RESEARCH ARTICLE

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2018, 9, 15472-Aminobenzimidazoles as antibiofilm agents
against *Salmonella enterica* serovar Typhimurium†William M. Huggins,^a T. Vu Nguyen,^a Nicholas A. Hahn,^a James T. Baker,^a
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Serovars within the species *Salmonella enterica* are some of the most common food and water-borne pathogens worldwide. Some *S. enterica* serovars have shown a remarkable ability to persist both inside and outside the human body. *Salmonella enterica* serovar Typhi can cause chronic, asymptomatic infection of the human gallbladder. This organism's ability to survive inside the gallbladder centers around its ability to form biofilms on gallstone surfaces. Currently, chronic carriage of *S. Typhi* is treated by invasive methods, which are not well suited to areas where *Salmonella* carriage is prevalent. Herein, we report 2-aminobenzimidazoles that inhibit *S. enterica* serovar Typhimurium (a surrogate for *S. Typhi*) biofilm formation in low micromolar concentrations. Modifications to the head, tail, and linker regions of the original hit compound elucidated new, more effective analogues that inhibit *S. Typhimurium* biofilm formation while being non-toxic to planktonic bacterial growth.

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Salmonella species are a frequent cause of food and water borne illness worldwide. They can cause a variety of disease syndromes, and are normally grouped into typhoidal and non-typhoidal species. *Salmonella enterica* serovar Typhi is the causative agent of enteric fever and *Salmonella enterica* serovar Typhimurium is typically associated with intestinal distress, or salmonellosis.¹ Typhoidal and non-typhoidal strains of *Salmonella* have shown a remarkable ability to persist in a variety of environments, including harsh environments within the human body. One of these survival strategies centers around the ability of *Salmonella* to form biofilms on gallstones.² Colonization of gallstones can then lead to chronic carriage of *S. Typhi*, which allows for the dissemination of *Salmonella* through fecal shedding. Biofilms can be up to 1000-fold more resistant to antibiotic treatment than their planktonic counterparts, rendering typical antibiotic treatment regimens ineffective at eradicating chronic carriage of *Salmonella*.^{3–5} Presently, chronic carriage of *Salmonella* is treated by invasive methods, which are not well suited to areas where *Salmonella* is prevalent.⁶ New treatment options for chronic carriage of *Salmonella* are needed, and compounds that perturb *Salmonella* biofilms could be a viable treatment option.

Recently, there have been reports of derivatized 2-aminoimidazoles (2-AIs) that inhibit *S. Typhimurium* biofilm formation.^{7–12} As our group has been deeply involved in exploring the antibiofilm potential of 2-AIs, we were curious if there were 2-AI derivatives in our library, or derivatives of other nitrogen-dense heterocycles that we have assembled and assayed for their antibiofilm activities, that possessed activity against *S. Typhimurium*. As *S. Typhi* is a human-specific pathogen, reliable murine pathogenic serovars such as *S. Typhimurium* have been used to model *S. Typhi* infections, allowing for future *in vivo* testing.^{13,14} We performed an initial library screen for inhibitors of *S. Typhimurium* biofilm formation and identified 2-aminobenzimidazole (2-ABI) compound **1** as one lead compound that returned an IC₅₀ value of 13.1 ± 0.6 μM. Previously, we have shown that 2-ABIs display a wide variety of biological activity including antibiotic activity against MRSA and MDR *A. baumannii*,¹⁵ antibiofilm activity against Gram-positive bacteria,¹⁶ and the ability to potentiate β-lactam antibiotic activity against

Fig. 1 Structure of original hit compound **1**, regions of modification.

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Mycobacterium smegmatis and *M. tuberculosis*.¹⁷ With this compound in hand, we decided to probe the structure–activity relationship (SAR) of the 2-ABIs against *S. Typhimurium* biofilms. Herein we report the results of this SAR study of the 2-ABI scaffold, focusing on three regions: the head region, the linker and the tail region (Fig. 1).

The first region of the molecule we decided to modify was the head region of the 2-ABI using a synthetic scheme previously described by our group (Scheme 1A).¹⁸ Briefly, 4-fluoro-3-nitroaniline **2** was acylated with 4-pentylbenzoyl chloride in the presence of triethylamine and 4-dimethylaminopyrimidine (DMAP) in DCM for 16 hours at room temperature to yield compound **3**. S_NAr substitution of compound **3** with commercially available amines in refluxing ethanol for 16 hours yielded compounds **4a–n**. Subsequent reduction of the nitro group with ammonium formate and 10% Pd/C in ethanol at reflux followed by cyclization with cyanogen bromide in DCM at room temperature yielded 2-ABIs **5a–n**. The unsubstituted 2-ABI derivative, compound **5o**, was prepared using a previously published method.^{15,19}

Three head group substitutions, *n*-octyl (**5a**), butyl phenyl (**5b**), and *n*-hexyl (**5c**) displayed improved activity (Table 1) over the parent compound. Isobutyl substitution (compound

5d) or substitution with tryptamine (**5e**) returned an IC₅₀ value between 10–15 μM, similar to that of parent compound **1** (IC₅₀ values were only quantified for compounds that had marked improvement over compound **1**). Substitution with cyclopentyl (**5g**) or cyclohexyl rings (**5h**) returned slightly higher IC₅₀ values compared to the parent (15–20 μM). Shortening the phenyl chain to two methylenes also reduced activity, with compound **5i** returning an IC₅₀ value of 20–40 μM (ESI[†]). Replacement of the phenyl ring on compound **5i** with an imidazole (**5j**) ring also returned an IC₅₀ value of 20–40 μM. Alkyl chains with less than four carbon atoms (**5k**, **5l**) or longer than eight carbon atoms (**5m**, **5n**) reduced or completely abolished antibiofilm activity. Removal of the head group (compound **5o**) from the 2-ABI lowers the IC₅₀ from 13.1 ± 0.6 μM to 20–40 μM, demonstrating the necessity of the head group for biofilm inhibitory activity.

After investigating different head group substitutions in the *para* position relative to the amide, analogues with the head group substitution *meta* to the amide were prepared. The synthetic route to these compounds (Scheme 1B), is identical to that of the *para* analogues (Scheme 1A) except the starting material is 3-fluoro-4-nitroaniline **6**. Compared to

Scheme 1 (A) Synthetic route to compounds **5a–n** and (B) synthetic route to compounds **9a–e**: (a) 4-pentylbenzoyl chloride, DMAP, Et₃N, DCM, 16 h (b) RNH₂, EtOH, reflux, 16 h (c) NH₄HCO₂, 10% Pd/C, EtOH, reflux, 3 h (d) CNBr, DCM, 16 h (e) HCl, MeOH.

Table 1 IC₅₀ values of *para* 2-ABI, compounds **5a–h**, and *meta* 2-ABI, compounds **9a–b**. Full inhibition data can be found in the ESI

Compound	R=	IC ₅₀ (μM)
5a		6.66 ± 0.32
5b		9.59 ± 0.23
5c		10.8 ± 0.56
1		13.1 ± 0.6
5d		10–15
5e		10–15
5f		15–20
5g		15–20
5h		15–20
9a		10–15
9b		10–15

their *para* substituted counterparts, the *meta* analogues did not display a significant increase in activity (Table 1). Only the butyl analogue **9a** (IC₅₀ 10–15 μM) displayed increased activity compared to butyl *para* analogue **5f** (IC₅₀ 15–20 μM). *n*-Hexyl analogue **9b** displayed almost identical activity to the *para* *n*-hexyl analogue **5c**, returning an IC₅₀ of 10–15 μM. Methyl (**9c**), ethyl (**9d**), and isopropyl (**9e**) analogues all displayed IC₅₀ values of 20–40 μM (ESI†).

After modifying the head region of the molecule, substitutions to the tail of the molecule were made in an effort to

Fig. 2 Structure of compounds **10a–t** with various tail substitutions.**Table 2** IC₅₀ values of 2-ABIs with different tails, compounds **10a–n**. Full inhibition data can be found in the ESI

Compound	R=	IC ₅₀ (μM)
10a		5.22 ± 0.11
10b		5.38 ± 0.79
10c		5.58 ± 0.21
10d		6.30 ± 1.29
1		13.1 ± 0.6
10e		10–15
10f		10–15
10g		10–15
10h		10–15
10i		10–15
10j		15–20
10k		15–20
10l		15–20
10m		15–20
10n		15–20

improve the antibiofilm ability of the parent compound. Previously, various 2-ABI derivatives with an identical head group to the parent compound **1** and various tails were prepared (Fig. 2).¹⁷ As these compounds were readily available in our laboratory, their anti-*Salmonella* biofilm activity was investigated. Halogenation (**10b**, **10c** and **10d**) of the aromatic ring lowers the IC_{50} value when compared to a 4-pentylbenzoyl group, all returning IC_{50} values of less than 7 μ M (Table 2). The 3,5-dichloro analogue **10e** displays lower activity compared to the other halogenated tails, but displays essentially equivalent activity to the parent compound **1**. Ad-

dition of a pentoxybenzoyl group (**10a**) returns the lowest IC_{50} observed, with an IC_{50} of 5.22 ± 0.11 μ M. Although substituted benzoyl groups are favored in the most active compounds, removal of the aromatic ring in favor of a straight alkyl chain is tolerated as observed with nonanoyl (**10h**) and decanoyl (**10i**) tails. 4-Butylbenzoyl (**10f**), and 4-propoxybenzoyl (**10g**) tails all returned an IC_{50} value of 10–15 μ M, similar to that of the parent compound (**1**) and the 3,5-dichlorobenzoyl compound (**10e**). Additional analogues were investigated, but all displayed lower activity in comparison to the parent compound. 4-Butoxybenzoyl (**10j**),

Scheme 2 (A) Synthetic route to compound **15** and (B) synthetic route to compound **20**: reagents and conditions (a) $SOCl_2$, MeOH, 0 °C to rt, 16 h (b) 3-phenyl-1-propylamine, EtOH, reflux, 16 h (c) NH_4HCO_2 , 10% Pd/C, EtOH, reflux, 3 h (d) CNBr, DCM, 16 h (e) NaOH, 1:1 MeOH : H_2O , reflux, 2 h (f) EDC, 3,4-dichloroaniline, Et_3N , DMAP, 2:1 DCM : DMF, rt, 16 h (g) HCl, MeOH (h) Boc_2O , DMAP, Et_3N , DCM, rt, 16 h (i) Alloc-Cl, Et_3N , $Sc(OTf)_3$, DCM, rt, 16 h (j) 30% TFA : DCM, rt, 4 h (k) 3,4-dichloroaniline, triphosgene, Na_2CO_3 , DCM- H_2O , rt, 16 h (l) $Pd(PPh_3)_4$, $NaBH_4$, EtOH, 0 °C to rt, 1 h (m) 12N HCl, pH 1, rt, 2 h.

4-propylbenzoyl (**10k**), 4-hexylbenzoyl (**10l**), 4-heptylbenzoyl (**10m**), and octanoyl (**10n**) tails all returned IC_{50} values of 15–20 μ M. 4-Ethylbenzoyl (**10o**) and hexanoyl (**10p**) tails returned IC_{50} values between 20–40 μ M (ESI[†]). Lastly, heptanoyl (**10q**), 4-octylbenzoyl (**10r**), undecanoyl (**10s**), and tridecanoyl (**10t**) tails returned IC_{50} values of greater than 40 μ M.

The last region of the 2-ABIs that we were interested in modifying was the linker region. The 2-ABIs previously synthesized in our laboratory^{15–18} that served as the basis for the 2-ABIs used in this study have featured an amide linkage, with the nitrogen in the amide connected to the 2-ABI head of the molecule. Previous studies on other 2-AI natural products have demonstrated the effect that modification of the amide moiety can have on a compound's ability to control bacterial behavior as well as its toxicity in *C. elegans*.²⁰ With this in mind, we aimed to synthesize 2-ABI derivatives with a reverse amide moiety as well as a urea moiety replacing the amide. Compound **10d** was chosen as the compound for further analog development due to its predicted reduced metabolic liabilities.

The synthesis of the reverse amide analogue (Scheme 2A) began by reacting 4-fluoro-3-nitrobenzoic acid with thionyl chloride in methanol at 0 °C, warming to room temperature overnight to yield the methyl ester **12**. Compound **12** was then reacted with 3-phenyl-1-propylamine in ethanol at reflux to yield **13**. Reduction of the nitro group followed by cyclization with cyanogen bromide in DCM at room temperature yielded the 2-ABI **14**. Lastly, saponification of the methyl ester with sodium hydroxide in 1:1 MeOH/H₂O followed by EDC coupling of the carboxylic acid with 3,4-dichloroaniline delivered the reverse amide 2-ABI **15**.

Synthesis of the urea analogue (Scheme 2B) began with the Boc protection of 4-fluoro-3-nitroaniline, yielding **16**. Compound **16** was then reacted with 3-phenyl-1-propylamine to yield the diaminobenzene **17**. Reduction of the nitro group using ammonium formate and 10% Pd/C in ethanol at reflux followed by cyclization with cyanogen bromide in DCM at room temperature yielded Boc-protected 2-ABI **18**. Alloc protection of the 2-ABI head proceeded smoothly utilizing scandium(III) triflate as a Lewis-acid catalyst in DCM overnight at room temperature, yielding alloc protected 2-ABI **19**. Boc deprotection of compound **20** followed by coupling with 3,4-dichloroaniline using triphosgene yielded the Alloc protected 2-ABI urea. Finally, Alloc deprotection with Pd(PPh₃)₄ and NaBH₄ yielded the target 2-ABI urea **20**.

While the reverse amide analogue **15** displayed a two-fold greater IC_{50} ($12.6 \pm 1.8 \mu$ M) than that of the parent compound **10d**, the urea analogue **20** displayed only a slight reduction in IC_{50} value from 6.30 ± 1.29 to $7.69 \pm 0.25 \mu$ M, demonstrating that the linker group of the 2-ABIs can be modified while retaining biofilm inhibitory activity against *S. Typhimurium*. Additionally, compounds **10a** and **20** showed no toxicity to planktonic bacterial growth at their IC_{50} values, indicating that they are non-toxic biofilm inhibitors (ESI[†]).

Conclusions

After the identification of 2-ABI compound **1** as a *S. Typhimurium* biofilm inhibitor, probing of the SAR of the 2-ABIs elucidated six new analogues with IC_{50} values of less than 10 μ M. Utilizing the same *para*-pentyl benzoyl tail as the parent compound **1**, structural modifications to the head of the 2-ABIs in the *para* position to the amide yielded compounds **5a** and **5b** with IC_{50} values of 6.66 ± 0.32 and $9.59 \pm 0.23 \mu$ M respectively. Modification of the tail region yielded compounds **15a** ($5.22 \pm 0.11 \mu$ M), **15b** ($5.38 \pm 0.79 \mu$ M), **15c** ($5.58 \pm 0.21 \mu$ M), and **15d** ($6.3 \pm 1.29 \mu$ M) that displayed improved activity. Modification of the linker between the head and the tail yielded the urea compound **20** with a comparable IC_{50} ($7.69 \pm 0.25 \mu$ M) to that of the amide parent compound, **10d** ($6.30 \pm 1.29 \mu$ M). Substitution the 2-ABI head at the *meta* position to the amide (**9a–e**) and reversal of the amide moiety (**15**) did not produce an increase in activity. Compounds **10a** and **20** were then shown to be non-toxic to planktonic bacterial growth at their IC_{50} values, demonstrating that they show specific anti-*S. Typhimurium* biofilm inhibitory activity. Additionally, compounds **10d** and **10e** and other related 2-ABIs¹⁷ displayed no toxicity to *G. mellonella* at 400 mg kg^{−1} (ESI[†]), thus making them potential candidates for future *in vivo* studies.

Conflicts of interest

There are no conflicts to declare.

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