Evaluating optimal therapy robustness by virtual expansion of a sample population, with a case study in cancer immunotherapy

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Cancer is a highly heterogeneous disease, exhibiting spatial and temporal variations that pose challenges for designing robust therapies. Here, we propose the VEPART (Virtual Expansion of Populations for Analyzing Robustness of Therapies) technique as a platform that integrates experimental data, mathematical modeling, and statistical analyses for identifying robust optimal treatment protocols. VEPART begins with time course experimental data for a sample population, and a mathematical model fit to aggregate data from that sample population. Using nonparametric statistics, the sample population is amplified and used to create a large number of virtual populations. At the final step of VEPART, robustness is assessed by identifying and analyzing the optimal therapy (perhaps restricted to a set of clinically realizable protocols) across each virtual population. As proof of concept, we have applied the VEPART method to study the robustness of treatment response in a mouse model of melanoma subject to treatment with immunostimulatory oncolytic viruses and dendritic cell vaccines. Our analysis (*i*) showed that every scheduling variant of the experimentally used treatment protocol is fragile (nonrobust) and (*ii*) discovered an alternative region of dosing space (lower oncolytic virus dose, higher dendritic cell dose) for which a robust optimal protocol exists.

robust therapies | cancer treatment | mathematical modeling | virotherapy | immunotherapy

Heterogeneity is a defining feature of cancer (1, 2). Interpatient heterogeneity manifests clinically in variable disease progression and treatment response between patients with the same diagnosis, whereas intrapatient heterogeneity describes variations that exist between tumor cells in a single patient. Intrapatient heterogeneity can be broken down further into intratumor heterogeneity, intrametastatic heterogeneity, intermetastatic heterogeneity, and temporal heterogeneity (2, 3). Intratumor heterogeneity is evident through the presence of multiple genetic subclones within a primary tumor (2), which have even been shown to exist in spatially distinct regions of the primary tumor (4). Intrametastatic heterogeneity is similar to intratumor heterogeneity, but describes heterogeneity within a single metastatic lesion instead of within the primary tumor (2). Intermetastatic heterogeneity, on the other hand, describes variations in subclones between different metastases in the same patient (2). Finally, temporal heterogeneity is defined as changes that take place in the tumor over time, whether they are a result of genomic instability, natural selection, non-Darwinian evolution, or selective pressures imposed by treatment (4–6). Note that, in each case, heterogeneity need not be genetic but may also be epigenetic, phenotypic, or microenvironmental (2, 7, 8).

For decades, cancer patients have been treated using standard of care, meaning they receive the best known treatment that has been deemed as efficacious and safe in epidemiological studies (9). However, in the face of such interpatient and intrapatient heterogeneity, standard of care fails to induce a strong antitumor response in some patients, and often loses its efficacy with time (7, 9). The notion of personalized medicine has emerged as an

alternative to standard therapy, and holds the promise of improving clinical care by using patient-specific data to tailor treatment protocols (5, 7, 9–11). The potential benefits of this approach are multifaceted, and include the expectation of a strong patient response with a minimal toxicity profile (5, 9, 10).

As heterogeneity presents challenges for standard of care therapy, it also poses challenges for personalized therapy. If a therapeutic protocol is individually tailored based on measurements from one region of a patient's tumor at a particular pretreatment time point, it is nontrivial to determine how regions of the primary tumor distinct from the biopsy region, not to mention metastatic legions, will respond to that protocol. Given that both standard of care and personalized cancer treatment protocols must confront the challenge of high levels of variability between and within patients, the design of robust therapeutic protocols is of the utmost importance. A robust treatment should result in the same qualitative response despite a reasonable level of uncertainty (12). In other words, it would be expected to exhibit an antitumor response across a large fraction of patients, and would be more likely to be effective across the range of spatial and temporal variations observed within an individual patient. In this work, we propose to study treatment robustness through a technique that we call Virtual Expansion of Populations for Analyzing Robustness of Therapies (VEPART, for short). VEPART provides a platform for assessing the robustness of treatment

Significance

A successful cancer therapy induces a strong antitumor response while causing minimal side effects. The heterogeneous nature of cancer observed across different regions of the primary tumor, across metastatic sites, across time, and across patients makes designing such a successful therapy challenging. Both standard of care and finely tailored treatment protocols run the risk of not exhibiting a robust antitumor response in the face of these uncertainties. Here we introduce a platform for exploring this robustness question using treatment response data from a sample population. Our method integrates these experimental data with statistical and mathematical techniques, allowing us to quantify therapeutic robustness. Using this approach, we identified a robust therapeutic protocol that combines oncolytic viruses with an immunotherapeutic vaccine.

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requires the identification of the optimal protocol (possibly among a list of clinically viable options) for each of the virtual populations, followed by an analysis of responses to the optimal protocol(s) across the virtual populations.

Case Study: Immunoenhanced Oncolytic Viruses with Dendritic Cell Vaccines

As a case study, we have applied this methodology to the experimental data of Huang et al. (31) on a mouse model of melanoma treated with oncolytic viruses (OVs) and dendritic cell (DC) vaccines. OVs are standard viruses genetically engineered to selectively replicate in cancer cells. Upon integration into a tumor cell, OVs replicate and, as part of their normal life cycle, lyse the cell (32). Lysis results in the release of more OVs that can infect additional tumor cells, spreading an infection throughout the tumor that, in theory, results in tumor regression while sparing normal cells. These OVs can be further genetically enhanced to act as a vector for delivering therapeutic genes to the tumor site (32). In ref. 31, an oncolytic virus, the adenovirus (Ad) in particular, is engineered to deliver genes that boost the immune system's ability to identify, target, and kill cancer cells. The transgenes of interest in this study are 4-1BB ligand (4-1BBL) and interleukin (IL)-12; 4-1BBL is a costimulatory molecule expressed on antigen presenting cells. Binding of 4-1BBL to its receptor, 4-1BB, promotes the development and expansion of type-1 T helper cells and cytolytic effector T cells (31). IL-12 is a cytokine that strongly promotes the differentiation of naïve $CD4^+$ T cells to type-1 T helper cells (33).

Huang et al. (31) have demonstrated that oncolytic adenoviruses can successfully infect cancer cells, locally deliver therapeutic genes, and cause tumor regression in their mouse model (Fig. 2A). Further, the therapy has been shown to have minimal side effects, as lysis and high expression of therapeutic genes were restricted to cancer cells (31). The efficacy of this treatment protocol has been shown to be enhanced by the use of DC vaccines (Fig. 2B). DCs are antigen-presenting cells that stimulate naïve T cells and generate memory T cells (31). Huang et al. (31) harvested these DCs from the bone marrow of mice, pulsed them *ex vivo* with tumor-associated antigens, and delivered them intratumorally to melanoma-bearing mice. This treatment exhibited antitumor activity in isolation, and potent antitumor effects in combination with oncolytic adenoviruses carrying 4-1BBL and IL-12 (31), which we will refer to as Ad/4-1BBL/IL-12 for brevity (Fig. 2B).

We have previously developed a mathematical model that describes well the data from Huang et al. (31) in all experimentally explored situations (Fig. 2): treatment with only OVs, OVs enhanced with one or more immunostimulatory molecules (4-1BBL, IL-12, or both), DC vaccines, and DC vaccines coupled with Ad/4-1BBL/IL-12 (34, 35). In this prior work, we used the best-fit parameters to address the question of the optimal treatment ordering when administering three doses of Ad/4-1BBL/IL-12 with three doses of the DC vaccine. We predicted that exactly one strategy, front loading the cytokine-bearing OVs (OV–OV–OV–DC–DC–DC), resulted in tumor eradication (35). However, upon further exploration, we found that the model displayed some unintuitive responses to altering the dosing order. For instance, the ordering of the protocols from maximal to minimal tumor response displayed no discernible pattern (35). A striking example of this phenomenon was when the optimal protocol became the worst-case scenario by simply moving the DC dose on the last day of treatment to day one (DC–OV–OV–OV–DC–DC). This extreme sensitivity to dosing order could be explained by our finding that the doses of Ad/4-1BBL/IL-12 and DCs used in the experiments of Huang et al. (31) were near a bifurcation point in our mathematical model. As a result, slightly altering the dose or sequence drastically changed the efficacy of the protocol (35).

Herein, we aim to gain insight into the unintuitive behavior observed in our single-population optimization study by performing a robustness analysis using VEPART. Specifically, we generated a large number of bootstrap replicates by sampling with replacement the experimental data in Huang et al. (31). As detailed in *Computational Methods*, pseudorandom sampling of the posterior distributions for the fit parameters approximated from these bootstrap replicates was used to generate 1,000 virtual populations. The procedure for finding the optimal dosing sequence (among a subset of protocols) was repeated for each virtual population. This process allows us to determine the probability that different protocols will successfully result in tumor eradication, and further allows us to compute the likelihood that a particular sequencing of drugs will be optimal across the virtual populations. We undertook this analysis in three different regions of dosing space: (i) at the experimentally used dose (31), (ii) at a 50% higher dose of Ad/4-1BBL/IL-12 but a 50% lower dose of DC than used in ref. 31, and (iii) at a 50% lower dose of Ad/4-1BBL/IL-12 but a 50% higher dose of DC than used in ref. 31.

Fig. 2. Experimental data from Huang et al. (31) in which mice with B16-F10 subcutaneous tumors are intratumorally injected with different treatments. Data points represent mean tumor volume $\pm$ SE in each group of six to nine mice. All injections occur on days 0, 2, and 4, unless otherwise specified. (A) Treatments include injection of PBS (control), injection of 10^{10} OVs, injection of 10^{10} OVs carrying the 4-1BBL transgene, injection of 5×10^9 OVs carrying the IL-12 transgene, and injection of 5×10^9 OVs carrying both the 4-1BBL and IL-12 transgene. (B) Treatments include injection of PBS, injection of 10^6 DCs on days 1, 3, and 5, injection of 2.5×10^9 Ad/4-1BBL/IL-12, and injection of 2.5×10^9 Ad/4-1BBL/IL-12 on days 0, 2, and 4 along with 10^6 DCs on days 1, 3, and 5. Also shown are the best-fit solution curves from the model presented in Eqs. 1–6 using the fitting procedure detailed in this work.

This robustness analysis uncovered something unexpected: Every scheduling variant of the experimentally used treatment protocol that involves administering three doses of Ad/4-1BBL/IL-12 and DCs is fragile (nonrobust). To detail, the protocol determined to be optimal at the experimentally used dose (OV–OV–OV–DC–DC–DC) only results in tumor eradication in 30% of the virtual populations. All suboptimal protocols are even less effective. The regime of higher OV and lower DC dose also proved to be fragile, as the optimal protocol leads to tumor eradication in only 43% of the virtual populations. On the contrary, a robust optimal protocol exists when treating with higher doses of DCs and lower doses of OVs than those used by Huang et al. (31). In this dosing regime, the protocol of front loading the DCs (DC–DC–DC–OV–OV–OV) not only proved to be optimal in all virtual populations but also resulted in tumor eradication in more than 84% of those virtual populations. This application serves as proof of concept that computational robustness studies, underpinned by experimental data, can aid in identifying treatment protocols predicted to have strong antitumor properties despite interpatient (and intrapatient) heterogeneity.

Results

The full system of six differential equations described in Eqs. 1–6 includes 14 parameters. Previously, we found that the value of 7 of the 14 parameter values (viral production rate, infected cell lysis rate, viral decay rate, naive and cytotoxic T-cell decay rates, base T-cell killing rate, and T-cell differentiation rate) could be reasonably approximated from the literature (Table S1), whereas the other 7 could not. Therefore, it is these 7 parameters that were fit to give the best-fit solution curves to the experimental data, as shown in Fig. 2.

The best-fit parameters for the full model (Ad/4-1BBL/IL-12 + DCs), determined using the hierarchical scheme detailed in *Computational Methods*, are found in Table S2 in *SI Results*. For reference, Table S2 also compares the best-fit parameters found herein to those previously found using a different algorithm and a slightly different metric of fit (35). Finally, Table S2 includes the 95% credible interval for each fit parameter, as calculated from the approximated posterior distributions (Fig. S1). These 95% credible intervals are compared to the results of a local sensitivity analysis, as shown in Figs. S2 and S3 of *SI Results*.

Robustness at Intermediate OV and DC Dose. Here we focus on identifying and classifying the response and robustness to 20 treatment protocols (all possible combinations of Ad/4-1BBL/IL-12 with DCs given at three doses a piece, separated by 1 d) at the experimentally used dose of 2.5×10^9 OVs per dose and 10^6 DCs per dose (31); we will refer to this as the “intermediate OV/DC dose” going forward. Our previous analysis revealed extreme sensitivity to parameters at this dosage (35), so here we expand upon that observation and undertake an exploration of treatment robustness using the VEPART approach.

To this end, we generate 1,000 virtual populations by pseudo-randomly sampling the posterior distributions on the fit parameters (Fig. S1), as described in *Computational Methods*. These virtual populations are alternative parameterizations to the model in Eqs. 1–6 that, when pooled together, in theory capture the underlying heterogeneity in the full patient population (16). The 20 treatment orderings considered in ref. 35 were administered to each of these virtual populations, allowing us to determine a “population-level” response to drug ordering at the intermediate OV/DC dose (Fig. 3).

Fig. 3A demonstrates the extent to which the virtual populations respond differently to the same treatment protocol. For a fixed ordering of the drugs (specified on the x axis), we see the frequency with which each protocol ranks in positions 1 through 20, with 1 being the optimal treatment ordering and 20 being the worst. Little consistency is observed in the ranking of the different treatment protocols across virtual populations. For instance, 65% of the treatment protocols rank in both the top two and bottom two positions in some of the virtual populations with nonzero probability (Fig. 3A). That number grows to 90% if we consider those that can rank in both the top and bottom five positions (Fig. 3A).

Another indication of the nonrobust (fragile) response across virtual populations at this dose is that, on average, each of the 20 treatment protocols achieve over 17 different rankings. As an example, the previously found optimal protocol of OV–OV–OV–DC–DC–DC achieves every rank between 1 and 20, with the exception of rank 13 (Fig. 3A). This protocol is most likely to rank in the top position, as it ranks in position 1 for 72.2% of the virtual populations (Fig. 3A). However, the next most likely ranking for this protocol is in the last position, as it ranks in position 20

Fig. 3. VEPART output at intermediate OV and DC doses. The x axis indicates the treatment protocol, with “V” representing Ad/4-1BBL/IL-12 treatment and “D” representing DC treatment on a given day. (A) For each of the 20 treatment protocols, we see the frequency at which it ranks in positions 1 to 20. Observe that there is little consistency across the virtual populations: On average, each of the treatment protocols can achieve over 17 different ranks. (B) The frequency of virtual populations for which the specified treatment protocol leads to tumor eradication (blue) or tumor escape (yellow).

for 13.8% of the virtual populations (Fig. 3A). The fact that the same protocol is the best case for some virtual populations and yet is the worst case for others strongly suggests the originally identified optimal protocol is not robust.

Because there was minimal support for a given treatment protocol to achieve a consistent ranking across the virtual populations, we then sought to address a simpler question: Do certain protocols result in tumor eradication in a large fraction of the virtual populations, independent of where they appear on the rank-ordered list of schedules? Consistent with the lack of a robust response at this dose, we found no protocols that lead to eradication in a majority of the virtual populations. Instead, we observed that the treatment that ranked as the top protocol most frequently (OV–OV–OV–DC–DC–DC) only results in tumor eradication in 30% of the virtual populations (Fig. 3B). All other protocols have a lower likelihood of tumor eradication, lending further evidence to the claim that the tumor eradication response in this dosing regime is fragile.

Digging more deeply into the binary classification of treatment protocols, we found that treatment response is more closely tied to the virtual population under consideration than to the treatment protocol itself (order in which the doses are given). By sorting our data by virtual population (instead of treatment protocol, as done in Fig. 3), we find that just over 76% of the virtual populations have the same binary response (tumor eradication or tumor escape) to all 20 treatment protocols tested. In particular, our simulations reveal that tumor escape will occur independent of the treatment protocol used in just over 66% of the virtual populations. Further, tumor eradication will occur independent of the treatment protocol in 10% of the virtual populations. Because the majority of virtual populations have a binary response (eradication or escape) independent of the dosing order, we learn that the parameters in the virtual population, and not the treatment order itself, are really driving the predictions in the intermediate OV/DC region of dosing space.

To conclude, in this region of dosing space treatment response can vary significantly across virtual populations. None of the treatment protocols, including the optimal one of front loading the OVs, robustly result in tumor eradication across the virtual populations. The situation only gets messier when we consider

the frequency with which each protocol achieves different rankings (from top protocol to worst protocol). Finally, the observation that treatment response is driven by the parameters that describe a virtual population, rather than the dosing order itself, further suggests that optimality predictions at this dosage may be of limited value. Because the population-level eradication response is fragile, we conclude that the combination of giving three doses of 2.5×10^9 Ad/4-1BBL/IL-12 and three doses of 10^6 DCs is not the ideal drug dosage to administer in this experimental system.

Robustness at Higher OV and Lower DC Dose. Because the intermediate OV/DC dose was found to be fragile, we next sought to analyze the robustness of the 20 protocols at a 50% higher OV dose (3.75×10^9 versus 2.5×10^9 OVs per dose) and a 50% lower DC dose (0.5×10^6 versus 10^6 DCs per dose). Like before, robustness of a protocol is assessed by measuring treatment efficacy across all virtual populations generated by pseudorandomly sampling the approximated posterior distributions for the fit parameters, as detailed in *Computational Methods*.

Similar to what was observed in the intermediate OV/DC dose, no treatment protocols result in a robust eradication response across the virtual populations (Fig. 4A). Although the protocol of front loading OVs consistently ranks as the top treatment across all virtual populations (Fig. 4B), the likelihood of tumor eradication using this protocol is only 43%. This outperforms the same protocol (which was also optimal) in the intermediate OV/DC dosing regime by 13%, but it still leaves over half of the virtual populations unsuccessfully treated. Further, all other protocols have a lower likelihood of eradication (with OV–OV–DC–OV–DC–DC being the only other one to cross the 33% threshold), suggesting that the tumor eradication response in this dosing regime is fragile.

That said, some responses in this dosing regime are robust, just not when it comes to the desired outcome of tumor eradication. Sixty percent of the protocols result in tumor escape in more than 90% of the virtual populations, and front loading the DCs results in treatment failure in 100% of the virtual populations (Fig. 4A). Further, unlike in the intermediate OV/DC regime, there is little variation in the rank each treatment protocol can achieve

Fig. 4. VEPART output at high OV/low DC doses. The x axis indicates the treatment protocol. (A) The frequency of virtual populations for which the specified treatment protocol leads to tumor eradication (blue) or tumor escape (yellow). (B) For each of the 20 treatment protocols, we see the frequency at which it ranks in positions 1 to 20. Observe that there is strong consistency across the virtual populations, with each protocol achieving a dominant ranking and other rankings only near that dominant one.

Discussion

In this work, we introduced the VEPART platform for assessing the robustness of treatment protocols. VEPART implementation begins with an experimental dataset describing the temporal evolution of some variable (for instance, tumor volume) in response to treatment in a sample population. A mathematical model (for instance, a system of differential equations) is then developed and fit to the experimental data. Once a reasonable mathematical model is attained, nonparametric statistical techniques are used to amplify the sample population and generate a large number of virtual populations. One benefit of this approach is that a large sample size, which can be difficult to attain experimentally, is not required to create these virtual populations. At this point, treatment optimization can be performed for each virtual population.

Although optimization is restricted to a set of clinically realizable protocols in this work, once the virtual populations have been created, classical tools from optimal control theory could also be applied (36, 37). An excellent overview of applying optimal control theory to cancer chemotherapy planning can be found in ref. 38. The use of optimal control in the context of the VEPART method would allow a more general optimal protocol to be identified, where “optimal” could be defined in many ways; for instance, one could seek to minimize tumor volume at an endpoint, minimize tumor volume over a time horizon, minimize drug concentration, minimize toxicity, minimize some weighted average the above, etc. (37–45). Further, this optimization can be performed subject to various types of constraints; for instance, toxicity could be introduced as a constraint on the amount of drug that can be administered (37, 46). These approaches can be used to find the optimal schedule for a single therapeutic agent, or for a mixture of drugs as done in refs. 43 and 47. Further, from any such optimal control problem, one could perform a sensitivity or elasticity analysis (similar to the one performed herein; see *SI Results*) to quantify how parametric changes influence the quality of an optimal treatment protocol, as in ref. 48.

A variety of solution methods exist for solving these optimal control problems (38). Analytical expressions for the optimal control can be obtained for simple models (see, e.g., ref. 49), and, when the model is too complex to obtain an analytic expression, approximation techniques can be used (see, for instance, ref. 43). When the optimal solution is computationally intractable, heuristic optimization algorithms, including genetic algorithm and simulated annealing can be used, as done in refs. 42, 47, and 50. Independent of how the optimization is carried out, an analysis of the optimal protocols across virtual populations using the VEPART method allows for the quantification of protocol robustness.

Related Work: Robust Cancer Therapies. In the cancer literature, examples of analogous efforts for designing robust treatment protocols can be found in radiotherapy (51–54). The challenges in designing a robust radiation treatment protocol can largely be attributed to geometric uncertainty and interpatient variability (54). Several studies have addressed the challenges posed by geometric uncertainty, including factors such as organ motion, variations in treatment setup, patient positioning errors, and fluctuations in machine output (54). For instance, Liu et al. (52) used “worst-case robust optimization” to identify intensity modulated proton therapy (IMPT) plans that are robust to uncertainties in the beamlet range and equipment setup. For the same beam arrangement, they computed a different dose distribution accounting for these uncertainties and compared this to the optimal dose distribution. They applied this approach to the lung, skull base and prostate and found that compared with IMPT plans optimized using conventional (nonrobust) methods, their method resulted in radiotherapy plans that are less sensitive to beamlet range and setup uncertainties (52). In a series of papers, robust optimization was used for planning intensity modulated radiation therapy for lung cancer in the face of uncertainty caused by breathing (51, 55, 56). This work introduces a data-driven model

of uncertainty to describe variations in breathing motion and allows the authors to determine the trade-off between ensuring the tumor receives sufficient radiation and minimizing the dose to normal tissue (51, 55, 56).

On the other hand, the work by Leder and colleagues (54) was the first to consider the challenge that interpatient variability poses to designing a robust radiotherapeutic protocol. They were particularly concerned about the potential risk that the optimal protocol (determined using the linear–quadratic model) may be weakly efficacious for some individuals, or may cause high toxicity to at-risk organs for others. To address this robustness concern, the authors used a stochastic optimization scheme in which the unknown parameters are assumed to be random variables with known distributions. The goal is to choose a dosing schedule (total dose, number of fractions, dose per fraction, and treatment duration) for which the objective function attains a high level with a given probability, and that the imposed constraints are also satisfied with a given probability. The authors found some important differences in the optimal schedule that accounts for parameter uncertainty compared with the optimal schedule found using the nominal parameter values in the absence of uncertainty (54). Our work is motivated by a similar robustness question to that considered by Leder and colleagues in ref. 54, although not restricted to radiotherapy.

Case Study: Immunoenhanced OV's with DC Vaccines. As a proof of concept, we applied VEPART to explore the robustness of treatment protocols that combine three doses of immunostimulatory OV's with three DC injections; to accomplish this, we worked with the experimental data from Huang et al. (31). The nature of this dataset previously led us to use a hierarchical method for fitting the parameter values (*Computational Methods*), which posed a challenge for directly fitting parameters for each of the virtual populations. To overcome this challenge, we began by bootstrapping the experimental data and using the best-fit parameters in each bootstrap replicate to approximate the posterior probability distribution for the parameters in our mathematical model (Eqs. 1–6). Pseudorandom sampling of these distributions, as detailed in *Computational Methods*, allowed for the creation of our 1,000 virtual populations.

In this case study, 20 treatment protocols (all possible combinations of Ad/4-1BBL/IL-12 with DCs given at three doses a piece, separated by 1 d) were ranked across the virtual populations in three regions of dosing space: (i) intermediate and experimentally used OV/DC dose, (ii) high OV/low DC, and (iii) high DC/low OV. VEPART revealed that the first two dosing regimes we considered are fragile. To detail, the protocol of administering Ad/4-1BBL/IL-12 on the first 3 d, and following up with a sequence of DCs, was optimal for both the intermediate dose of OV's and DCs and the high OV/low DC dose. However, in neither case did this protocol cause a robust eradication response: The protocol only causes tumor eradication in 30% of the virtual populations at the intermediate dose, and in 43% of the virtual populations at the high OV/low DC dose. As a consequence of VEPART's nonrobust prediction, it is not expected that any ordering of three OV's and three DCs at these two doses will exhibit a strong antitumor response in the face of interpatient and inpatient heterogeneity.

On the other hand, in the high DC/low OV region of dosing space, a robust optimal protocol was identified. This protocol administers DCs on the first 3 d, and follows up with a sequence of Ad/4-1BBL/IL-12 on the last 3 d. Not only did this protocol rank as optimal in all virtual populations, it also resulted in tumor eradication in 84% of those virtual populations. Therefore, VEPART predicts that, in the high DC/low OV region of dosing space, the treatment that front-loads DCs will be effective across different individuals in a population, and will be more robust to spatial and temporal variations within an individual.

In future work, the use of approximate optimal control techniques or heuristic optimization algorithms (38, 42) can extend the current implementation of VEPART. In particular, we can optimize over a wider range of protocols (for instance, considering

a variable number of OV and DC doses, and/or allowing variable spacing between doses), and over a more complete range of dosing space. This extended analysis of our system could result in finding robust protocols beyond treating with DC–DC–DC–OV–OV–OV in the high DC/low OV region of dosing space.

This case study is intended to illustrate how the VEPART method can be used to assess therapeutic robustness. By combining experimental data with mathematical modeling and statistical analyses, the VEPART approach can contribute to tackling a significant problem clinicians face when treating cancer using standard of care or an individualized treatment protocol: high levels of interpatient and inpatient variability.

Computational Methods

In this section, we introduce our mathematical model that includes immunostimulated OVs and DC vaccines. Although this model has been successfully fit to the experimental data of Yun and colleagues (31, 35), we introduce an alternative approach to fitting the data that results in small but significant improvements in the model fits. To further study the relationship between the model and its parameters, in this section, we also describe two key steps of our proposed VEPART approach: (i) We detail the bootstrapping procedure that produces posterior distributions and credible intervals for each of the fit parameters, and (ii) we explain how these posterior distributions are used to generate virtual populations that provide the foundation of our robustness analysis.

Mathematical Model. In this study, the data-validated model from Wares et al. (35) was used. The model is represented by the following initial value problem:

$$\frac{dU}{dt} = rU - \beta \frac{UV}{N} - (k_0 + c_{kill}I) \frac{UT}{N} \quad U(0) = U_0, \quad [1]$$

$$\frac{dI}{dt} = \beta \frac{UV}{N} - \delta_I I - (k_0 + c_{kill}I) \frac{IT}{N} \quad I(0) = 0, \quad [2]$$

$$\frac{dV}{dt} = u_v(t) + \alpha \delta_I I - \delta_V V \quad V(0) = 0, \quad [3]$$

$$\frac{dT}{dt} = c_T I + \chi_A A + \chi_D D - \delta_T T \quad T(0) = 0, \quad [4]$$

$$\frac{dA}{dt} = c_A I - \delta_A A \quad A(0) = 0, \quad [5]$$

$$\frac{dD}{dt} = u_D(t) - \delta_D D \quad D(0) = 0, \quad [6]$$

where U is the number of uninfected tumor cells (the only variable with a nonzero initial condition that gets fit at each step of the hierarchical fitting process), I is the number of tumor cells infected by the OV, V is the number of free virions (oncolytic adenoviruses), T is the number of tumor-targeted T cells, A is the number of naïve T cells, D is the number of injected DCs (note that endogenous DCs are not directly modeled), and N is the total number of cells (tumor cells and T cells) at the tumor site. The individual terms that contribute to the rate of change of each population are explained in more detail in [SI Computational Methods](#).

This model was hierarchically developed to fit the increasingly complex data in Fig. 2. Although more details are provided in [SI Computational Methods](#), here we briefly note the different hierarchical stages of the model:

- i) Model 0 is a model of untreated tumor growth in the absence of any virions, T cells, or DCs.
- ii) Model 1 is a model of tumor growth and treatment with oncolytic virotherapy that does not carry any immunostimulatory transgenes. T cells and DCs are still not considered.
- iii) Model 2a is a model of tumor growth and treatment with the immunostimulatory oncolytic virotherapeutic Ad/4-1BBL. The immunostimulation results in the inclusion of tumor-

targeted T cells, but naïve T cells and DCs are still not considered.

- iv) Model 2b is a model of tumor growth and treatment with the immunostimulatory oncolytic virotherapeutic Ad/IL-12. The immunostimulation results in the inclusion of tumor-targeted T cells, including the naïve T cells, but DCs are still not considered.
- v) Model 3 is a model of tumor growth and treatment with the immunostimulatory oncolytic virotherapeutic Ad/4-1BBL/IL-12. The immunostimulation results in the inclusion of tumor-targeted T cells, including the naïve T cells, but DCs are still not considered.
- vi) Model 4 is a model of tumor growth and treatment with Ad/4-1BBL/IL-12 and DC vaccines. At this point, the model includes all equations and all terms within each equation, including the tumor-targeted T cells, the naïve T cells, and the DCs. A subcase of model 4 in which all virus-related parameters are set to zero gives us the case of treating with only the DC vaccine.

Fitting Model to Experimental Data. Just as the structure of the model was hierarchically developed, the parameter fitting scheme was also hierarchical in nature (34, 35). As illustrated in [Fig. S4](#), parameters that could not be well-estimated from the literature were fit at each hierarchical model stage, and some subset of the fit parameters are inherited to the subsequent models in the hierarchical scheme. Details of this hierarchical fitting scheme can be found in [SI Computational Methods](#). The initial condition for the number of uninfected tumor cells was always refit at each model stage, as each model was fit to a different data set. All other initial conditions are identically zero, as the administration of treatment is captured through the time-varying terms [$u_v(t)$ and $u_D(t)$] in Eqs. 1–6.

The parameters in models 1 through 4 were previously fit using the Levenberg–Marquadt algorithm as implemented by MATLAB's `lsqnonlin` command (35). Like most parameter fitting schemes, the Levenberg–Marquadt algorithm can get stuck at local minimum, and the algorithm's performance deteriorates as a function of the dimension of parameter space. To have more control over parameter fitting, and to attempt to avoid getting trapped at local minimum, here we instead use simulated annealing for model fitting.

Goodness of model fit to the experimental data is sought by minimizing the objective function S , as defined by

$$S = \sum_t v_t \frac{(v_t^{model} - v_t)^2}{\sigma_t^2}, \quad [7]$$

where v_t is the average experimental tumor volume at day t , v_t^{model} is the tumor volume at day t predicted by Eqs. 1–6, and σ_t^2 is the variance in the experimental tumor volume at day t . Within Eq. 7, the fractional term is a dimensionless measure of the error: The numerator gives the sum of the square errors between the model predictions and the (average) experimental data, and the denominator is the variance in the experimental data. By dividing the sum of the square errors by the variance, we impose the condition that, at time points where the variance is small, we want a better fit to the average volume than is required at time points where the variance is large, in accordance with the principle of maximum likelihood estimation (57).

Because volume measurements made using calipers are imprecise for smaller tumor sizes [instrumentation error in measuring length and width is independent of tumor volume (58, 59)], the dimensionless term in Eq. 7 is weighted by the average tumor volume at day t . Our weighting has the additional advantage of leading to a better-posed numerical optimization problem, as this objective function does not artificially bias the algorithm to fit well at small tumor sizes (when the variance is very small) at the expense of fitting well over a majority of the data points. Details on the simulated annealing algorithm being used to minimize Eq. 7 can be found in [SI Computational Methods](#) and [Table S3](#) therein.

Analyzing Best-Fit Parameters. For each dataset (control, Ad only, etc.) consisting of N mice (N was between six and nine, depending on the dataset), 1,000 bootstrap replicates have been created. Each bootstrap replicate was created by sampling N mice from the original dataset with replacement (36) using the programming language R. The simulated annealing protocol was run on each bootstrap replicate, and the best-fit parameter values for each bootstrap replicate were binned in a histogram. This process provides a visualization of the estimated posterior marginal distribution on each fit parameter. Given the distributions were empirical and asymmetric, we defined the 95% credible interval for that parameter (the interval for which we can be 95% confident that the true value of the parameter occurs in) as the range that excluded 2.5% of the values on each end of the distribution. Bootstrapping has been used by others to explore parameter sensitivity in mathematical biology models with implications for cancer; see, for instance, refs. 60–62.

A local sensitivity analysis was also performed to give an alternative mechanism for analyzing the fit parameters. Details on this are found in *SI Computational Methods*.

Ranking and Robustness of Treatment Protocols. Previous work determined the optimal protocol among the 20 possible orderings of three OV injections and three DC injections at the experimentally used dose of 2.5×10^9 OVs and 10^6 DCs (35). To rank these 6-d-long protocols, tumor growth was tracked over 1 mo, and the volume predicted by the model after this period was recorded. That analysis revealed that the only ordering of the drugs that led to tumor eradication (defined as tumor volume dropping below the assumed volume of one cell, 10^{-6} mm³) was the one that front-loads the OV (that is, OV–OV–OV–DC–DC–DC) (35). However, further analysis revealed that the dose used in the experimental work was near a bifurcation point in the system, and small changes in the drug dose could result in a different optimal solution (35).

This finding motivated us to extend our analysis to investigate treatment robustness, as such an analysis gives more information about therapeutic efficacy than a single-population optimization study. To explore robustness, the data obtained from the bootstrap replicates were used to construct virtual populations, similar to the nonparametric approach described in ref. 29. In particular, herein, 1,000 virtual populations were created by pseudorandomly picking the six fit parameters (r , β , c_A , c_T , c_{kill} , and χ_D) from their posterior distributions approximated using the bootstrap replicates and the hierarchical fitting process. Two constraints were imposed when creating these virtual populations. First, to maintain the covariance structure between the parameters, those that were fit together were selected together to create a new virtual population. This criterion impacted the selection of parameters from model 3, and we implemented this parameter selection by randomly choosing a value of c_A from its approximated posterior distribution, and subsequently setting the values for c_T and c_{kill} to the best-fit value in the same bootstrap replicate that gave that value of c_A .

Note that an implicit assumption in creating virtual populations in this way is that parameters fit at different stages of the hierarchical fitting process are minimally correlated. This assumption is grounded in our fitting methodology, in which parameters fit at each step of the hierarchy are treated as independent biological phenomenon. For instance, the fitting of the tumor growth rate r at the first step of the hierarchy isolates the behavior of the tumor

without treatment, and the fitting of the viral infectivity parameter β at the next step of the hierarchy is meant to isolate the behavior of the OV. In effect, this setup establishes a regime of tumor growth and viral activity that the system responds to. Therefore, by independently sampling from the distribution for r and β to create a virtual population, we are trying to select a reasonable tumor growth rate and a reasonable viral infectivity rate, as seen in the experimental data. Others have given consideration to further preserve the covariance structure among all variables; see, for instance, refs. 16, 23, 25, 26, 29, and 30. The second constraint imposed is that only virtual populations whose parameter values are all within their respective 95% credible intervals were considered; this can be thought of as an “inclusion–exclusion” criterion that refines the virtual population pool to be statistically similar to the experimental population (29), including mirroring its heterogeneity. That said, this approach does constrain the virtual populations to statistically resemble the experimental data, which, because of the small sample size in our experiments, runs the risk of not resembling the population data (29). However, the experimental data considered herein inject genetically identical mice with the same number of cells from the same cancer cell line. Therefore, at least for these data, it is reasonable that the small sample can be expanded to result in virtual populations that accurately reflect the true heterogeneity in the population.

Therapeutic robustness was assessed by ranking the 20 protocols of interest for each of the virtual populations that were generated. Within a virtual population, a treatment was ranked using a two-component criterion. First, for tumors that were eradicated within 30 d (volume drops below 10^{-6} mm³), the time until eradication is used to measure the effectiveness of the protocol, with faster eradication times considered superior to slower ones. Second, for tumors that were not eradicated within 30 d, the volume after 30 d was used to measure the effectiveness of the treatment, with smaller volumes considered superior to larger ones. By analyzing the response to the 20 protocols across the 1,000 virtual populations, the robustness of each protocol can be studied.

Given that our previous work suggested that the experimentally used dose does not have a robust optimal solution, we performed this robustness study in three different regions of dosing space: (i) at the experimentally used dose (OV dose of 2.5×10^9 , DC dose of 10^6), (ii) at a 50% higher dose of OV but a 50% lower dose of DC (OV dose of 3.75×10^9 , DC dose of 0.5×10^6), and (iii) at a 50% lower dose of OV but a 50% higher dose of DC (OV dose of 1.25×10^9 , DC dose of 1.5×10^6). These two other regions of dosing space were chosen because they are sufficiently distinct from the experimental dose such that we could expect different optimal protocols and a different robustness profile. Further, both of these regions of dosing space involve increasing the dose of one drug and decreasing the dose of the other by the same relative amount; this was done in an attempt to preserve the toxicity profile across the three dosing regimes.

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