations, but interestingly have a similar clinical prognosis to *RAS* WT patients [77]. These results demonstrate the importance of identifying the particular mutation and its change in the protein function when determining treatment strategies.

EMERGING COLORECTAL CANCER SUBTYPES

PIK3CA Mutant

PIK3CA is the gene that encodes for the p110 α catalytic subunit of PI3K, a phosphoinositide kinase important in the PI3K/mTOR signaling pathway. Activation of this pathway leads to enhanced protein synthesis, cell cycle progression, cell growth, and survival. Mutations in *PIK3CA* are found in approximately 18% of CRC cases with 48% of those mutations occurring in the kinase domain (most commonly H1047R) and 43% occurring in the helical domain (most commonly E542K and E545K) [78]. While genetic profiling of this gene is not routinely recommended for metastatic CRC patients, many medical centers nationwide have begun including *PIK3CA* sequencing to guide clinicians in choosing appropriate targeted therapies as part of genetic sequencing panels.

Currently, there are several inhibitors of the PI3K/mTOR pathway which directly target PI3K alone, mTOR alone, and or the combination (dual PI3K/mTOR inhibitors). To date, no clinical trial has looked specifically at these inhibitors in the context of patients with *PIK3CA* mutant CRC. Our group has demonstrated the preclinical benefit of targeting MTORC1/2 for *PIK3CA* mutant CRCs. Using a novel transgenic mouse model possessing the hotspot *Pik3ca^{H1047R}* mt and loss of APC, mTORC1/2 inhibition resulted in treatment response. Using tumor derived spheroid cultures we demonstrated that these cancers were resistant to MTORC1 inhibition with everolimus and the PI3K alpha isomer specific inhibitor BYL-719. These treatment strategies need to be prospectively examined in *PIK3CA* mt CRCS without concomitant mutations in RAS or RAF. [79]

HER2-Amplified

Human epidermal growth factor receptor 2 (HER2) is a member of the epidermal growth factor receptor family in which activation leads to upregulation of the RAS/MAPK and PI3K/mTOR pathways. HER2 amplification is found in about 3–5% of all CRC cases and can lead to relative resistance to anti-EGFR therapies [80]. It is not currently standard of care to examine for HER2 amplification at most centers, though this subtype of CRC is targetable using agents that target HER2. This is especially true for those cancers with a copy number greater than 10 [81].

One of the initial studies to target this subset of CRC was the HERACLES phase II trial which examined combination of trastuzumab, a HER2 directed antibody, and lapatinib, an oral dual HER2/EGFR kinase inhibitor. Of those enrolled in the study, 30% of patients (8/27 patients) had an objective response [82]. The MyPathway trial enrolled patients with HER2-amplified CRC to receive the combination of pertuzumab (anti-HER2) and trastuzumab. In the initial report of this study, 38% of the cohort have received an objective response rate [83]. Additionally, there is an ongoing phase II clinical trial SWOG1613 examining the

combination of trastuzumab and pertuzumab vs cetuximab and irinotecan in metastatic HER2-amplified CRC. Given the promising results of those studies, it is anticipated that it will become a national standard to test all patients with metastatic CRC for HER2 amplification.

NTRK Fusions

The neurotrophic tropomyocin receptor kinase (NTRK) genes encode for the TrkA, TrkB, and TrkC receptor tyrosine kinases that are important in the function of the nervous system in human neuronal tissue. These proteins are activated by the nerve growth factors neurotrophins (NTs) which leads to activation of the RAS/MAPK pathway, PI3K pathway, or the phospholipase C- γ (PLC- γ) pathway resulting in neuronal differentiation and survival. However, gene fusions of the *NTRK* gene can occur in which the 3' region is joined with the 5' sequence of a fusion partner gene leading to constitutive activation or overexpression of the Trk kinase [84–87]. These fusion proteins have been reported to occur in ~1.5% of all CRCs. Although these fusion proteins are rare in CRC, the FDA has recently approved larotrectinib, a Trk inhibitor, for all solid tumors with *NTRK1*, *NTRK2*, and *NTRK3* fusion genes. This is the second time the FDA has approved a drug independent of cancer type but the first time a targeted therapy has been approved in this setting. Three clinical trials were used to assess the efficacy of this drug: a phase I trial in adults, a phase 1–2 study in children, and a phase 2 study in adolescents and adults. The combined analysis endpoint had an overall response rate of 75%, including achieving complete response[88]. Given these results and FDA approval, it is anticipated that examining NTRK fusions will become a part of the standard molecular screening of CRCs.

Summary and Conclusions

Colorectal cancer is a diverse disease with multiple histological and molecular subtypes. Standard of care molecular testing for metastatic CRC includes the assessment of mismatch repair deficiency and analysis for mutations in *KRAS*, *NRAS*, and *BRAF*. This is now often being performed clinically using IHC to determine the MMR status and a sequencing panel for the mutation analyses. This information directly informs clinical practice with both prognostic and predictive information. Each of the identified molecular subtypes of CRC outlined above have distinct treatment strategies that continue to evolve. The sequencing analyses being performed now should be done as part of large clinical sequencing platforms. Large panel testing allows for the standard of care testing to be done, and additionally, those alterations that we believe will soon be part of standard testing including *PIK3CA* mutations, *HER2* amplification and *NTRK* fusions among others. As further testing is being incorporated into clinical practice further information will be generated regarding the prognostic and therapeutic options for these and other CRC subtypes in the future.

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Table 1.**Molecular Subtypes of Colorectal Cancer and Select Treatment Options**

CRC Subtypes	Select Treatment Options
<i>RAS/RAF</i> wild-type, microsatellite stable	cetuximab or panitumumab
<i>KRAS/NRAS</i> mutant, microsatellite stable	cytotoxic chemotherapy, no current targeted therapy options
MSI-H/MMR deficient	pembrolizumab or nivolumab or nivolumab in combination with ipilimumab
<i>BRAF</i> V600 mutant	vemurafenib, cetuximab and irinotecan or MEK plus BRAF inhibitor in combination with cetuximab or panitumumab
<i>HER2</i> amplified	trastuzumab in combination with pertuzumab or lapatinib
<i>NTRK</i> fusion	Larotrectinib
<i>PIK3CA</i> mutant, <i>RAS/RAF</i> wild-type	PI3K inhibitor clinical trials