Published in final edited form as:

Transl Cancer Res. 2017 March ; 6(Suppl 2): S366–S368. doi:10.21037/tcr.2017.03.26.

KDM1 links nuclear GSK3 β to epigenetic alterations

Weijie Guo and Binhua P. Zhou

Department of Molecular and Cellular Biochemistry, and Markey Cancer Center, University of Kentucky, College of Medicine, Lexington, USA

Accumulating evidence indicates that cancer stem cells (CSCs) play an important role during the processes of tumor development, progression, and recurrence and are primarily responsible for therapeutic resistance and poor clinical outcome of patients (1). Epigenetic alterations, particularly histone methylation, have been increasingly recognized as a global transcriptional regulator that contributes to stem cell self-renewal and differentiation under physiological and pathological conditions (2). For example, methylations of Lys9 and Lys27 residues of histone H3 (H3K9me_{2/3} and H3K27me₃), associate with heterochromatin and transcriptional repression; whereas H3K4me_{2/3}, often found in active gene promoters, is associated with transcriptional activation. These epigenetic modifications create unique promoter architectures that control gene expression. Glycogen synthase kinase 3 beta (GSK3 β) has been shown to take part in the regulation of histone methylation including H3K4 methylation in the promoter regions of multiple genes (3,4). However, the molecular mechanisms for GSK3 β in mediating alterations of histone methylation remain to be defined. In September 2016 issue of *Nature Cell Biology*, Zhou *et al.* (5) addressed this challenge and singled out a connection between nuclear GSK3 β and epigenetic aberration via regulation of lysine-specific histone demethylase 1A (KDM1A) stability.

Histone lysine methylation is a dynamic and reversible process. KDM1A (also known as LSD1) is the first identified lysine-specific histone demethylase that selectively remove the methyl group from H3K4me_{1/2} through flavin-adenine-dinucleotide (FAD)-dependent oxidative reaction (6). KDM1A exerts its gene repression functions by forming as a key catalytic component in several corepressor complexes, including Co-REST, NuRD, CtBP, HDAC, or SIRT1 (7,8). KDM1A is often overexpressed in breast (9), bladder, lung, colon (10) cancers and glioblastoma (11), and is required for the maintenance and differentiation of stem cells (12) and CSCs (13). Despite its determinant roles in stem cell pluripotency and differentiation and tumorigenesis, the mechanisms that lead to KDM1A dysregulation in these tumors remain to be better delineated.

Correspondence to: Binhua P. Zhou, MD, PhD, Professor. Department of Molecular and Cellular Biochemistry, Markey Cancer Center, University of Kentucky School of Medicine, BioPharm Rm# 581, 789 South Limestone, Lexington, KY 40536, USA. peter.zhou@uky.edu.

Conflicts of Interest: The authors have no conflicts of interest to declare.

Provenance: This is an invited Editorial commissioned by Section Editor Chen Qian (Center for Inflammation & Epigenetics, Houston Methodist Hospital Research Institute, Houston, TX, USA).

Comment on: Zhou A, Lin K, Zhang S, *et al.* Nuclear GSK3 β promotes tumorigenesis by phosphorylating KDM1A and inducing its deubiquitylation by USP22. *Nat Cell Biol* 2016;18:954–66.

Consistent with the notion that KDM1A is a labile protein that under constant protein ubiquitination and degradation (6,9), Zhou *et al.* found that nuclear GSK3 β stabilizes KDM1A by decreasing its ubiquitylation. GSK3 β prefers substrates that are primed phosphorylated by another kinase at a Serine or Threonine residue located 4 or 5 amino acids downstream or upstream of GSK3 β phosphorylation site (14). The authors found that KDM1A contains several potential GSK3 β phosphorylation sites harboring a canonical -S/TxxxS/T-motif. They further identified that Ser687 of KDM1A is phosphorylated by casein kinase 1 alpha (CK1 α) and this priming phosphorylation facilitates GSK3 β phosphorylation at Ser683. As GSK3 β does not directly deubiquitylate KDM1A, the authors proposed that there should be deubiquitylase (DUB) to cooperate the deubiquitylation of KDM1A. Thus, they screened a panel of DUBs and identified ubiquitin-specific protease 22 (USP22) as a direct DUB for KDM1A. Subsequently, the authors explored the roles of GSK3 β -USP22-KDM1A axis in glioma. They showed that GSK3 β -USP22-KDM1A axis is required for tumorigenesis of CSCs and associated with the grade of human glioma malignance.

A picture appears in which KDM1A connects nuclear GSK3 β with epigenetics (Figure 1). First, KDM1A is phosphorylated by CK1 α at Ser687 which facilitates its binding with and phosphorylation by GSK3 β . Subsequently, USP22, a direct KDM1A DUB, deubiquitylates KDM1A. Consequently, stabilized KDM1A removes the activation marks on H3K4 in promoter regions of stem cell pluripotency inhibition genes, including *BMP2*, *CDKN1A* and *GATA6*. Finally, as reported here, silencing of these tumor suppressors promotes tumorigenesis of CSCs. More importantly, the authors also showed therapeutic potential of tideglusib, a selective non-ATP-competitive GSK3 inhibitor with good blood-brain barrier penetration and now in a phase II trial for the treatment of Alzheimer's disease, along or in combination with temozolomide, a FDA approved drug for glioma. Although accumulating numbers of oncogenes have been extensively characterized, many of these oncogenes are not good drug targets. Thus, there are urgent needs for alternative therapeutic approaches to interfere these oncogenes. As KDM1A is structurally related monoamine oxidases A & B (MAO-A and MAO-B), a KDM1A inhibitor may exhibit side-effect by suppressing MAO-A and MAO-B. In addition, blood-brain barrier makes it harder for KDM1A as a therapeutic target for treating glioma. Thus, targeting the proposed GSK3 β -USP22-KDM1A signaling axis represents an excellent approach in suppressing the function of KDM1A in glioma. Recent works suggest that onco-DUBs, which contribute substantially to the stability of onco-proteins and pathways and are amenable to pharmacologic inhibition by small molecules, possess great potential for cancer treatment (15). Though not explored in their work, USP22, may also serve as a promising target for treating glioma and deserve further investigation. Nevertheless, the therapeutic potential established in this outstanding study will warrant further translational investigation and inspire many areas of basic inquiry that will further our understanding of the concerted connections between aberrant cell signaling and epigenetics during malignant tumor progression.

Acknowledgements

Funding: This work was supported by the Shared Resources of the University of Kentucky Markey Cancer Center (P30CA177558), grants from NIH (CA125454 and CA188118), DoD (BC140733P1), and Mary Kay Ash Foundation.

References

1. Nguyen LV, Vanner R, Dirks P, et al. Cancer stem cells: an evolving concept. *Nat Rev Cancer* 2012;12:133–43. [PubMed: 22237392]
2. Klose RJ, Zhang Y. Regulation of histone methylation by demethylation and demethylation. *Nat Rev Mol Cell Biol* 2007;8:307–18. [PubMed: 17342184]
3. Hill EV, Ng TH, Burton BR, et al. Glycogen synthase kinase-3 controls IL-10 expression in CD4(+) effector T-cell subsets through epigenetic modification of the IL10 promoter. *Eur J Immunol* 2015;45:1103–15. [PubMed: 25627813]
4. Ougolkov AV, Bone ND, Fernandez-Zapico ME, et al. Inhibition of glycogen synthase kinase-3 activity leads to epigenetic silencing of nuclear factor kappaB target genes and induction of apoptosis in chronic lymphocytic leukemia B cells. *Blood* 2007;110:735–42. [PubMed: 17463171]
5. Zhou A, Lin K, Zhang S, et al. Nuclear GSK3 β promotes tumorigenesis by phosphorylating KDM1A and inducing its deubiquitylation by USP22. *Nat Cell Biol* 2016;18:954–66. [PubMed: 27501329]
6. Shi YJ, Matson C, Lan F, et al. Regulation of LSD1 histone demethylase activity by its associated factors. *Mol Cell* 2005;19:857–64. [PubMed: 16140033]
7. Mulligan P, Yang F, Di Stefano L, et al. A SIRT1-LSD1 corepressor complex regulates Notch target gene expression and development. *Mol Cell* 2011;42:689–99. [PubMed: 21596603]
8. Wang Y, Zhang H, Chen Y, et al. LSD1 is a subunit of the NuRD complex and targets the metastasis programs in breast cancer. *Cell* 2009;138:660–72. [PubMed: 19703393]
9. Wu Y, Wang Y, Yang XH, et al. The deubiquitinase USP28 stabilizes LSD1 and confers stem-cell-like traits to breast cancer cells. *Cell Rep* 2013;5:224–36. [PubMed: 24075993]
10. Hayami S, Kelly JD, Cho HS, et al. Overexpression of LSD1 contributes to human carcinogenesis through chromatin regulation in various cancers. *Int J Cancer* 2011;128:574–86 [PubMed: 20333681]
11. Sareddy GR, Nair BC, Krishnan SK, et al. KDM1 is a novel therapeutic target for the treatment of gliomas. *Oncotarget* 2013;4:18–28. [PubMed: 23248157]
12. Wang J, Hevi S, Kurash JK, et al. The lysine demethylase LSD1 (KDM1) is required for maintenance of global DNA methylation. *Nat Genet* 2009;41:125–29. [PubMed: 19098913]
13. Wang J, Lu F, Ren Q, et al. Novel histone demethylase LSD1 inhibitors selectively target cancer cells with pluripotent stem cell properties. *Cancer Res* 2011;71:7238–49. [PubMed: 21975933]
14. Kaidanovich-Beilin O, Woodgett JR. GSK-3: Functional Insights from Cell Biology and Animal Models. *Front Mol Neurosci* 2011;4:40. [PubMed: 22110425]
15. Potu H, Peterson LF, Pal A, et al. Usp5 links suppression of p53 and FAS levels in melanoma to the BRAF pathway. *Oncotarget* 2014;5:5559–69. [PubMed: 24980819]

Figure 1.

Schematic illustration of GSK3β-USP22-KDM1A axis in the regulation of tumorigenesis. Ubiquitylated KDM1A either enters the proteasomal degradation. Alternatively, nuclear GSK3β phosphorylates KDM1A after the priming phosphorylation by CK1α. Phosphorylation of KDM1A by GSK3β enhances its binding with and deubiquitylation by USP22, leading to its stabilization. Stabilized KDM1A promotes demethylation of histone H3K4 and suppresses the expression of *BMP2*, *CDKN1A* and *GATA6*, which in turn favors tumorigenesis.