Published in final edited form as:

Biochimie. 2017 April ; 135: 1–5. doi:10.1016/j.biochi.2017.01.001.

Medium Throughput Biochemical Compound Screening Identifies Novel Agents for Pharmacotherapy of Neurofibromatosis Type I

Galina Semenova^{1,3,5}, Dina Stepanova^{2,3}, Sergey M. Deyev^{1,4}, and Jonathan Chernoff³

¹Shemyakin–Ovchinnikov Institute of Bioorganic Chemistry, Russian Academy of Sciences, Moscow, Russia

²Russian National Research Medical University, Moscow, Russia

³Cancer Biology Program, Fox Chase Cancer Center, Philadelphia, PA

⁴Tomsk State University, Tomsk, Russia

Abstract

The variable manifestation of phenotypes that occur in patients with neurofibromatosis type 1 (NF1) includes benign and malignant neurocutaneous tumors for which no adequate treatment exists. Cell-based screening of known bioactive compounds library identified the protein phosphatase 2A (PP2A) inhibitor Cantharidin and the L-type calcium channel blocker Nifedipine as potential candidates for NF1 pharmacotherapy. Validation of screening results using human NF1-associated malignant peripheral nerve sheath tumor (MPNST) cells showed that Cantharidin effectively impeded MPNST cell growth, while Nifedipine treatment significantly decreased local tumor growth in an MPNST xenograft animal model. These data suggest that inhibitors of PP2A, as well as calcium channel blockers, might be used in broader MPNST preclinical studies as single agents or in combinatorial therapeutic strategies.

Keywords

neurofibromatosis type 1; chemical screening; Cantharidin; Nifedipine

1. Introduction

Neurofibromatosis type 1 (NF1), also known as *von Recklinghausen's disease*, is a frequent human cancer predisposition syndrome affecting many organ systems, peripheral nervous system in particular (1–3). Typical NF1 symptoms include neuroectodermal tumors and lesions of other tissue origins, pigmentation patterns, as well as skeletal abnormalities and

⁵Correspondence to: Galina Semenova, Cancer Biology Program, Fox Chase Cancer Center, 333 Cottman Ave W422, Philadelphia, PA 19111, 215 728 5320, Galina.Semenova@fccc.edu.

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

cognitive deficits (3). Most NF1 patients develop dermal and plexiform neurofibromas - benign lesions associated with peripheral nerves. Dermal neurofibromas are tumors in the skin, while plexiform neurofibromas (PNF) develop along the nerve plexus. PNFs can reach large sizes and compress nearby nerves, causing pain and a range of dysfunctions; moreover, PNF can transform into malignant peripheral nerve sheath tumors (MPNSTs) (4). These cancers exhibit highly aggressive metastatic growth and serve as major source of morbidity for NF1 patients (4, 5). To date there is no established pharmaceutical treatment for benign and malignant NF1-associated tumors, thus, the search for novel therapeutic targets for this disease is of intense scientific and clinical interest (6).

Neurofibromatosis type 1 is a monogenic dominantly inherited autosomal disorder. It occurs due to inactivating mutations in the *NF1* tumor suppressor gene that encodes the Ras-regulatory protein neurofibromin (6, 7). Neurofibromin is broadly expressed and plays important roles in regulating multiple cellular processes, as evidenced by the variety of symptoms arising from its loss (3, 6). However, specific functions of neurofibromin remain unclear. Although many neurofibromin interactions with other proteins have been reported in addition to controlling Ras (3), the biological meaning of these additional interactions is largely undefined.

Genetic and biochemical screenings can be useful tools for defining new protein functions as well as therapeutic targets. Here we perform cell-based phenotypic screening assay to identify novel compounds that inhibit proliferation and survival of cells lacking neurofibromin. We show that small molecules of two distinct classes – the protein phosphatase 2 (PP2A) inhibitor Cantharidin and the calcium antagonist Nifedipine - exhibit selective toxicity towards NF1-deficient mouse embryonic fibroblasts (MEFs). We demonstrate that Cantharidin effectively inhibits growth of human NF1-associated MPNST cells, suggesting that PP2A might represent an attractive new target for MPNST pharmacotherapy. Finally, we show that Nifedipine inhibits cell growth as well as xenograft tumor growth of an aggressive MPNST cell line, S462TY. These results suggest that unbiased chemical screens may yield unexpected candidates for NF1 pharmacotherapy.

2. Materials and Methods

2.1. Cell lines

Immortalized murine *Nf1*-negative and wild type mouse embryonic fibroblasts (MEF *Nf1*-/- – *E1A-p53* and MEF *Nf1*^{wt} *E1A-p53*) were generously provided by Dr. Karen Cichowski (Brigham and Women's Hospital and Harvard Medical School, Boston, USA). MPNST cell lines sNF96.2, sNF02.2 and sNF94.3 were obtained from the American Type Culture Collection, S462TY cells were generously provided by Dr. Timothy Cripe (Nationwide Children's Hospital, OH), ST8814, ST88-3 and 90-8 were generously provided by Dr. Nancy Ratner (Cincinnati Children's Hospital, OH). Cells were maintained in Dulbecco's Modification of EAGLE Medium (DMEM) supplemented with 10% fetal bovine serum and penicillin/streptomycin and cultured on tissue culture-treated plates under standard conditions of 37°C and 5% CO₂.

Cells were maintained in Dulbecco's modified Eagle medium (DMEM) supplemented with 10% FBS, 2 mM L-glutamine, 1 mM sodium pyruvate and 1% penicillin/streptomycin (all Invitrogen, USA) under standard conditions of 37°C and 5% CO₂.

2.2. In Vitro Cytotoxicity Assay, Half Inhibitory Concentration (IC₅₀) Calculation

MEFs were plated in triplicate for each dose of the compound on 96-well plates at 10000 cells/well in complete medium. Twenty-four hours after plating, cells were treated with carrier alone (0.1% DMSO) or bioactive compounds. Cell viability was evaluated seventy-two hours after addition of drugs using alamarBlue fluorescent assay (Life Technologies, USA). The values of IC₅₀, the drug concentration required for 50% growth inhibition, were calculated using software package Calcsyn (Biosoft).

2.3. Small Bioactive Compound Library Screening

The ICCB Known Bioactives library (Enzo Life Sciences, USA, BML-2840-0100) was used for the screening. The screening was performed using MEF *Nf1*^{-/-} *E1A-p53* in triplicate and repeated 2 times. Drug transfer was performed 24 hrs after plating using CyBi Cell Vario automated pipettor, transfer volume 100 nL. DMSO and staurosporin at the final concentration of 5 uM were used as negative and positive controls respectively. Read-outs were done in 72 hours using alamarBlue cytotoxicity assay. Targets were counted positive if the viability ratio was < 0.85 and false discovery rate (FDR) was < 20%.

2.4. Xenograft studies

Female 6–7 weeks old *nu/nu* immunodeficient mice were injected subcutaneously with 10⁷ S462TY cells in 0.1 ml 30% Matrigel (BD Biosciences)/PBS. Treatment started when the average tumor size reached 100 mm³. Nifedipine (Santa Cruz) was prepared as a suspension in aqueous 0.5% hypromellose/0.2% Tween 80 and administrated by daily gavage at 50 mg/kg. Norcantharidin (Santa Cruz) was formulated in PBS and IP injected at 10 mg/kg/day. Tumors were measured with a caliper every tree days. Mice were sacrificed when tumor size reached 10% of body weight, tumors were isolated and weighted.

2.5. Statistics

Statistical significance for the IC₅₀ values was calculated by Holm-Sidak method with $\alpha=5\%$. For xenograft experiment statistical significance was estimated using the two-tail Student's *t*-test.

3. Results

We used MEF *Nf1*^{-/-} *E1A-p53* cells to assess the effects of 472 small molecules on the growth and survival of NF1-deficient neoplastic cells. *Nf1*^{-/-} *E1A-p53* MEFs were created by transforming MEFs harboring a germline bi-allelic inactivating *Nf1* mutation, with an E1A oncogene and dominant-negative p53 (8). Such transformation partially recapitulates p53 and Rb inactivation that are often seen in NF1-associated malignancies (9, 10).

Figure 1a shows preliminary screening results identifying 142 compounds that satisfied selection criteria (ratio between vital cells exposed compounds and negative control < 0.85

and FDR according to Benjamini—Hochberg method $< 20\%$). Compounds with poor target specificity and acute toxicity were excluded from the hit list. For further studies we selected candidate compounds from the drug classes most broadly represented in the “hit” list: Cerulenin, AA.861 (lipid biosynthesis agents), Calphostin C (PKC inhibitors), LY.294002 (PI3K inhibitors), Nifedipine, Nigericin (ion channel ligands), Cantharidin (PP2A inhibitors).

To access selective toxicity of indicated compounds toward NF1-deficient cells we exposed MEF *Nf1*^{−/−} *E1A-p53* cells and isogenic control MEF *Nf1*^{wt} *E1A-p53* cells to various doses of the compounds. The IC₅₀ values for both cell lines were determined for candidates identified by the screening. Of seven initially selected compounds, only two: Cantharidin and Nifedipine showed consistent significant differences in suppression of NF1-negative versus control MEF cell growth (Figure 1b).

These results were then validated on a panel of seven NF1-associated human MPNST cell lines S462TY, ST88-3, ST8814, 90-8, sNF96.2, sNF02.2 and sNF94.3 (Figure 1c). Cantharidin IC₅₀ values for all MPNST cell lines tested were comparable to those obtained in MEF *Nf1*^{−/−} *E1A-p53* (Figure 1c). Interestingly, S462TY cells were extremely sensitive to the calcium channel blocker Nifedipine, however other MPNST cell lines exhibited little sensitivity to this compound (Figure 1c).

We next asked whether our *in vitro* observations might be recapitulated *in vivo*. The S462TY MPNST cell line was used to establish subcutaneous xenograft tumors and therapy was initiated when average tumor size reached 100 mm³. For these experiments we used Nifedipine at 50 mg/kg/day and Norcantharidin (NCTD) (demethylated form of cantharidin with reduced toxicity (11)), at 10 mg/kg/day. Mice in NCTD treated group were sacrificed at day 12 when two of six xenograft tumors reached 10% body weight. The experiment was stopped at the day 15 when most control mice required sacrifice, and the tumors were collected. Exposure to Nifedipine significantly attenuated S462TY xenograft growth as indicated by tumor volume (~492.06 mm³ versus ~1638.13 mm³ in control group at study termination, $p = 0.029$) (Figure 2a) and tumor weight (~0.256 g versus ~1.191 g in control group, $p = 0.020$) (Figure 2b) and was well tolerated.

4. Discussion

Current therapeutic options for benign and malignant NF1-associated tumors are restricted to surgical resection, a treatment that does not significantly improve prognosis (6). In this study we aimed to identify additional, and perhaps more effective, treatment options for NF1 by identifying novel pharmaceutical agents that suppress tumor growth without toxic side effects. We took advantage of a pair of isogenic MEFs (wild type and NF1-deficient), as well as human NF1-associated MPNST cells, and screened known bioactive compounds to identify agents that selectively inhibit growth of NF1-deficient cells.

The first compound selected from the screen, Cantharidin, is a potent inhibitor of protein phosphatase 2A (PP2A), an enzyme that plays an important role in cell proliferation, survival, and cell-fate determination (12). PP2A targets many oncogenic signaling cascades

and was initially characterized as a tumor suppressor (12), however, accumulating evidence points to its anti-apoptotic role in multiple cancer cell types (12), as well as a contribution to chemotherapy resistance mechanisms (13). Pharmaceutical inhibitors of PP2A compromise cancer cell proliferation, survival and invasion (12, 14, 15) and are considered promising for treatment of particular types of cancer (12, 13). In our study, NF1-deficient MEFs as well as all NF1-associated MPNST cells were significantly more sensitive to Cantharidin compared to wild type MEFs, suggesting that pre-malignant and malignant NF1-associated tumor cells biology depends on PP2A activity. As selective PP2A inhibitors, including Cantharidin analogs (11, 16, 17), have been developed, such small molecules might represent promising therapeutic agents for neurofibromas and MPNSTs alone or in combination with chemotherapeutic agents.

Another compound identified in our screen, Nifedipine, is a dihydropyridine calcium antagonist (Figure 1b). Nifedipine disrupts extracellular calcium influx through voltage-dependant calcium channels and is broadly used as antihypertensive drug (18). Given the importance of calcium entry for cell division (19) potential anticancer effects of calcium antagonists are being discussed (19–21). Although Nifedipine showed moderate selective toxicity toward NF1-deficient cells in our MEF-based system, the majority of NF1-associated MPNST cells tested, with the notable exception of S462TY cells, exhibited little to no response to the compound. It is not known whether MPNST cells express functioning voltage-dependent calcium channels and whether alternative mechanisms of calcium signaling system are involved in regulation of MPNST cell growth. Thus these data suggest that the observed effects is cell-type specific and that Nifedipine might be a perspective candidate only for selective MPNSTs, such as that represented by the S462TY cell line.

Out of seven MPNST cell lines used, S462TY, exhibited the most aggressive growth and was the only cell line that gave rise to xenograft tumors after subcutaneous injection. Unlike other cell types, S462TY showed high sensitivity to Nifedipine, but was the least sensitive to Cantharidin *in vitro*. Similar results were obtained in a xenograft experiment. Strikingly, exposure to Nifedipine over a period of two weeks significantly impeded S462TY tumor growth, while treatment with the PP2A inhibitor Norcantharidin (NCTD) showed no tumor suppressive effect (Figure 2). These observations point to substantial heterogeneity amongst human MPNSTs in terms of expression of drug targets, response to inhibitors, and mechanism of drug resistance.

In conclusion, we show that neurofibromin deficiency sensitizes cells to the protein phosphatase 2A inhibitor Cantharidin and the calcium antagonist Nifedipine, addressing potential new roles of neurofibromin in calcium signaling and specific cascades targeted by PP2A. Potent inhibition of NF1-associated MPNST cell viability by Cantharidin suggests that PP2A should be considered as a potential therapeutic target for these types of tumors. Further investigations shall focus on mechanisms underlying sensitivity of various NF1-deficient cell lines to Nifedipine. That these unexpected targets emerged from an unbiased screen for small molecules that inhibit the growth of NF1-deficient cells suggests that similar approaches will have value in uncovered new therapeutic targets in NF1-associated tumors.

Acknowledgments

We thank Dr. Karen Cichowski for providing immortalized MEF cell lines, Dr. Timothy Cripe and Dr. Nancy Ratner for providing MPNST cell lines. This work was supported by grants from the NIH (R01 CA148805 and R01 CA098830), and the Children's Tumor Foundation to Jonathan Chernoff, NIH CORE Grant CA06927, RSF grant 14-24-00106 and an appropriation from the state of Pennsylvania to the Fox Chase Cancer Center.

References

1. Lakkis MM, Tennekoon GI. Neurofibromatosis type 1. I. General overview. *Journal of neuroscience research*. 2000; 62:755–763. [PubMed: 11107159]
2. Ferner RE. Neurofibromatosis 1. *European journal of human genetics: EJHG*. 2007; 15:131–138. [PubMed: 16957683]
3. Ratner N, Miller SJ. A RASopathy gene commonly mutated in cancer: the neurofibromatosis type 1 tumour suppressor. *Nature reviews Cancer*. 2015; 15:290–301. [PubMed: 25877329]
4. Evans DG, Baser ME, McGaughan J, Sharif S, Howard E, Moran A. Malignant peripheral nerve sheath tumours in neurofibromatosis 1. *Journal of medical genetics*. 2002; 39:311–314. [PubMed: 12011145]
5. Tonsgard JH. Clinical manifestations and management of neurofibromatosis type 1. *Semin Pediatr Neurol*. 2006; 13:2–7. [PubMed: 16818170]
6. Rad E, Tee AR. Neurofibromatosis type 1: Fundamental insights into cell signalling and cancer. *Seminars in cell & developmental biology*. 2016; 52:39–46. [PubMed: 26860753]
7. Trovo-Marqui AB, Tajara EH. Neurofibromin: a general outlook. *Clinical genetics*. 2006; 70:1–13. [PubMed: 16813595]
8. Johannessen CM, Reczek EE, James MF, Brems H, Legius E, Cichowski K. The NF1 tumor suppressor critically regulates TSC2 and mTOR. *Proceedings of the National Academy of Sciences of the United States of America*. 2005; 102:8573–8578. [PubMed: 15937108]
9. Carroll SL. The Challenge of Cancer Genomics in Rare Nervous System Neoplasms: Malignant Peripheral Nerve Sheath Tumors as a Paradigm for Cross-Species Comparative Oncogenomics. *The American journal of pathology*. 2016; 186:464–477. [PubMed: 26740486]
10. Zhu Y, Guignard F, Zhao D, Liu L, Burns DK, Mason RP, et al. Early inactivation of p53 tumor suppressor gene cooperating with NF1 loss induces malignant astrocytoma. *Cancer cell*. 2005; 8:119–130. [PubMed: 16098465]
11. Wang GS. Medical uses of mylabris in ancient China and recent studies. *Journal of ethnopharmacology*. 1989; 26:147–162. [PubMed: 2689797]
12. Kiely M, Kiely PA. PP2A: The Wolf in Sheep's Clothing? *Cancers*. 2015; 7:648–669. [PubMed: 25867001]
13. McDermott MS, Browne BC, Conlon NT, O'Brien NA, Slamon DJ, Henry M, et al. PP2A inhibition overcomes acquired resistance to HER2 targeted therapy. *Molecular cancer*. 2014; 13:157. [PubMed: 24958351]
14. Shen M, Wu MY, Chen LP, Zhi Q, Gong FR, Chen K, et al. Cantharidin represses invasion of pancreatic cancer cells through accelerated degradation of MMP2 mRNA. *Scientific reports*. 2015; 5:11836. [PubMed: 26135631]
15. Le AP, Zhang LL, Liu W, Shi YF. Cantharidin inhibits cell proliferation and induces apoptosis through G2/M phase cell cycle arrest in hepatocellular carcinoma stem cells. *Oncology reports*. 2016; 35:2970–2976. [PubMed: 26986084]
16. To KK, Ho YP, Au-Yeung SC. In vitro and in vivo suppression of growth of hepatocellular carcinoma cells by novel traditional Chinese medicine-platinum anti-cancer agents. *Anti-cancer drugs*. 2005; 16:825–835. [PubMed: 16096430]
17. Abe K, Niwa H, Iwase K, Takiguchi M, Mori M, Abe SI, et al. Endoderm-specific gene expression in embryonic stem cells differentiated to embryoid bodies. *Experimental cell research*. 1996; 229:27–34. [PubMed: 8940246]

18. Sorkin EM, Clissold SP, Brogden RN. Nifedipine. A review of its pharmacodynamic and pharmacokinetic properties, and therapeutic efficacy, in ischaemic heart disease, hypertension and related cardiovascular disorders. *Drugs*. 1985; 30:182–274. [PubMed: 2412780]
19. Capiod T. Cell proliferation, calcium influx and calcium channels. *Biochimie*. 2011; 93:2075–2079. [PubMed: 21802482]
20. Taylor JM, Simpson RU. Inhibition of cancer cell growth by calcium channel antagonists in the athymic mouse. *Cancer research*. 1992; 52:2413–2418. [PubMed: 1533173]
21. Loughlin KR. Calcium channel blockers and prostate cancer. *Urologic oncology*. 2014; 32:537–538. [PubMed: 24814406]

Highlights

1. Effective targeted therapies for Neurofibromatosis type 1 (NF1) associated tumors await discovery.
2. Chemical screening identified two compounds showing selective toxicity toward NF1-deficient mouse embryonic fibroblasts: the protein phosphatase 2A (PP2A) inhibitor Cantharidin and the calcium channel blocker Nifedipine.
3. Cantharidin significantly suppressed growth of human NF1-associated malignant peripheral nerve sheath tumor (MPNST) cells.
4. Nifedipine potently inhibited growth of subcutaneous MPNST xenografts

Figure 1.

Primary screening results and screen confirmation. A) ICCB small bioactive compound library of 472 known bioactive small molecules screened in *Nf1*-null mouse embryonic fibroblasts. Cell viability data is expressed as ratio to negative control. Out of 142 compounds, inhibiting cell viability (ratio < 0.85), 7 hits were chosen for further verification. B) Dose-response curves and IC₅₀ values of MEF *Nf1*^{-/-} *E1A-p53* cells and control MEF *Nf1*wt *E1A-p53* cells exposed to Cantharidin or Nifedipine, biochemical formulas of Cantharidin and Nifedipine. C) Half-inhibitory concentrations (IC₅₀s) for

Cantharidin and Nifedipine in human NF1-associated MPNST cell lines. Error bars represent standard deviation (**= $p<0.005$).

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

Figure 2. Effects of Norcantharidin (NCTD) and Nifedipine on the growth of subcutaneous S462TY xenografts. A) Volumetric changes of S462TY xenograft tumors in control, NCTD treated and Nifedipine treated groups (n=6). B) Xenograft tumors weights at study termination. Error bars represent standard error of the mean (*= $p < 0.05$).