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Phosphothreonine (pThr)–Based Multifunctional Peptide Catalysis for Asymmetric Baeyer–Villiger Oxidations of Cyclobutanones

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Abstract

Biologically inspired phosphothreonine (pThr)-embedded peptides that function as chiral Brønsted acid catalysts for enantioselective Baeyer–Villiger oxidations (BV) of cyclobutanones with aqueous H₂O₂ are reported herein. Complementary to traditional BINOL-derived chiral phosphoric acids (CPAs), the functional diversity of the peptidic scaffold provides the opportunity for multiple points of contact with substrates via hydrogen bonding, and the ease of peptide synthesis facilitates rapid diversification of the catalyst structure, such that numerous unique peptide-based CPA catalysts have been prepared. Utilizing a hypothesis-driven design, we identified a pThr-based catalyst that contains an *N*-acylated diaminopropionic acid (Dap) residue, which achieves high enantioselectivity with catalyst loadings as low as 0.5 mol%. The power of peptide-based multi-site binding is further exemplified through reversal in the absolute stereochemical outcome upon repositioning of the substrate-directing group (*ortho*- to *meta*). Modifications to the β -3 residue (^L-Dap to ^L-Phe) lead to an observed enantiodivergence without inversion of any stereogenic center on the peptide catalyst, due to noncovalent interactions. Structure–selectivity and ¹H-¹H-ROESY studies revealed that the proposed hydrogen bonding interactions are essential for high levels of enantioinduction. The ability for the phosphopeptides to operate as multifunctional oxidation catalysts expands the scope of pThr catalysts and provides a framework for the future selective diversification of more complex substrates, including natural products.

Graphical Abstract

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Author Contributions

The manuscript was written through contributions of all authors. All authors have given approval to the final version of the manuscript.

The authors declare no competing financial interest.

Crystallographic data are deposited with the Cambridge Crystallographic Data Center under the accession numbers CCDC 1865894 (**4d**), 1867752 (**5c**), 1865895 (**5e**), 1865896 (**5f**), 1865897 (**5s**), 1865899 (**P4**), and 1865898 (**7**).

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website.

Additional figures, synthetic procedures, characterization data for all compounds, and crystallographic information (PDF).

X-ray crystallographic data for compounds **4d**, **5c**, **5e**, **5f**, **5s**, **P4**, **7** (CIF).

Keywords

asymmetric catalysis; Baeyer–Villiger; chiral phosphoric acid; peptides; phosphothreonine

INTRODUCTION:

Phosphate esters are ubiquitous in biochemistry.¹ Phosphothreonine (pThr), among other proteinogenic amino acids (*e.g.* pSer, pTyr), is the product of site-selective post-translational modifications of proteins, which both induces global conformational changes^{2,3} and binds signaling molecules, as part of cellular regulation (Figure 1a).⁴ Yet, no known catalytic site of an enzyme engages pThr as part of the bond-forming or bond-breaking mechanistic machinery. However, chiral phosphoric acids (CPAs) have emerged as powerful catalysts for an ever-expanding list of asymmetric transformations.⁵ So far, BINOL- or SPINOL-derived CPAs have dominated the expansive literature of CPA-catalyzed reactions (Figure 1b).⁶ As a result of their C₂-symmetry and rigidity, BINOL CPAs have a well-defined binding pocket that facilitate high levels of enantioinduction, predominately arising from the steric bulk on the 3,3'-positions.^{6e,7} While further modifications to the scaffold can attenuate the electronics and enhance enantioselectivities in select transformations, the ability to construct diverse CPAs with new opportunities for secondary interactions, especially hydrogen bonding, is limited.^{6,8}

As a primary focus of our group is to functionalize complex molecules, we sought to develop an analogous strategy with added opportunities for non-covalent interactions (NCIs).⁹ Chiral catalysts that operate on highly functionalized substrates often require less rigid catalysts that possess an increased capacity to interact with substrates through multivalent NCIs, though the explicit, rational design of a suitable catalyst is not always straightforward.¹⁰ Additionally, the choice of functionalized model substrates provides an opportunity to examine substrate–catalyst interactions with drug-like molecules.¹¹ Accordingly, we have initiated a program that explores pThr as a biologically inspired catalytic residue in peptide-based Brønsted acid catalysts for a broad array of synthetically interesting reactions. Our first proof-of-principle study established that pThr-based catalysts could indeed mediate enantioselective reactions, including archetypal transfer

hydrogenations of functionalized quinolines (Figure 1c)¹² and reductive aminations of functionalized ketones.¹³ More recently, we have also found that these catalysts facilitate atroposelective cyclodehydrations in complex molecular settings.¹⁴ Reported herein are applications of pThr-based catalysts to an entirely orthogonal mode of reactivity, the oxidative ring-expansion of cyclobutanones.

Discovered in 1899, the Baeyer–Villiger (BV) reaction¹⁵ remains an indispensable tool for accessing chiral esters and lactones from the corresponding ketone precursors through the use of enzymes,¹⁶ organocatalysts,¹⁷ and transition metal complexes.¹⁸ Inspired by the pioneering work of Ding and co-workers on CPA-catalyzed BV oxidation^{17b}, we endeavored to learn whether pThr-based CPAs could facilitate oxidation chemistry with comparable or complementary selectivities and achieve a benchmark for peptide catalysts for the diversification of more complex substrates (Figure 1d). The enantioselective BV reaction of symmetrical, cyclic ketones is particularly appealing for elucidating catalyst control given the mechanistic ramifications. In general, the migratory aptitude is determined by the substituent best able to stabilize the partial positive charge (e.g. 3° alkyl > 2° alkyl > phenyl > 1° alkyl > methyl).¹⁹ However, for symmetrical ketones, the migratory aptitude of the substituents is identical. Furthermore, upon addition of H₂O₂, two Criegee intermediates may be formed (*syn*-**6** and *anti*-**6**), both of which can go on to form either enantiomer of product.²⁰ Therefore the stereochemical outcome is dictated by the ~90° bond rotation of the O–O peroxy group to obtain the necessary σ_{C-C} to σ^*_{O-O} orbital overlaps for migration (Scheme 1). The consequence of possible *syn*- and *anti*- Criegee intermediates is that the catalysts must possess the ability to control delivery of H₂O₂, through the differentiation of the diastereotopic faces of the ketone and/or position the dihedral angle of the peroxy O–O bond with high fidelity. As we show below, the application of pThr-based CPAs in the BV reaction resulted in a remarkable level of catalyst-control through the explicit design of peptide sequences that exploit secondary NCIs, similar to those often associated with enzymes.²¹ Taken together, the results herein expand the specific capabilities of CPAs for both enantioselective BV and pThr-based catalysts to the previously unreported realm of oxidation catalysis. Yet, it could be that the most exciting ramification of this work is the generalization of CPA catalysis to include scaffolds beyond C₂-symmetry, including those that contain catalyst functionality that enables specific interactions with functionalized substrate chemotypes. Shown below are dramatic examples of stereodivergence that highlight this point.

Results and Discussion

Preliminary studies in our laboratory on pThr-based peptide catalysts for the enantioselective BV oxidation commenced with 3-phenylcyclobutanone (**4a**) and peptide catalyst **P2** (Scheme 2). Employing similarly established reaction conditions^{17b}, the Baeyer–Villiger oxidation of **4a** proceeded to give **5a** with high conversion, albeit with low enantioselectivity (55:45 er). With **4b**, bearing a prochiral quaternary carbon, γ -butyrolactone **5b** was obtained in 95% conversion with a slight increase in enantioselectivity (57:43 er). Given the modularity of peptide catalysts, we focused on sequential residue optimization first. We chose peptide sequences that are predicted to adopt well-defined secondary structures,

namely β -turns, and identified aspects of the peptide catalyst responsible for enantioselectivity (Table 1).²²

Given the significant influence of the *C*-terminal group on enantioselectivity in prior studies, the dimethyl amide analog **P3** was prepared first, which afforded **5b** in 87% conversion and 64:36 er (Table 1, entry 1) with 10 mol% catalyst loading.²³ While other reactions investigated from our group have demonstrated that the *i*+2 residue has a pronounced effect on the enantioselectivity, modifications to the *i*+2 or *i*+3 positions (**P4–P7**) provided little influence on the enantioselectivity in the BV oxidation of **4b** (entries 2–5) (see Table 1 for “*i*+*n*” nomenclature).²³ Investigation into the diastereomeric series of the tetrameric peptides revealed the *L*-stereochemical configuration of the *i*+3, and *D*-stereochemical configuration of Pro *i*+1, was crucial (entry 6–7). While a number of reaction parameters were evaluated, permutations from the aforementioned conditions did not offer any advantages (see SI). However, when H₂O₂ is substituted with *t*-BuOOH, **5b** is obtained in only 19% conversion and poor enantioselectivity (entry 8). This observation suggests that a similar mechanism proposed by Ding and co-workers is operative, which invokes the phosphoric acid as a bifunctional catalyst.²⁴ Furthermore, when the phosphate moiety is removed, as in **P10**, catalysis is lost, highlighting the catalytic role of the phosphorylated amino acid (entry 9).

Given the relatively subtle effects on enantioselectivity observed thus far, we sought to make alterations to the catalytically active residue, as well as modifications nearest to pThr, that might enhance enantioselectivity. In contrast to rigid C₂-symmetric CPAs, non-equivalent rotamers about the β -C–O_{pThr} bond, in addition to the stereogenic P(V) and rapid tautomerization of the acidic proton, provide numerous, chemically distinct catalyst conformers.²⁵ When the phosphate benzyl group is removed, thereby producing a non-stereogenic phosphorous atom, the enantioselectivity is diminished (55:45 er, entry 10). Inversion of the Thr β -stereochemistry to *allo*-pThr ablates catalyst activity and selectivity, resulting in 47% conversion and 51:49 er (entry 11). Substitution of the β -methyl group with a more bulky β -phenyl substituent (**P13**) affords the product in 97% conversion, although surprisingly the enantioselectivity remains unaffected (67:33, entry 12) given the close proximity of the β -substituent to the phosphate moiety. The importance of the β -substituent is further highlighted as replacement of pThr with pSer resulted in 78% conversion and 58:42 er (entry 13).²⁶ While elongation of the *C*-terminus did not attenuate enantioselectivity (entry 2), an additional *D*-Pro at the *N*-terminus resulted in increased selectivity (73:27 er), (entries 14–17). At low temperatures (–15, –25 °C) **5b** could be obtained in up to 78:22 er with a slight decrease in yield (entries 18–19). Notably, **P18** provided higher enantioselectivity than commercially available BINOL derived CPA, (*R*)-TRIP **1b** (64:36 er), and comparable selectivities (81:19 er) to those previously reported.^{17b}

In order to gain additional information about the catalyst structure and develop hypotheses about NCIs, we determined the solid-state structure of **P4** using X-ray crystallography (Figure 2a). Interestingly, there were three intramolecular hydrogen bonds, including the predicted 10-membered β -turn hydrogen-bond between N–H(*i*+3)⋯O(*i*) and N–H(*i*)⋯O(*i*+4).²² Recently, the structures of various dimethylaminoalanine (Dmaa)-embedded β -turn catalysts were extensively studied, and we wondered whether the principles learned would be applicable in ongoing catalyst design.²⁷ In order to make a direct comparison, we

obtained a crystal structure of the Dmaa analog (**7**) (Figure 2b).²⁸ Indeed, both catalysts showed nearly identical secondary structures in the solid-state, despite the rather large change to the catalytically active residue (Fmoc-pThr(Bn) vs. Boc-Dmaa) as depicted by the similar hydrogen bond lengths and structural overlays (see SI for details). Moreover, both peptides exhibit similar ϕ and ψ dihedral angles that are consistent with a type II' β -turn secondary structure, and only show a significant difference in the conformation of the *i*+3 Phe side chain orientation.²² One observation from the solid-state structure of **P4** was the relatively exposed nature of the phosphate moiety, which may position substrates in such a way as to prevent favorable NCIs needed to achieve a highly enantioselective BV reaction.

Given the numerous examples of peptide catalyst-substrate interactions involving hydrogen bonding networks and the diversity of functionalization in drug-like molecules, we hypothesized an additional hydrogen bond donor or acceptor on the substrate might engage in further secondary contacts with the catalyst and perturb enantioselectivity.^{23,29} Using substrate **4c**, bearing a carbamate *ortho*-substituent and 10 mol% of **P7**, product **5c** was obtained in 97% yield and 66:34 er (Scheme 3). Initially, this result suggested additional functionality on the substrate had no influence on the enantioselectivity of the reaction, and that perhaps the selectivities observed thus far are due largely in part to steric hindrance. Based on the relative positions of the *i*pThr and *i*+3 Leu side chains in the solid state of **P4**, we postulated that an *i*+3 side chain-directing group (*e.g.* amide functionality) would afford the opportunity for substrate **4c** to engage in attractive secondary interactions with the amino acid side chains synergistically.

While more than half of proteinogenic amino acids possess side-chain functionality that can engage in hydrogen bonding interactions, for our purposes, we envisioned an ideal amino acid residue to be neutral, easily diversifiable, and sufficiently short as to reduce the rotational degrees of freedom. We imagined that the readily available 2,3-diaminopropionic acid (Dap) residue, a biologically significant non-proteinogenic amino acid, would fulfill all these requirements.³⁰ Indeed, when Dap(Cbz) (**P19**) was incorporated in the peptidic scaffold, the reaction delivered product **5c** in 97% yield and 78:22 er, lending credence to our hypothesis (Table 2a). Encouraged by these results, we began to further investigate the role of the *i*+3 Dap residue on the enantioselectivity of the BV reaction.

Hydrogenation of the Cbz-protecting group allowed access to a variety of side chain analogs, following amino functionalization, and in comparing catalysts **P19–P31**, a number of differences were realized (Table 2a). Initially, acetamide **P20** and benzamide **P21** were prepared, which both resulted in increased enantioselectivity (82:18 er and 80:20 er, respectively), whereas the serine analog (**P22**) afforded **5c** in decreased selectivity (69:31 er). Replacement of the acetamide (**P20**) with electron-deficient amides (**P23–P24**) resulted in generally lower selectivity, while the electron-rich amide **P25** gave similar results (80:20 er). There was little difference observed in enantioselectivity among bulkier amides (**P26**, **P27**), signifying that the increased selectivity of the catalyst is likely not attributable to steric factors alone. Catalysts containing a phenyl urea performed equally well (**P28**), although carbamate **P29** and sulfonamide **P30** afforded the product with reduced enantioselectivity. The directionality of the amide group was shown to be crucial, as the Asn analog **P31** resulted in a substantial decrease in selectivity (67:33 er). Together, these results suggest that

while the N–H of the side chain might not be directly involved in the putative secondary interactions, an amido group on the side chain is necessary for achieving high selectivities.³¹ The enantioselectivity could then be further increased through *N*-terminal modification (Table 2b, **P32–P39**). It was found that an electron-rich *N*-terminal amide was advantageous (**P34**, 86:14 er), while bulkier amides afforded decreased selectivities. Although a steric effect is possible, the increased enantioselectivity is thought to be associated with changes in the electronic nature of the *N*-terminal amide that alters the intramolecular hydrogen bonding of the peptide.¹³

With **P34** identified as a more selective catalyst, we set out to perform an optimization of the reaction conditions. A catalyst loading screen with 10, 5, and 2.5 mol% peptide **P34** resulted in a slight increase in enantioselectivity with no loss in conversion at 4 °C in 24 hours (Table 3, entries 1–3). By reducing the reaction concentration from 0.10 M to 0.05 M in CHCl₃, **5c** was obtained in 99% yield and 88:12 er (entry 4). The enantioselectivity was further increased by lowering the reaction temperature to –15 °C (89:11 er), although at –25 °C the selectivity was not improved (entries 5–6). We chose to briefly re-examine the *i*+2 position and found that when Aib was replaced with Aic (**P40**), the product was obtained in 99% conversion and 92:8 er under the optimized reaction conditions. Additionally, by switching the dimethylamide *C*-terminal group with a diethylamide (**P41**), the lactone could be obtained with 99% yield and 94:6 er. With 0.5 mol% of **P41**, **5c** could be obtained in 88% yield and nearly the same enantioselectivity (92:8 er) with prolonged reaction time, which demonstrates both the robustness and efficiency of the pThr-embedded catalysts.³² It should be noted that when (**R**)-**1b** is used as the catalyst under the reaction conditions optimized for the peptide catalyst, **5c** is obtained in 74% yield and 77:23 er (entry 10). Although the mode of catalysis is likely the same, the ability for the peptide catalyst to engage in additional hydrogen bonding interactions offers an advantage in achieving highly selective transformations of more functionalized substrates. With the optimized conditions in hand, peptide **P41** was used to evaluate the substrate scope of the enantioselective BV of cyclobutanones.

As shown in Table 4, **P41** provides a number of *N*-substituted substrates with high enantioselectivities. In general, *N*-aryl carbamates are processed with high enantioselectivity, and electron-donating or electron-withdrawing groups in the *para*-position do not lead to significant perturbations in the enantioselectivity, such that substrates **4d–4g** were smoothly converted to the corresponding lactones in nearly quantitative yield and good enantioselectivities within 24 hours (93:7 – 94:6 er) (Table 4, entries 2–5). When the reaction was carried out with a cyclobutanone containing a *p*-*t*-butyl phenyl carbamate, the product was obtained in 97% yield and 94:6 er (entry 6). Additionally, the dioxolane product can be isolated in 96% yield and 93:7 er (entry 7). Benzyl or alkyl substituted carbamates are generally less well tolerated, requiring longer reaction times (72–84 hours) and decreased enantioselectivity (82:18 – 85:15 er) (entries 8–9). Using the carbamate-functionalized analog of **4b**, which contains a methyl group at quaternary position C3, lactone **5i** can be isolated in 94% yield and 80:20 er, albeit with increased reaction time (entry 10). Lastly, we were able to establish the absolute configuration of the lactone products **5e** and **5f** by X-ray diffraction (see SI for details).

Further studies were then undertaken to gain a better understanding of the apparent directing group effects and key substrate-catalyst interactions that are required to achieve a highly enantioselective transformation. To probe the putative hydrogen bonding with the *i*+3 Dap residue, the phenylalanine analog **P42** was prepared, containing only a single-point mutation (Phe) at the *i*+3 position (Figure 3a). Various substrates were then subjected to the optimized reaction conditions using 2.5 mol% of **P41** or **P42** for 24 hours. In agreement with our earlier observations, catalyst **P42** (Phe) afforded product **5c** in 78% yield and 79:21 er, which corresponds to a decrease in $\Delta G^\ddagger$ of 0.73 kcal·mol⁻¹ compared to **P41** (Dap) (Figure 3). When the carbamate directionality is inverted, **5m** was obtained with excellent enantioselectivity using **P41** (Dap, 91:9 er) and again diminished yield and enantioselectivity with catalyst **P42** (Phe). The same trend is observed for products **5n–5p**, wherein urea³² and amide groups are better matched for the Dap-containing peptide **P41**, which contains more functionality for attractive secondary interactions, than for **P42** containing Phe. The presence of a hydrogen bond donor is necessary for achieving high selectivities, which we believe arises in part to essential hydrogen bonding interactions occurring in the transition structure of the reaction.²⁹

Altogether, these results demonstrate a synergistic effect between the nature of the appended substrate directing group and the multifunctional nature of the peptide catalysts. It is notable that in all of the substrates tested thus far, the yields are consistently better with **P41** (Dap), which suggests associative interactions that lead to a rate acceleration for oxidations compared with **P42** (Phe). Using catalyst **P41** (Dap), with carbonate substrate **4q** lacking an N–H, **5q** was obtained in only 37% yield and 66:34 er. In contrast, catalyst **P42** (Phe) performs better, generating the product in 49% yield and 78:22 er. The difference in selectivities achieved with catalyst **P41** when the N–H of **4c** is replaced with an oxygen atom (**4q**) corresponds to a $\Delta G^\ddagger$ of 1.07 kcal·mol⁻¹ (94:6 er for **5c** and 66:34 er for **5q**). Substrates **4r** and **4a** are again obtained in better enantioselectivity with the Phe catalyst **P42**, although the phenol is obtained in higher yield with **P41** (Dap). Tuning of the *i*+3 position provides a complementary pair of catalysts that can obtain good-to-high enantioselectivities for a range of substrates through secondary NCIs.

Having demonstrated that *ortho*-directing groups lead to high enantioselectivity, we were interested in the question, whether alternative placement of the identical directing group could allow for switching of catalysis trajectories. To examine whether the peptides could recognize more distal functionality we prepared substrate **4s**, which repositions the *N*-aryl carbamate from the *ortho*- to the *meta*-position. Under the optimized conditions, lactone **5s** was obtained in 97% yield and 13:87 er with 2.5 mol% **P41** (Scheme 4). Moreover, the X-ray structure of **5s** revealed that the opposite absolute configuration was obtained, compared to substrates containing an *ortho*-directing group, now favoring the *S*-enantiomer (see SI for X-ray analysis).³⁴ The results of the divergence in the absolute stereochemistry can be explained by either (1) delivery of H₂O₂ to the diastereotopic face of the ketone with the same C–C σ -bond migration or (2) the substrate–catalyst complex repositioning the peroxy group in the Criegee intermediate ~90° to achieve the opposite, but equivalent C–C bond migration (Scheme 1). It is possible that both explanations can account for the divergence, as the addition of H₂O₂ to either face of the prochiral ketone leads to *syn* and *anti* isomers,

which can both afford either enantiomer of product. The ability for **P41** to recognize both substrates in a manner that achieves the opposite absolute stereochemistry of the lactone products (**R-5c** and **S-5s**), under otherwise identical reaction conditions, is notable.

In continuing our analysis of the effect of Dap(Ac) and Phe substitution at the *i*+3 position on enantioselectivity, we applied catalyst **P42**, bearing the Phe *i*+3 residue, to the oxidation of substrate **4s** (Figure 4a). With 2.5 mol% **P42**, the opposite enantiomer lactone (**R-5s**) is obtained in 74% yield and 66:34 er under the optimal reaction conditions, relative to **P41** (Dap).³⁵ However, it should be noted that the absolute configuration of product (**R-5s**) is the same as the γ -butyrolactone products **5** bearing an *ortho*-directing group, previously obtained from the BV reaction with **P42** (Figure 3). This observed inversion in enantioselectivity is all the more impressive given the constituent amino acids in both peptides (**P41** and **P42**) have the same stereochemical configuration, yet afford opposite enantiomers of product **5s**.^{23d} The $\Delta G^\ddagger$ of 1.3 kcal·mol⁻¹ is a consequence of how the two catalysts are able to recognize the substrate upon binding to form different substrate–catalyst complexes. Moreover, when the reaction is carried out with (**R**)-**1b** as the catalyst, (**R**)-**5s** is obtained in 86% yield and 73:27 er, which matches the absolute stereochemistry and selectivities obtained in the oxidation of **4c** with (**R**)-**1b** (Table 3, entry 10). This divergence in enantioselectivity could stem from either (1) a conformational change in the catalyst structure upon mutation of the *i*+3 position or (2) a perturbation in the secondary, non-covalent interactions between the side-chains of the peptide and the functionalized substrate. Once again, it appears that the catalyst **P42** with an *i*+3 Phe residue and (**R**)-TRIP (**1b**) achieve selectivity through steric hindrance, whereas **P41** bearing the Dap residue operates through multivalent NCIs, resulting in the products bearing opposite absolute configurations.

After realizing this intriguing divergence in the absolute sense of enantioinduction by catalysts **P41** and **P42** with **4s**, we prepared substrate **4t**, which contains a remote *para-N*-aryl carbamate directing group (Figure 4b). When we applied optimized catalyst **P41** bearing the *i*+3 Dap(Ac) residue to the oxidation of **4t**, lactone **5t** was obtained in 81% yield and diminished selectivity (39:61 er). Alternatively, phenylalanine analog **P42** afforded the product in 56% yield and 79:21 er, similar to the results obtained with (**R**)-**1b** (74% yield, 76:24 er). Once again, **P42** and (**R**)-**1b** afford the opposite absolute stereochemistry of lactone product relative to **P41**. Based on the results presented hitherto, catalyst **P42** appears to provide enantioinduction through steric repulsive interactions, whereas catalyst **P41**, containing an *i*+3 Dap residue can engage the substrate in additional hydrogen-bonding interactions. These differences in mechanism can lead to a divergence in the absolute stereochemistry of product without inversion of a catalyst stereocenter, and seem to suggest that even a remote directing group can engage in NCIs with the peptidic scaffold and perturb the enantioselectivity.

To explain the emerging trends from the structure–selectivity studies and intriguing divergence in observed enantiomeric products **5s–t**, we sought to understand the effect of *i*+3 substitution on peptide secondary structure using 2D NMR spectroscopy (Figure 5a) (see SI for details).³⁶ In the case of **P41**, a number of nOe contacts were present between the *i*+3

Dap residue and the $i+1$ D-Pro residue that indicated the Dap side chain may be orientated on the top face of the peptide turn. However, for **P42** no nOe contacts were observed between the $i+3$ Phe side chain and the peptide backbone, which suggests the phenyl ring is positioned away from the peptide turn. Yet, nOe contacts are still observed between the β_{C-H} of the i pThr residue and the β_{C-H} of the $i+3$ residues for both peptides, indicating some similarity between the positioning of the Phe and Dap residues in the peptide secondary structure. Furthermore, the orientation of the pThr(Bn) residue is likely the same in each catalyst as indicated by five additional conserved interresidue contacts present at the i catalyst position. DMSO titration studies were then conducted to elucidate the hydrogen bonding environments of each N-H proton, which further showed structural similarities between **P41** and **P42** (Figure 5b).³⁷ While the N-H(i) exhibits a subtle downfield shift, the N-H($i+2$) and N-H($i+3$) peaks remain constant and shift upfield respectively, suggesting both N-H($i+2$) and N-H($i+3$) being previously engaged in intramolecular hydrogen bonding. Together with the observed nOe contacts, these observations are suggestive of both peptides adopting a type I' double β -turn.^{18,22} It is worth noting that **P41** and **P42** appear to adopt a type I' β -turn secondary structure in solution, whereas previously **P4** adopted a type II' β -turn in the solid-state.³⁸

With these considerations in mind, a mechanistic hypothesis for the formation of the enantioenriched products could be assembled (Figure 6a). In our thinking, we preserve the type I' β -turn secondary structure in accord with our experimental observations. Given the similarity between the two structures, as determined by 2D NMR and DMSO titration studies, we believe that the incorporation of the Dap(Ac) residue does not result in a different conformation of the peptide. Rather, the $i+3$ Dap side chain of **P41** is orientated on the top face of the catalyst with the C=O of the $i+3$ side chain acetamide free to engage in a hydrogen bond with the substrate, whereas the $i+3$ Phe side chain in **P42** does not provide additional hydrogen bonding. We envision that the bifunctional phosphate moiety of pThr activates the ketone for addition of H₂O₂.²⁴ Although speculative, the peroxy group of the Criegee intermediate could engage in further hydrogen bonding with the peptide backbone amides. In the instances of switching in stereoselection, it is plausible to consider a similar binding model (Figure 6b). However, upon substrate binding with the catalyst side-chain amide, a slight change in the conformation of the peroxy group results in the opposite stereochemical outcome. Since the $i+3$ Phe side chain in **P42** is unable to engage in further hydrogen bonding, steric hindrance is the predominating factor in stereoselectivity, and thus the absolute stereochemistry of product is unchanged when the carbamate directing group is placed in the α -, m -, or p - positions. The multi-point binding of the substrate-catalyst complex allows for stabilization of the Criegee intermediate and facilitates a highly selective rearrangement, akin to those interactions found in BVO-monooxygenases.^{16c}

Conclusion:

In summary, we have reported a multifunctional Brønsted-acid pThr-based peptide catalyst for the enantioselective BV reaction of functionalized cyclobutanones with low catalyst loadings (0.5–2.5 mol%). By utilizing hypothesis-driven design, aided by detailed NMR and crystallographic studies, we have identified the inclusion of an $i+3$ Dap residue to be advantageous for achieving high selectivities through secondary non-covalent interactions

with a variety of directing groups. Moreover, the ability of the peptide to interact with a carbamate directing group in multiple positions of an aryl ring leads to an observed divergence in the absolute stereochemistry (94:6 er to 13:87 er *R/S*) of the γ -butyrolactone products (**5**) with a pThr-embedded peptide catalyst containing Dap. Permutations to the β -3 residue provide a complementary pair of catalysts for a further range of cyclobutanone substrates, which affords the products **5** in high yield and stereoselectivities, and in select cases opposite absolute configuration.³⁹ This work provides a framework for the further development of pThr-based catalysts for applications in asymmetric catalysis and modifications of complex molecules, where multiple sites of engagement are necessary to achieve selective functionalization.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Figure 1.

(a) Phosphorylation event as a post-translational modification. (b) BINOL-derived chiral phosphoric acids. (c) Peptide-mediated transfer hydrogenation of 8-aminoquinolines using pThr as a catalytic residue. (d) Ding and co-workers' pioneering Brønsted acid catalyzed Baeyer-Villiger oxidation of 3-substituted cyclobutanones. (e) Proposed mechanism for the CPA catalyzed BV reaction utilizing H₂O₂ as the oxidant.

Figure 2. Comparison of X-ray crystal structures of pThr- and Dmaa-embedded peptides. (a) X-ray structure of **P4**. Two 1,4-dioxane solvent molecules omitted for clarity (see SI for details). (b) X-ray structure of **7**. Two distinct packing polymorphs were observed in the unit cell (**7a**, **7b**), only one is shown for clarity (See SI for details). (c) Catalytically active residues (pThr and Dmaa) embedded in *i* position of conserved peptide sequence.

Figure 3.

(a) General reaction scheme for BV oxidation with peptide catalyst and **(b)** structure–selectivity study with **P41** and Phe analog (**P42**). Reaction conditions: Cyclobutanone **4** (0.05 mmol, 1.0 equiv), catalyst (2.5 mol%), and H₂O₂ (30% w/w aq. 1.5 equiv) at –15 °C with [**4c**] = 0.05 M CHCl₃. Yield was determined by ¹H NMR analysis of the crude reaction mixture using an internal standard (1,3,5-trimethoxybenzene). Enantiomeric ratios were by CSP-HPLC analysis. Reported results are the average of two trials. The absolute configuration of products **5m–r** were assigned in analogy to **5e** and **5f**. The absolute

configuration of **5a** was assigned as (*R*)-(-)-**5a** by comparison of the measured optical rotation to literature values.^{17a}

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Figure 4.

Divergences in enantioselectivity in *meta*- and *para*-*N*-aryl carbamate without inversion of a catalyst stereogenic center.^{a,b}

^a Conditions: Cyclobutanone **4** (0.05 mmol, 1.0 equiv), H_2O_2 (30% w/w aq. 1.5 equiv), catalyst (2.5 mol%) at $-15\text{ }^\circ\text{C}$ with $[\mathbf{4}] = 0.05\text{ M CHCl}_3$. ^b Yield was determined by ^1H NMR analysis of the crude reaction mixture using an internal standard (1,3,5-trimethoxybenzene), and enantiomeric ratios determined by CSP-HPLC analysis. Reported results are the average of two trials. ^c Absolute configuration of **5t** assigned in analogy to **5s**.

Figure 5.

(a) ^1H - ^1H ROESY NMR of peptide catalysts **P41** and **P42** show both unique and conserved interresidue nOe correlations (800 MHz, 5.0 mM in CDCl_3 at 25 °C). (b) ^1H NMR solvent titration curve of $\text{DMSO-}d_6$ to identify solvent exposed and hydrogen bonded amides for peptides **P41** and **P42** (5.0 mM concentration in CDCl_3 (referenced to 7.26 ppm) at 25 °C).

Figure 6.

(a) Speculative transition structures showing putative substrate-catalyst hydrogen bonding interactions with an antiperiplanar orientation of the peroxy group in the Criegee intermediate, which is consistent with the absolute configuration of the product. (b) Speculative transition structures for the switching in stereoselection in substrate **4s**, which is *meta*-substituted.

Scheme 1.
Syn/anti Criegee intermediates.

Scheme 2.

Preliminary pThr-catalyzed Baeyer–Villiger oxidation of 4a–b.

Scheme 3.
Baeyer–Villiger oxidation of carbamate **4c**.

Scheme 4. Observed reversal in absolute stereochemistry in the BV of *meta*-4s.

Table 1.

Initial Peptide sequence optimization.^{a,b}

Reaction scheme: 4b (cyclobutanone derivative) reacts with catalyst (10 mol%), H₂O₂ (30%, 1.5 equiv), and CHCl₃ (24 h) to form 5b (peptide derivative).

Catalyst structure: A cyclic peptide with side chains R², R³, R⁴, R⁵, R⁶ and a phosphate group R¹O-P(=O)(OH)-. The peptide backbone is labeled with *i*, *i* + 1, *i* + 2, and *i* + 3.

Catalyst								
entry	PG	<i>i</i>	<i>i</i> + 1	<i>i</i> + 2	<i>i</i> + 3	conv (%) ^c	er ^d	
1	P3	Fmoc	pThr(Bn)	D-Pro	Acpc	Leu-NMe ₂	87	64:36
2	P4	Fmoc	pThr(Bn)	D-Pro	Acpc	Leu-D-Phe-NMe ₂	67	66:34
3	P5	Fmoc	pThr(Bn)	D-Pro	Acpc	Phe-NMe ₂	88	64:36
4	P6	Fmoc	pThr(Bn)	D-Pro	Acpc	Ala-NMe ₂	85	62:38
5	P7	Fmoc	pThr(Bn)	D-Pro	Aib	Phe-NMe ₂	92	69:31
6	P8	Fmoc	pThr(Bn)	D-Pro	Aib	D-Phe-NMe ₂	69	52:48
7	P9	Fmoc	pThr(Bn)	Pro	Aib	Phe-NMe ₂	54	44:56
8 ^e	P8	Fmoc	pThr(Bn)	D-Pro	Aib	Phe-NMe ₂	19	51:49
9	P10	Fmoc	Thr(Bn)	D-Pro	Aib	Phe-NMe ₂	<5	n/a
10	P11	Fmoc	pThr(H)	D-Pro	Aib	Phe-NMe ₂	44	55:45
11	P12	Fmoc	<i>allo</i> -pThr(Bn)	D-Pro	Aib	Phe-NMe ₂	47	51:49
12	P13	Fmoc	pPhSer(Bn)	D-Pro	Aib	Phe-NMe ₂	98	67:33
13	P14	Fmoc	pSer(Bn)	D-Pro	Aib	Phe-NMe ₂	73	58:42
14	P15	Fmoc-Phe	pThr(Bn)	D-Pro	Aib	Phe-NMe ₂	96	70:30
15	P16	Fmoc-D-Phe	pThr(Bn)	D-Pro	Aib	Phe-NMe ₂	88	67:33
16	P17	Fmoc-Pro	pThr(Bn)	D-Pro	Aib	Phe-NMe ₂	88	72:28
17	P18	Fmoc-D-Pro	pThr(Bn)	D-Pro	Aib	Phe-NMe ₂	93	73:27
18 ^f	P18	Fmoc-D-Pro	pThr(Bn)	D-Pro	Aib	Phe-NMe ₂	73 (75) ^g	78:22
19 ^h	P18	Fmoc-D-Pro	pThr(Bn)	D-Pro	Aib	Phe-NMe ₂	50	77:23
20 ^h	(R)-1b						97	64:36

^a Reaction conditions: Cyclobutanone **4b** (0.05 mmol, 1.0 equiv) at 0 → 4 °C (ambient temperature) with [**4b**] = 0.10 M CHCl₃.^b Reported results are the average of two trials.^c Determined by ¹H NMR relative integrations.^d Enantiomeric ratios were determined by CSP-HPLC analysis (Chiralpak IA, 210 nm). Absolute configuration was not determined.^e Reaction performed with *t*-BuOOH (70% w/w aq. 1.5 equiv).^f Reaction was carried out at -15 °C.

^gYield in parentheses determined by ¹H NMR internal standard (1,3,5-trimethoxybenzene).

^hReaction was carried out at −25 °C.

Abbreviations: pThr, phosphothreonine; Bn, benzyl; Acpc, 1-aminocyclopropane carboxylic acid; Aib, 2-aminoisobutyric acid; pPhSer, *β*-threo-phosphophenylserine; pSer, phosphoserine; Bz, benzoyl.

Table 2.

Peptide *i*+3 and *N*-terminus optimization for directed BV reaction^{a,b,c}

^a Reaction conditions: Cyclobutanone **4c** (0.025 mmol, 1.0 equiv), catalyst (10 mol%), and H₂O₂ (30% w/w aq. 1.5 equiv) at 0 → 4 °C (ambient temperature) with [4c] = 0.10 M CHCl₃.

^b Conversion determined by ¹H NMR relative integrations and enantiomeric ratios determined by CSP-HPLC analysis (Chiralpak IA, 230 nm).

^cReported results are the average of two trials.

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Table 3.In depth optimization of peptide and reaction conditions.^a

entry	cat.	<i>i</i> + 2	X	cat. (mol%)	conc. [M]	T (°C)	conv. (%) ^b	er ^c
1	P34	Aib	NMe ₂	10	0.10	4	100	85:15
2	P34	Aib	NMe ₂	5	0.10	4	100	86:14
3	P34	Aib	NMe ₂	2.5	0.10	4	100	86:14
4	P34	Aib	NMe ₂	2.5	0.05	4	100	88:12
5	P34	Aib	NMe ₂	2.5	0.05	−15	99	89:11
6 ^d	P34	Aib	NMe ₂	2.5	0.05	−25	100	88:12
7	P40	Aic	NMe ₂	2.5	0.05	−15	99	92:8
8	P41	Aic	NEt ₂	2.5	0.05	−15	99	94:6
9 ^e	P41	Aic	NEt ₂	0.5	0.05	−15	88	92:8
10	(R)-1b			2.5	0.05	−15	74	77:23

^aReaction conditions: Cyclobutanone **4c** (0.05 mmol, 1.0 equiv) and H_2O_2 (30% w/w aq. 1.5 equiv). Reported results are the average of two trials (0.05 mmol).

^bConversion determined by ¹H NMR relative integrations.

^cEnantiomeric ratios were determined by CSP-HPLC analysis (Chiralpak IA, 230 nm).

^dReaction performed for 48 hours.

^eReaction performed for 96 hours.

Abbreviations: Aic, 2-aminoindane-2-carboxylic acid.

Table 4.

Carbamate substrate scope.^a

entry	R ¹	R ²	time (h)	yield (%) ^b	er ^c
1	H	C ₆ H ₅ (c)	24	99	94:6
2	H	4-OMeC ₆ H ₄ (d)	24	99	93:7
3	H	4-CF ₃ C ₆ H ₄ (e)	24	98	93:7
4	H	4-BrC ₆ H ₄ (f)	24	94	94:6
5	H	(g)	24	95	92:8
6	H	4- <i>t</i> -BuC ₆ H ₄ (h)	24	97	94:6
7	H	(i)	24	96	93:7
8	H	CH ₃ CH ₂ (j)	84	96	85:15
9	H	C ₆ H ₅ CH ₂ (k)	72	95	82:18
10	CH ₃	C ₆ H ₅ (l)	72	94	80:20

^aReaction conditions: Cyclobutanone **4** (0.10 mmol, 1.0 equiv), **P41** (2.5 mol%), and H₂O₂ (30% w/w aq. 1.5 equiv) at -15 °C with [**4c**] = 0.05 M CHCl₃. Reported results are the average of two trials.

^bIsolated yield after chromatography.

^cEnantiomeric ratios were determined by CSP-HPLC analysis.

See supporting information for details on assignment of absolute configuration. The configuration of products **5c-d**, **5g-l** were assigned in analogy to **5e** and **5f**.