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The microgenderome revealed: sex differences in bidirectional interactions between the microbiota, hormones, immunity and disease susceptibility

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Abstract

Sex differences in immunity are well described in the literature and thought to be mainly driven by sex hormones and sex-linked immune response genes. The gastrointestinal tract (GIT) is one of the largest immune organs in the body and contains multiple immune cells in the GIT-associated lymphoid tissue, Peyer's patches and elsewhere, which together have profound effects on local and systemic inflammation. The GIT is colonised with microbial communities composed of bacteria, fungi and viruses, collectively known as the GIT microbiota. The GIT microbiota drives multiple interactions locally with immune cells that regulate the homeostatic environment and systemically in diverse tissues. It is becoming evident that the microbiota differs between the sexes, both in animal models and in humans, and these sex differences often lead to sex-dependent changes in local GIT inflammation, systemic immunity and susceptibility to a range of inflammatory diseases. The sexually dimorphic microbiome has been termed the 'microgenderome'. Herein, we review the evidence for the microgenderome and contemplate the role it plays in driving sex differences in immunity and disease susceptibility. We further consider the impact that biological sex might play in the response to treatments aimed at manipulating the GIT microbiota, such as prebiotics, live biotherapeutics, (probiotics, synbiotics and bacteriotherapies) and faecal microbial transplant. These alternative therapies hold potential in the treatment of both psychological (e.g.,

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Compliance with ethical standards

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anxiety, depression) and physiological (e.g., irritable bowel disease) disorders differentially affecting males and females.

Keywords

Adaptive immunity; Innate immunity; Sex differences; Sex hormones; Probiotics; Faecal microbiota transplant; Bacteriotherapy

The healthy microbiota and its role in immune homeostasis

The intestinal mucosal interface is a site of intense immune homeostasis whereby antigens from food are surveyed and processed, and the balance between adverse inflammation and pathogen surveillance is maintained. The microbiota consists of commensal, symbiotic and pathogenic bacteria, as well as fungi and viruses, and the microbial genome of these microorganisms is termed the microbiome. The body contains at least an equal number of microbes as there are cells in the human body [1], with the majority of these microbes occurring in the gastrointestinal tract (GIT). The GIT is the largest immune organ in the body, and multiple studies have shown that these resident microbial communities, termed the GIT microbiota, can educate immune development and modulate host inflammatory status. This occurs via microbiota effects on both innate and adaptive immunity (Table 1; Fig. 1) [2–12, 15–29]. The microbiota differs depending on the region of the GIT (oesophagus, stomach, upper and lower intestine, colon), but this review will focus on the colonic microbiota where an estimated 70% of the GIT microbiota reside [4].

Healthy microbiota versus dysbiosis

Colonisation of the GIT commences at birth and early colonisation events (e.g., vaginal birth, antibiotics) are thought to have profound influences on GIT development, immune system function and metabolic homeostasis, thereby influencing long-term health and disease susceptibility [30]. For the purposes of this review, dysbiosis is an imbalanced or maladapted microbiota that can occur at any time in life and is often associated with localised or systemic inflammation (Fig. 1). A person's healthy microbiota rarely leads to excessive local or systemic inflammation, and often, certain microbial communities are considered healthier than others, though this is not the same for each individual [31]. The mammalian GIT microbiota consists of three major phyla, namely the Bacteroidetes, Firmicutes and Actinobacteria [4]. The diversity of Bacteroidetes is limited, thus individual species are typically the dominant species by abundance. Bacteroidetes are thought to play an important role in degradation of complex sugars and proteins [32]. The Firmicutes, in contrast, contain the most diversity but typically occur at lower levels than Bacteroides [33]. The Actinobacteria are early colonisers associated with breast milk that persist and are detectable in most healthy people [34]. The most prevalent members of the Actinobacteria phyla are from the *Bifidobacterium* genus and include *Prevotella* and *Faecalibacteria*, which are typically associated with health in humans. On the other hand, *Lactobacillaceae* (Firmicutes) are absent or occur at low levels in the healthy human GIT but are highly prevalent in the murine GIT. Indeed, while Bacteroidetes and Firmicutes dominate in the microbiota of many laboratory rodents (e.g., mice, rats, hamsters) [35–37], marked

differences between the rodent and human GIT microbiota at the genus level indicate that rodent studies may not be generalizable to humans [38]. Furthermore, individual rodents may vary greatly in their microbiota even when co-housed; thus, rigorous experimental techniques are required to ensure consistency in experiments [39].

Importantly, certain bacteria, such as *Bifidobacteriaceae* (Actinobacteria), are thought to have anti-inflammatory effects [40]. By contrast, others including many members of the *Lactobacillaceae*, such as *Lactobacillus acidophilus* [41], and Proteobacteria including *Enterobacteriaceae* [12] are highly pro-inflammatory. However, the picture is highly complex and it remains unclear whether the Proteobacteria drive inflammation or merely survive better in a pro-inflammatory environment than Firmicutes and Bacteroidetes [42]. Further, this greatly oversimplified view does not consider that the community of microbes (including viruses and fungi) interact in a cohesive way to maintain host homeostasis, a description of which is beyond the scope of this review.

Sex differences in the GIT microbiota and immunity—rodent studies

Sex differences in GIT microbiota

Many studies have shown that the GIT microbiota composition differs in adult male and female rodents [39, 43–46]. For example, while the pro-inflammatory *Lactobacillaceae* are more abundant in females, the highly pro-inflammatory *Ruminococcaceae* and *Rikenellaceae* are more prevalent in males [43]. Sex differences in composition, however, also depend on the species and strain being studied. For example, B6 females have greater *Lactobacillaceae* and *Bacteroides* than B6 males, whereas BALB/c females have greater *Bifidobacteriaceae* than BALB/c males [44]. These sex differences in the microbiota have been shown to correlate with sex differences in GIT expression of multiple genes controlling immunological function, including genes affecting inflammation and leukocyte migration [44]. For instance, in a study examining differential gene expression in the GIT mucosa, males had greater transforming growth factor beta (TGF- β), interleukin 1 beta (IL-1 β) signalling, and type 1 interferon (IFN) pathway regulation than females [43], further suggesting the important role sex plays in the microbiota-immunity interface.

Sex differences in the immunological effects of microbiota transfer

While the GIT microbiota shapes both innate and adaptive immune responses (Table 1), the reciprocal is also true in that the innate and adaptive immune systems can alter the composition of the local microbiota [47]. Thus, sex differences in systemic immunity are likely to contribute to sex differences in GIT microbiota and vice versa. A recent study addressed this reciprocal relationship by performing GIT microbiota transfer experiments from conventional [i.e., specific pathogen free (SPF)] to germ-free (GF) mice of the same or opposite sex, and examining effects on weight loss, organ-specific T and B cell immunity and gene expression in the GIT [43]. Female mice receiving female transplants maintained their normal body mass, whereas females receiving male microbiota or male recipients of either male or female microbiota lost weight, suggesting the female microbiota may be less pro-inflammatory, further confirmed by the analysis of local genes and pathways affected in these experiments [43]. The microbiota first adapts to the sex of the recipient, but by 4

weeks, some donor-specific sex differences become apparent [43]. Mice receiving female microbiota had higher levels of double-negative T cell precursors compared to mice receiving male microbiota, suggesting a sex-dependent effect of the GIT microbiota on T cell development [43]. However, there were no sex differences in T helper (Th) types Th1, Th2 or Th17 cell subsets in PP, mesenteric lymph nodes (MLNs) or spleens [43]. In another study, transfer of male microbiota to GF males led to greater ROR γ t⁺ Foxp3⁺ T cells (i.e. those that inhibit Th2 mediated pathology) in PP and MLN compared to male recipients of female microbiota [48], which may explain the greater propensity of females to food allergies.

The above transfer study also investigated the effect of the microbiota on antibody levels [43]. Notably, GF females had higher baseline antibodies (i.e. titers of IgM and IgE) compared to GF males, but similar levels to conventional females, suggesting that the microbiota may not influence specific aspects of immunity in females [43]. In contrast, GF males have lower baseline antibody titers (i.e. IgG2a and IgG2b) than conventional males, and the transfer of female microbiota significantly lowers IgA levels compared to male microbiota transfer, all supporting a microbiota-induced effect on antibodies in males [43]. However, these data do not confirm whether the microbiota influence how the immune system may respond to a true infection.

Sex differences prior to puberty

Most studies examining the role of sex in GIT microbiota have generally investigated adult animals. However, investigations of the GIT microbiota in young animals suggest that these microbial sex differences do not appear until after puberty [37]. For example, deep sequencing of the colonic luminal contents from pre-pubescent C57BL/6 mice show no effect of sex on colonic bacterial community composition [37]. In this study, however, whole-genome profiling of colonic and small intestinal tissue from pre-pubertal mice revealed multiple genes that exhibit sexually dimorphic expression, many of which were autosomal and involved in infection, immunity and inflammatory pathways, particularly in the small intestine [37]. Thus, even in the absence of high levels of circulating sex hormones, mice with identical microbiota have intrinsic sex-specific gene regulation in the GIT.

In summary, there are multiple descriptions of sex differences in the adult rodent microbiota, which in turn influence local and systemic immunity in a sex-specific manner. These include effects on innate and adaptive immunity generally that drive an anti-inflammatory GIT in females and a more inflammatory environment in males. How profound this effect is in contributing to sex differences in systemic immunity is not known at present but is likely to be considerable given the major contribution of the GIT microenvironment to immune homeostasis (Table 1).

Effect of the microbiota on sex-specific behaviour in rodents

Changes in the GIT microbiota can influence behaviour in sex-dependent ways, and many studies have supported the microbiota's role in behavioural abnormalities (e.g., anxiety-like behaviour). For example, in Siberian hamsters, treatment with a broad-spectrum antibiotic

not only affects faecal microbial communities in both sexes, but there is a strong sex difference in the behavioural response to treatment. Specifically, two, but not one, antibiotic treatment is associated with marked decreases in aggressive behaviour in males, but in females, there is a decrease in aggression after only a single treatment [36]. Further, Neufeld and colleagues showed that GF Swiss Webster mice exhibit decreased anxiety-like behaviour in the Elevated Plus Maze (EPM) (e.g., increased time spent in the open arm), which was accompanied by increased brain-derived neurotrophic factor (BDNF) expression and decreased serotonin receptor 1A (5HT1A) expression in the hippocampus compared to SPF mice [49]. In a separate study, male and female Swiss Webster mice spent less time near a conspecific, as well as less time investigating a novel conspecific, when compared with a familiar one, suggesting increased anxiety-like behaviour [50]. These contradictory data suggest that normal GIT microbial communities may modulate important neurotransmitters that can greatly influence behavioural responses in sex-dependent ways, yet the precise role of the microbiota in modulating these responses is not completely understood.

Sex hormones drive microbiota differences in rodent models

The lack of sex differences in pre-pubertal mice supports a role for sex hormones in driving sex differences in the microbiota. Sex hormones are known to have many immunological effects with estrogens generally being considered pro-inflammatory and androgens anti-inflammatory [51]. The GIT microbiota modulate the enterohepatic circulation of non-ovarian estrogens in men and post-menopausal women thereby affecting local and systemic sex hormone levels and likely act in the same manner across animal models [52].

Oestrogen receptors are expressed by murine intestinal macrophages and nerve cells as well, the former of which can be activated by exogenous oestrogen administration [53, 54]. Oestrogen therefore acts at multiple levels of the GIT and plays an important role in maintaining health or causing disease. Estrogens are implicated in driving experimental colitis in mouse models [55], although certain murine colitis models are not affected by estrogens [56].

Rodent studies further suggest that GIT bacteria regulate local production of testosterone leading to protection against the development of type 1 diabetes in male mice via altered IFN- γ and IL-1 β -mediated signalling [57, 58]. Oral gavage of type 1 diabetes prone non-obese diabetic (NOD) females with male caecal contents increases testosterone levels in females and reduces insulinitis, insulin antibodies and disease incidence, an effect reversed by the anti-androgen drug flutamide [57]. A metabolomics analysis in this study showed that the sex of the microbiota differentially affected metabolic outcomes such as serum long-chain fatty acid levels. Again, these effects were abrogated by androgen receptor blockade suggesting that increased testosterone following male microbiota transfer was a key factor in the metabolomic effects. This provides direct evidence for a role for the GIT microbiota in influencing male sex hormone levels and altering autoimmune disease susceptibility. Androgens may similarly influence GIT microbial composition in lupus-susceptible mice and protect males against the development of lupus [47].

Sex differences in the GIT microbiota—human studies

Human studies have similarly shown that sex influences the GIT microbial composition, with the female microbiome having lower abundance of the Bacteroidetes phylum than males [59, 60]. A recent study showed sex differences in the GIT microbiome in a Japanese population; however, this was confounded by stool consistency, making it difficult to determine if this represents a biologically relevant difference or a sampling bias [61]. In conflict with these latter studies, a study investigating geographical variation in the microbiome among 1020 healthy individuals from 23 different populations across four continents found a similar abundance of Firmicutes and Bacteroidetes in males and females [62]. This study compiled data from six studies, and thus methodology and population differences may have masked a sex effect.

Healthy females reportedly have increased relative abundance of species in the *Bacteroides* genus than males in the GIT [63, 64]. In contrast, higher relative abundance of *Escherichia* [64] and *Veillonella* genera and lower relative abundance of the *Bilophila* genus have all been described in males [63]. However, another study showed increased *Bacteroides* in males compared to females [59]. *B. fragilis*, a member of the *Bacteroides* genus that is rarely prevalent in healthy individuals, activates T cell-dependent immune homeostatic mechanisms via the production of polysaccharide A, leading to Treg activation and suppression of Th17 responses and could therefore contribute to sex differences in immunity [13].

Generally, females have more robust innate and adaptive immune responses than males [51], but the role that the GIT microbiota plays in this effect is not known. Healthy adult females have greater immune activation and inflammation-associated gene expression in small intestine mucosal biopsy samples compared with males, in conjunction with higher peripheral blood T cell activation and proliferation and upregulated CD4 T cell IL-1 β and Th17 pathway genes [14]. This suggests that gut inflammation drives systemic inflammation in a sex-specific manner even in healthy asymptomatic individuals and likely contributes to the superior immune systems of females.

Further, oestrogen levels in men and post-menopausal women directly correlate with GIT microbiome richness and diversity, whereas there is no correlation in pre-menopausal women who are at varying stages of the menstrual cycle [52]. In particular, Clostridia taxa and several *Ruminococcaceae* family members are affected, suggesting that GIT microbes influence levels of non-ovarian oestrogens via the enterohepatic circulation [52]. The human microbiota changes throughout the various stages of pregnancy further implicating a modulatory role for sex steroids [65]. Diversity appears to decrease throughout pregnancy while certain bacteria such as *Proteobacteria* and *Actinobacteria* increase in the majority of women, the former known to be associated with inflammatory mediated dysbiosis [66]. A study of diverse human populations (Venezuela, Malawi, USA) across the lifespan showed distinct changes in the microbiota at puberty further implicating a role for sex hormones [67]. In an investigation of American twins, no significant sex differences in microbiota relatedness among infant twin pairs of the same sex compared to the opposite sex were found. However, there was greater faecal microbiota dissimilarity in opposite sex teenage

(13–17 years) twin pairs compared to same sex pairs, supporting microbiota sex differences in older individuals [67].

Additionally, microbial fermentation in the GIT leads to the production of short-chain fatty acids (SCFAs) such as butyrate, propionate and acetate. These have a myriad of anti-inflammatory effects including induction of Tregs (Table 1), inhibition of nuclear factor- κ B (NF- κ B) activation and suppression of tumour necrosis factor (TNF)- α and IL-6 release [12]. Males have been shown to have higher serum SCFA levels compared to females [68], which may account for higher Tregs in males [51] and contribute to the lower systemic inflammatory status among males as compared to females [21–24].

The human microbiota-gut-brain axis and sex differences in behaviour

Changes in the GIT microbiota may also contribute to sex-specific behavioural abnormalities across clinical populations. For example, subsets of autism spectrum disorder patients (mostly male) show both changes in GIT microbial composition, as well as gastrointestinal symptoms [69]. Another study found that while the microbiota compositions were similar between the sexes, there were sex-specific interactions between the Firmicutes and myalgic encephalomyelitis (also known as chronic fatigue syndrome) symptoms, suggesting sex-specific functional effects of GIT microbiota [70]. This bidirectional crosstalk between the GIT and brain via neural, hormonal and immunological pathways, called the ‘microbiota-gut-brain axis’ (Fig. 1), has received considerable attention in recent years and may be associated with sex differences in a number of psychological and neurological conditions [69]. This gut-brain axis also provides the link between certain sexually dimorphic GIT disorders, such as the greater female susceptibility to irritable bowel syndrome and the behavioural comorbidities that occur with this disorder [71].

Diet, GIT transit and adiposity have sex-differential effects on GIT microbiota

A study specifically investigating the sex*diet interaction in four vertebrate species including fish, mice and humans confirmed that diet has a sex-specific effect on the GIT microbiome in humans and two species of fish, but no such interactions were apparent in laboratory mice fed a standard controlled diet [72]. Interestingly, dietary effects were not observed in fish if the statistical models failed to account for sex, illustrating the importance of taking sex into account in studies of GIT microbiota. The impact of dietary fibre intake on the microbiome composition also appears sexually dimorphic [60], possibly via the effects of fibre on oestrogen levels [73]. Furthermore, adipose tissue is a source of sex hormones [74] and several studies suggest that adipose tissue contributes to sex differences in the GIT microbiota [32, 75]. Importantly, focusing on sex differences, particularly estradiol and testosterone, can provide information on the regulation and motor function (e.g. transit time) in the GIT and ultimately the normal function of the GIT microbiome. Indeed, faster GIT transit in females as compared to males is thought to contribute to sex differences in the GIT microbiota [76, 77].

Dysbiosis triggers diseases that manifest differently in the sexes

Increasing evidence suggests that GIT dysbiosis can lead to a preferential skewing towards effector T cell development and trigger inflammatory and autoimmune diseases [47]. In some cases, the same bacteria may provide protection against certain conditions and predisposition to others. For example, segmented filamentous bacteria may predispose individuals to Th17-mediated diseases and asthma in murine studies [78, 79] but can protect against type 1 diabetes in NOD mice [80]. Intestinal dysbiosis has also been linked to systemic lupus erythematosus [81, 82], and it is speculated that sex differences in the GIT microbiota play a role in the greater female susceptibility to autoimmune diseases [57, 58, 83].

The GIT microbiota can also affect sites distant to the GIT. For instance, *Clostridium leptum* can induce protection against allergic airways disease via the induction and/ or activation of Tregs [84]. Changes in the GIT microbiota composition, specifically altered anaerobes and facultative anaerobes, can be linked to the development of hepatocellular carcinoma in female but not male mice, including increased abundances of *Allobaculum*, Erysipelotrichaceae, Neisseriaceae, *Sutterella*, Burkholderiales and *Prevotella* species [85]. The microbiota has been linked to sex differences in susceptibility to cardiometabolic syndromes as well [86].

Treatments for dysbiosis should consider the sex of the recipient

Prebiotic, probiotic and symbiotic therapy

There is increasing interest in manipulating the host microbiota to treat diseases that result from dysbiosis [1], but few studies have taken recipient sex into account. Administration of a probiotic mixture of five *Lactobacillus* strains to lupusprone mice improved renal function and had anti-inflammatory effects by lowering IL-6, IgG2a and IgA and increasing the levels of IL-10 in female and castrated male mice, but not in gonadally intact males [82]. Another probiotic mixture has been used to prevent cortical bone loss in female mice [87], and a different probiotic prevented bone loss in oestrogen-deficient mice after ovariectomy via anti-TNF factors [88]. Interestingly, supplementation of aged obese mice with the probiotic *L. reuteri* helped restore testosterone levels and decreased IL-17 levels in males [89], suggesting the important role that age plays in the response to shifts in microbial communities. Understanding host metabolism will be vital to treatment success. Male and female rodents, for example, metabolise a diet supplemented with the prebiotic oligofructose (OF) differently, with females having higher levels of *Bacteroidetes* compared to males [90]. Faecal butyrate levels are also increased in males, and liver IgA, IL-6 and caecal IL-6 levels are higher in males, while anti-inflammatory IL-10 levels are higher in females [90]. Further, mice given an oral anti-ageing oil treatment show sex-based differences in GIT microbiota modulation [46]. Collectively, these studies point to sex differences in the immunological effects of pre- and probiotic treatments and may hold great potential in the treatment of microbiota-associated disease (Fig. 1).

In a study specifically designed to examine the effect of sex on host immunity following *Mycobacterium avium* subsp. *paratuberculosis* (MAP) challenge in mice either fed the

probiotic *Lactobacillus animalis* or control fed, circulating cytokines (e.g. IL-1 α / β , IL-6, IFN- γ) differed between the sexes [91]. This suggests that Th1, Th2, Th17 and Tregs may all be regulated by sex-linked factors in the mouse GIT. Furthermore, females show an increase in Firmicutes, including *Staphylococcus* and *Roseburia*, in all groups except controls [91] indicating that male and female microbiota respond differently to certain dietary organisms, and that the female microbiota may be more susceptible to dietary manipulation than males. In an extension of these studies, the same authors examined cytokine transcription levels and found that males have greater expression of Th2 and B cell factors, while females have decreased pro-inflammatory cytokine expression after MAP infection [91]. Further, male mice show increased *Tgf- β* , *Il-10* and *Foxp3* expression (indicative of Treg induction) and *Il-17* and *Il-23A* (typical of Th17 responses). Presumably, these sex differences in gene transcription modulate T cell function via cytokine-dependent mechanisms.

FMT and bacteriotherapies

Faecal microbial transplant (FMT) is becoming of increasing interest as a treatment for multiple conditions associated with GIT dysbiosis in humans (Fig. 1). In particular, FMT has been used for the treatment of *Clostridium difficile*-associated colitis [88], but studies indicate that it may also be used to treat inflammatory bowel disease, irritable bowel syndrome, obesity, diabetes, depression and anxiety [92–95]. As described above, all these conditions can be sex-dependent and associated with sex differences in the GIT microbiota. One study showed sex differences in mouse GIT microbiota colonisation after receiving FMTs from a male on a short-term vegetarian and inulin-supplemented diet, demonstrating that males and females utilise the same microbiota differently [96]. Murine FMT was also reported to protect against radiation-induced toxicity in a sex-specific manner thereby improving the prognosis of tumour patients after radiotherapy [97]. To our knowledge, no one has suggested that the sex of the donor should be considered when screening for suitability as an FMT donor, but the studies in this paper indicate that this could be an important consideration. Indeed, a recent human study found that female sex was associated with failure of FMT to cure *C. difficile* infection [98]. Advances in methods of cultivation, measurement and preparation of gastrointestinal bacteria are now progressing research from FMT to the development of rationally selected mixtures of bacteria with proven therapeutic efficacy, termed bacteriotherapies. This approach overcomes the issues of disease risk, patient acceptability and ease of delivery associated with FMT; however, consideration of sex differences may also prove essential in the design of these therapeutic options.

Concluding remarks

Flak and colleagues introduced the concept of the microgenderome in 2013 to define the interaction between the microbiota, sex hormones and immunity [99], but since that time, the concept has not been widely adopted. Indeed, because gender is a social construct, and sex is a biological construct, the term ‘microgenderome’ may not be accurate because most of the factors driving male-female differences in the microbiota are determined by biological sex rather than gender. Whatever term is used moving forward, there is no doubt that there are sex differences in GIT microbial composition and function, in part driven by sex

hormones, and these in turn contribute to sex differences in immunity and susceptibility to a multitude of infections and chronic diseases. Understanding the role of the GIT microbiota in the development of chronic inflammatory diseases may have major therapeutic implications in the future. This therefore leaves little doubt that studies of the microbiota should take sex into consideration, as should therapeutic manipulation of the microbiota with pre-, pro-, syn- and post-biotics, since it is likely the sexes will respond differently to these treatments.

It should be borne in mind that the majority of these studies used 16s rRNA profiling to determine the microbiota composition. This approach is unable to provide the species and strain resolution generally required for detailed understanding of bacterial diversity [100]. With the wider adoption of metagenomic sequencing, standardised data reporting and more sophisticated bioinformatics tools alongside novel culture techniques, we should get a far better picture of the nature of sex differences in microbial communities [33, 100–102].

The studies reviewed herein support sex differences in microbiota, as well as sex-specific responses to the same microbiota. These data show that the microbiota can influence the immune, endocrine and metabolic systems in hosts. Further, the microbiota can affect susceptibility to autoimmune, psychiatric and neuro-inflammatory conditions in a sex-specific manner, likely contributing to male/female disparities in susceptibility to these conditions. The relationship between the microbiota, hormones, metabolism and immunity appears multi-directional, with dysbiosis causing systemic imbalance in these factors and systemic imbalance leading to dysbiosis. Collectively, these studies provide evidence that future investigations into the composition and function of the microbiota must continue partitioning data for sex-differential interactions to understand these intricate relationships more fully.

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Abbreviations

DC	dendritic cell
MØ	macrophage
IECs	intestinal epithelial cells

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Fig. 1.

The microgenderome revealed. The gastrointestinal tract (GIT) is a site of intense immune homeostasis in which the microbiota plays a pivotal role. Many studies show that the GIT microbiota differs in males and females. Likely causes include differing sex hormone levels in males and females, in part driven by sex differences in systemic sex hormone concentrations, but also influenced by the microbiota themselves. Sex differences in the microbiota composition drive sex differences in both innate and adaptive immunity, and the sex-differential innate and adaptive immune systems in turn drive sex differences in the

microbiota composition. A normal, healthy microbiota allows immune homeostasis to be maintained, but loss of the healthy microbiota (dysbiosis) can drive inflammation and susceptibility to inflammatory and autoimmune diseases. Moreover, the GIT microbiota communicates with the brain and vice versa in what has been termed the microbiota-gut-brain axis, likely contributing to sex differences in susceptibility to a number of neuropsychiatric conditions. The response to therapeutic alteration of the microbiota using prebiotics, live biotherapeutics (probiotics, synbiotics and bacteriotherapies) or FMT is also likely to be different in males and females, although this has not been specifically studied to date. DC = dendritic cell, MØ = macrophage, IECs = intestinal epithelial cells

Table 1

Effect of the GIT microbiota on innate and adaptive immunity

Immune factor	How it is affected by the GIT microbiota	Refs
Innate immunity		
Dendritic cells (DCs), macrophages in gut associated lymphoid tissue (GALT)	Microbiota expression of pathogen-associated molecular patterns (PAMPs) stimulate pattern recognition receptors (PRRs) on DCs and macrophages including toll-like receptors (TLRs), nucleotide-binding oligomerization domain containing protein (NOD) receptors, retinoic acid-inducible gene-I-like (RIG-I-like) receptors, leading to downstream signalling events. Many GIT microbiota-induced innate responses are anti-inflammatory leading to homeostasis via the production of suppressive cytokines, such as IL-10 and TGF-β.	[1–9]
Type 3 innate lymphoid cells (ILCs)	RORγt ⁺ type 3 innate lymphoid cells (ILCs) are stimulated directly and indirectly by the GIT microbiota.	[4, 10]
Invariant natural killer T cells (iNKT cells)	Induced by GIT microbiota to produce pro- and anti-inflammatory cytokines, regulate neutrophil recruitment and function, important in GIT immune homeostasis	[11]
Intestinal epithelial cells (IECs)	Microbiota PAMP-induced stimulation of PRRs leading to production of antimicrobial peptides, such as defensins, C-type lectins, and cathelicidins.	[12]
Endocrine cells	Secrete serotonin and neuropeptides allowing crosstalk with the immune system	[12]
Adaptive immunity		
Th1, Th2, Th17 immunity	GIT microbiota directly induces T helper 1 (Th1), Th2 and Th17 cells.	[13–17]
Regulatory T cells (Tregs)	Microbiota cause the generation of GIT and thymic-derived Tregs thereby maintaining tolerance. Bacterial-derived short-chain fatty acids (SCFAs) induce Tregs.	[18–24]
B cells and secretory IgA	GIT microbiota directly stimulates B cell development and regulates the generation of secretory IgA by plasma cells.	[7, 25–28]
Regulatory B cells	GIT microbiota can induce CD5 ⁺ regulatory B cells in mice.	[29]