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A Pause-then-Cancel model of stopping: evidence from basal ganglia neurophysiology

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Many studies have implicated the basal ganglia in the suppression of action impulses ('stopping'). Here, we discuss recent neurophysiological evidence that distinct hypothesized processes involved in action preparation and cancellation can be mapped onto distinct basal ganglia cell types and pathways. We examine how movement-related activity in the striatum is related to a 'Go' process and how going may be modulated by brief epochs of beta oscillations. We then describe how, rather than a unitary 'Stop' process, there appear to be separate, complementary 'Pause' and 'Cancel' mechanisms. We discuss the implications of these stopping subprocesses for the interpretation of the stop-signal reaction time—in particular, some activity that seems too slow to causally contribute to stopping when assuming a single Stop processes may actually be fast enough under a Pause-then-Cancel model. Finally, we suggest that combining complementary neural mechanisms that emphasize speed or accuracy respectively may serve more generally to optimize speed–accuracy trade-offs.

This article is part of the themed issue 'Movement suppression: brain mechanisms for stopping and stillness'.

1. Introduction

Inhibition of behaviours that are currently maladaptive is a key feature of normal executive function [1]. The various forms of behavioural inhibition include being more prepared to stop if required ('proactive' inhibition), and terminating movements that have already begun [2–4]. Here, we use the term 'stopping' to mean suppressing a current impulse to act, in response to a Stop cue ('reactive' inhibition). This suppression of action impulses is specifically compromised in a wide range of conditions, including attention-deficit hyperactivity disorder, obsessive–compulsive disorder, Tourette syndrome, schizophrenia and drug abuse [5].

The basal ganglia have long been implicated in response inhibition [6,7], and the classic layout of distinct functional pathways in the basal ganglia [8] was originally devised to account for neurological disorders involving movements that are either insufficiently, or excessively, suppressed. Even accounts of response inhibition that primarily emphasize cortical or midbrain mechanisms have often postulated a key role for basal ganglia circuits [7,9–11]. Despite this, there have been relatively few studies using the high temporal resolution of electrophysiology to investigate in real time how stopping is achieved within the basal ganglia.

The 'stop-signal task' (SST) is a standard test of behavioural inhibition, and the 'race model' [12] is the conventional theoretical framework that provides a quantitative account of stop-signal behaviour (see detailed reviews elsewhere in this issue [13,14]). In the race model, Go and Stop cues elicit corresponding Go and Stop processes that each race towards completion. If the Go process finishes first, movement is initiated. If the Stop process wins, movement is suppressed.

One useful aspect of the SST is that it provides a measure of the speed of stopping (the ‘stop-signal reaction time’), by inferring how fast the Stop process must be to account for reaction time (RT) distributions. The race model has been extended and refined, e.g. to include an interaction between Go and Stop processes towards the end of their evolution [15,16]. However, a basic property of existing race models is that the Stop process is a single entity. One might therefore expect to find a single distinct neural population whose firing rate time course corresponds to the developing race model Stop process. Based on our recent work in rodents we challenge this idea, and provide an updated account of the neural basis of stopping.

2. A stop-signal task in rats

To study neural manifestations of Go and Stop processes in the basal ganglia, we developed a rat version of the SST [17]. Inspired by seminal SST work in non-human primates employing well-controlled saccadic eye movements [18–21], we used ballistic head/neck movements triggered by auditory cues. At the start of each trial, the rat placed its nose into an illuminated nose-poke port. After a brief, randomized time delay (500–1200 ms), one of two auditory Go cues (high or low tone) prompted a corresponding leftward or rightward movement to an adjacent side port. On Stop trials (30% of total), the Go cue was followed by the Stop signal (a white noise burst), indicating that the animal had instead to remain in the centre port (for another approximately 500–900 ms). Correct performance in both Go and Stop trials was rewarded with a sugar pellet.

Behavioural performance in this rat SST was qualitatively and quantitatively similar to human and non-human primate stop tasks. In particular, Go trials showed a characteristic broad RT distribution, with RTs for Failed Stop trials resembling the early (faster) portion of this distribution [17,22,23]. This is consistent with the race model: Failed Stop trials occur when the Go process is quicker than average (Fast Go), while on Correct Stop trials the Go process is slower than average (Slow Go) and so loses the race.

The combination of neurophysiology and the SST enables a powerful and elegant approach for examining the temporal evolution of behavioural control, by comparing neural activity between trial types. On Correct Stop and Failed Stop trials, all the cues presented to the subject are identical, so differences in activity (and behavioural outcome) reflect internal differences—such as trial-by-trial variation in the speed of the Go process. Conversely, to help isolate neural mechanisms involved in stopping one can examine activity differences between Stop and Go trials. This is even more effective if one compares trials for which the Go process is assumed to have a similar initial time course (‘latency-matching’)—i.e. comparing Failed Stop to Fast Go trials, and Correct Stop to Slow Go trials. As this comparison effectively subtracts away activity patterns associated with preparation for movement, any remaining activity differences just after Stop cue onset are good candidates for involvement in a stopping mechanism.

To facilitate such comparisons we used a fixed stop-signal delay (the time between Go and Stop cue onset) within each recording session, so that we could readily align neural activity on the Stop cue (or in the case of Go trials, align on

the time that the Stop cue would have occurred had it been a Stop trial). Although in other variants of the stop-signal task the stop-signal delay is commonly randomized to discourage ‘waiting’ for the stop signal, we did not find any evidence that the rats used a waiting strategy [17]. The stop-signal delay was adjusted between sessions to obtain a similar number of Correct and Failed Stop trials.

3. Does the striatum provide the race model Go process?

A core concept in the physiology of the basal ganglia is disinhibition. Basal ganglia output provides a tonic, GABAergic suppression of structures that promote movements, and interruptions in this suppression facilitate those movements. This mechanism was nicely demonstrated for eye movements in non-human primates, for which the substantia nigra pars reticulata (SNr; a basal ganglia output nucleus) inhibits saccade-promoting neurons in the superior colliculus [24–26]. The SNr itself receives GABAergic input from a subset of striatal neurons (the ‘direct’ striatonigral pathway; [25]), and increases in striatal direct pathway activity facilitate movement [27] by interrupting SNr firing [28]. Though best established for orienting-type movements like saccades, such disinhibition of brainstem motor centres seems to be a general mechanism for controlling a range of fundamental behaviours including locomotion [4]. The impact of basal ganglia output on thalamic targets may be more complex [29] and lead to an influence over neocortex that is more subtle than simple increases or decreases in activity [30]. There is also a lingering controversy over whether the basal ganglia help select which action to perform [31] or just invigorate actions chosen and initiated elsewhere [32,33]. Nonetheless, there is broad agreement that the striatal direct pathway provides some form of ‘Go’ signal. Within the specific context of the SST, human fMRI studies found evidence for Go-related activation in motor striatum (e.g. [7]) as part of an overall cortical–basal ganglia network involved in movement preparation and execution.

We therefore looked for activity patterns of striatal neurons that could map onto the Go process imagined in the race model [22]. What properties ought such activity patterns to have? They should change after the Go cue that initiates the Go process, but substantially before the onset of actual movement. Furthermore, to be involved in selecting (or at least invigorating) a specific movement, activity should distinguish between the different movements. Furthermore, following the latency-matching logic, Go-related activity should be very similar when comparing Fast Go and Failed Stop trials (because both involve a faster-than-usual Go process) and very similar when comparing Slow Go and Correct Stop trials (because both involve a slower-than-usual Go process).

As typically observed in other tasks (e.g. [34]), activity of striatal neurons was heterogeneous and sparse. Nonetheless, a significant subpopulation distinguished the direction of upcoming movement (ipsi- versus contralateral) at least approximately 130 ms before movement onset (figure 1a) and these cells exhibited a sharp firing rate increase during movement initiation (figure 1b). The time course of this activity was also virtually identical between Fast Go and Failed Stop trials (figure 1b). These observations support

Figure 1. Go-related activity in the basal ganglia. (a) Coding of movement direction in striatal units during Go trials, reprinted from [22]. Bars indicate the fraction of units with a significantly higher firing rate in the indicated trial type. Green arrow indicates when the striatal population reached a significant level of direction coding, approximately 130 ms before the onset of movement. (b) Average firing rate of the subset of striatal units that coded for movement direction before movement onset, reprinted from [22]. Different colours indicate mean firing rates in different trial types as indicated. Coloured bars at the top mark significant differences between trial types (cyan, Fast Go versus Slow Go; blue, Slow Go versus Correct Stop; purple, Fast Go versus Failed Stop). (c) Mean spectrogram of the local field potential in globus pallidus (GP) aligned to the onset of the Stop signal (vertical dashed line), reprinted from [17] with permission from Elsevier. Note that there are two transient beta pulses in Correct Stop trials (top panel), but only a single transient beta pulse in Failed Stop trials (bottom panel).

the idea that the striatum conveys the race model Go process that initially evolves independently from a Stop process. Consistent with the disinhibitory character of basal ganglia output, this striatal Go process would enable movements by producing an interruption in SNr firing, that in turn facilitates firing of, for example, superior colliculus neurons [22].

However, the activity of this striatal subpopulation has other noteworthy properties. Although its time course is initially very similar between Slow Go and Correct Stop trials, consistent with conveying a similar slowly evolving Go process, it then abruptly diverges approximately 150 ms after the Stop signal (compare red, dark blue lines in figure 1*b*). The sudden decrease in striatal firing on Correct Stop trials may be due to inhibitory inputs from the globus pallidus (GP; see §6) and might correspond conceptually to a late interaction between Go and Stop processes [15,16].

RT differences between individual Go trials are typically imagined to arise from the Go process building-up at a different rate. A direct correspondence was reported between eye movement RT and a progressive build-up in neuron firing rates in a frontal cortical area [18]. By contrast, when aligned on movement onset the striatal neurons showed a similar build-up rate for both Fast and Slow Go trials (figure 1*a*). This might reflect the involvement of striatal neurons only in some late subcomponent of Going—for example, once they receive more than a critical level of excitatory drive from cortex [35]. Alternatively striatal networks may have Go-relevant dynamics that are not readily apparent in a crude summation of firing rates [36,37], or that do not manifest as a firing rate ‘threshold’ [38,39].

4. Beta rhythms and sensorimotor gating

Activity dynamics that are part of, or interact with, a Go process may also be reflected in local field potential oscillations. Beta (approx. 13–30 Hz) oscillations throughout the basal ganglia and thalamocortical networks seem to have a particular, negative relationship to motor output [40,41]. Spontaneous or evoked beta oscillations seem to delay or slow movements (e.g. [17,42,43]), and there appears to be an important link between beta oscillations and dopamine [44–46]. Beta power is exaggerated in Parkinson’s disease, consistent with an ‘akinetetic’ state, and dopaminergic drugs (and other therapies) that alleviate Parkinsonian akinesia also reduce beta power [47]. However, the role of beta oscillations in sensorimotor processing appears more complex than simply enabling or retarding movements [48,49].

We examined beta oscillations in a set of rat behavioural tasks including the SST [17]. Spontaneous brief epochs of elevated beta (approx. 20 Hz) power occur coherently throughout cortical-basal ganglia networks. Elevated beta can also be prompted by both Go and Stop cues, suggesting that these rhythms are not involved solely in either the Go or in the Stop process of the race model. Notably, however, the Stop cue provoked a beta increase only on Correct, but not Failed, Stop trials (figure 1*c*; see also [50]), even though the sequence of cues presented was identical. We argued that only cues that are actually *used* to direct behaviour evoke beta increases.

Speculatively, the elevated beta may indicate a relatively closed ‘gate’ within the basal ganglia that reduces responsiveness to incoming stimuli. As one possible manifestation of

this, Stop cues were ineffective at arresting behaviour if they arrived during the time of elevated beta produced by the Go cue [17]. By delaying the evolution of a striatal Go process, the beta network state may normally serve the adaptive function of impeding impulsive responding, but become exaggerated and maladaptive in Parkinson’s disease.

Such ideas remain speculative, in large part because beta oscillations are a broadly distributed phenomenon whose origins, propagation and functional impact remain less than clear. Furthermore, pronounced changes in beta power appeared only several hundred milliseconds after cue onset, so are unlikely to be directly part of the critical fast development of a Stop process.

5. Fast progression of Stop cue information through the basal ganglia

We therefore examined the activity of individual basal ganglia neurons during the SST [22], comparing firing rates between latency-matched Go and Stop trials. Prior human imaging work had found evidence that the ‘hyperdirect’ pathway from frontal cortex to the subthalamic nucleus (STN) is an anatomical substrate of the race model Stop process [7] that may suppress movements at the level of basal ganglia output structures such as SNr. Consistent with this, we found neuronal subpopulations in both STN and SNr that showed significant short-latency firing rate increases to the Stop cue (figure 2). No such short-latency Stop response was seen in striatum, consistent with the race model idea that Go and Stop processes initially evolve independently.

The STN neurons had a more ‘sensory’ character, responding to the Stop signal quickly (peak approx. 15 ms) and regardless of whether stopping was actually successful or not. By contrast the responses downstream in SNr were a little slower (peak approx. 35 ms) and more ‘motor’—they strongly correlated with whether the rat would successfully stop in that trial (figure 2), as if reflecting the *outcome* of a race rather than the Stop process alone. Note that this SNr firing increases only on Correct, rather than Failed, Stop trials and is similar to the Stop-cue evoked increase in beta power described in §4 (albeit much faster).

The selective responding of SNr neurons on Correct Stop trials is a form of sensorimotor gating, arising from the relative timing of different inputs [22]. As described earlier (§3), a key late step in the Go process seems to be increases in GABAergic input to SNr from the striatal direct pathway. If this is already underway, then the glutamatergic STN input evoked by the Stop cue is ineffective at driving SNr activity (and behaviourally stopping fails too). In this way, the fundamental idea of a race between Go and Stop processes may map onto a race between distinct anatomical pathways converging on individual SNr neurons.

This is—of course—too simple to serve as a full account of behavioural inhibition, for a variety of reasons. Human studies have often presented the SST as a paradigm of executive function, with a correspondingly prominent role for frontal cortical regions and their hyperdirect projections to STN. Yet in our rat SST it seems unlikely that there is enough time for significant information processing in frontal cortex to occur before the approximately 15 ms latency Stop cue responses we observed in STN (even in primary auditory cortex, neurons typically require approximately 12 ms to

Figure 2. Stop-related activity in the STN (top panels) and SNr (bottom panels; reprinted from [22]). Mean firing rates of Stop cells in Correct and Failed Stop trials and latency-matched control Go trials. Note the sharp increase in firing rate in STN in response to the Stop signal in both Correct and Failed Stop trials. By contrast, SNr stop cells respond to the Stop signal only in Correct Stop trials, and show instead a movement-related decrease during Failed Stop trials. The grey vertical lines mark stop-signal reaction times in the corresponding recording sessions.

respond to white noise stimuli [51]). An alternative potential source of fast auditory STN input is the pedunculo-pontine tegmental nucleus (PPN) [52], and indeed we have recorded neurons there with Stop cue latencies of approximately 9 ms or less [53].

Even more importantly, this fast response is observed not just for the Stop cue, but for the Go cue as well—not a feature one would necessarily expect for a Stop process. In addition, the Stop response is highly transient—for many STN neurons just a single spike—and our modelling suggested that a more sustained change would be necessary to counteract the wave of striatal GABAergic inhibition that normally arrives in SNr just before movement onset.

So, do such speed, transience and lack of selectivity invalidate the fast PPN-STN-SNr signal as a neural mechanism of stopping? Rather, we proposed that stopping actually involves multiple component subprocesses, with this fast signal serving to briefly delay ('Pause') actions rather than fully cancel them [22] (see also figure 3). For example, we note that in variants of the SST where a 'Continue' cue is sometimes played instead of a Stop cue, actions proceed but with prolonged RTs [54], consistent with engagement of a Pause mechanism without complete cancellation. In addition, there is substantial evidence that unexpected cues transiently suppress not just the preparation of one particular

action, but have a more global impact on even unrelated actions (and thoughts also) [55]. This is broadly consistent with the idea of a 'hold-your-horses' role for STN in reactive behavioural inhibition [56] that buys time for more informed decisions on how to proceed.

6. Multiple, complementary Stop mechanisms

Which circuits are then responsible for actually cancelling the movement? As noted in §3, we found evidence for a late interaction between Go and Stop processes at the level of the striatum. Specifically, on Correct Stop trials the developing movement-related activity in striatum abruptly dropped away (figure 1b). We hypothesized that this was due to a strong inhibitory input from the GP. Although the classic ('prototypical') GP neurons project downstream to STN and SNr, the recently characterized 'arkypallidal' cells form massive GABAergic projections exclusively back to striatum [57].

To identify the cell type of the GP units recorded in the SST, we first demonstrated that identified arkypallidal, but not prototypical neurons, greatly reduce their firing rate during slow wave sleep. This signature of arkypallidal neurons allowed us to discriminate them in freely moving rats, by monitoring the same neurons in both the SST and during natural sleep [23].

Consistent with participation in a slower and more selective Stop process, GP Stop response latencies were longer (approx. 60–80 ms) and more selective (unlike STN and SNr, the neurons did not also respond to the Go cue). Arkypallidal neurons had significantly stronger and faster Stop responses than prototypical neurons [23], and intriguingly, these responses immediately preceded the inhibition of striatal movement-related activity in Correct Stop trials (figure 1b). We concluded that arkypallidal neurons have the appropriate timing, selectivity and connectivity to help cancel actions by suppressing the striatal Go process (figure 3b).

There is an obvious survival advantage in being able to respond quickly to events. Yet the more rapid the response, the less sophisticated the preceding information processing can be. In the case of reactive stopping, this presents an inherent trade-off: at one extreme a subject could rapidly interrupt ongoing behaviour for any salient sensory change, while at the other extreme the subject could be much more selective about whether to abandon ongoing motor plans, but much slower to do so.

Having separate, complementary Pause and Cancel mechanisms allows the advantages of both speed and selectivity. The less selective but very fast PPN-STN-SNr response to stimuli blocks movement execution, but only for a brief period, buying time for more detailed assessment of stimulus identity. If this more detailed assessment indicates that stopping is unnecessary, actions can proceed with only a brief delay (tens of milliseconds, compared with the hundreds of milliseconds involved in normal movement preparation). In this way, multiple Stop mechanisms allow a more effective speed–accuracy trade-off than would be possible with a single mechanism alone [58].

As a side note, the two stopping subcomponents are quite reminiscent of the distinct phases of dopamine cell responses to unexpected events: the well-known signalling of reward prediction errors is preceded by a faster, transient response

Figure 3. Sketch of the Pause-then-Cancel model (reprinted from [23] with permission from Elsevier). (a) Illustrations of how multiple stop circuits in the brain may be mapped into the race model framework. In Go trials (left), different reaction times are due to variable evolution of the Go process (green lines). All salient sensory events (Go and Stop cues) activate the Pause circuit (orange line) which leads to a transient elevation of the Go threshold (black dotted line). In Failed Stop trials (middle), the Go process reaches the threshold before this Pause mechanism is effectively engaged. In Correct Stop trials (right), the fast transient elevation of the threshold buys additional time for the slower Stop process to win the race. (b) Simplified circuit diagrams. For the Go process, the direct pathway from striatum to SNr contributes to the initiation of movement (left). Salient sensory stimuli can evoke the rapid Pause mechanism in PPN, STN and SNr that transiently delays the initiation of movements (middle). This delay is exploited by a slower Cancel mechanism involving GP arky pallidal ('Arky') projections to the striatum. (c) Average firing rate time courses for stop-related neurons during Correct Stop trials. Note the successive 'waves' of activity in different basal ganglia subpopulations (STN; SNr; Arky), reflecting different stages of Stop cue information processing.

that is less selective [59]. This is likely no coincidence, because STN provides a major input to dopamine cells [60].

The slower, more selective Stop responses of arky pallidal neurons presumably reflect more elaborate information processing in structures that provide input to GP. It is not yet known which structures these are because specific inputs to arky pallidal neurons have not yet been mapped. However, GP receives direct inputs from various cortical regions including supplementary motor areas [61,62] that have been previously related to stopping [63,64].

7. Implications and limitations

Our experiments confirmed the basic idea that Go and Stop processes race for completion. However, our finding that stopping can be decomposed into complementary subprocesses means that certain assumptions may need to be revisited. In particular, the stop-signal reaction time has been used as a criterion to decide whether neural responses occur early enough to contribute causally to stopping or not (e.g. [65]), but our Pause-then-Cancel model suggests this may be too simple. This is because the stop-signal reaction time should be dominated by the relatively brief time required to engage the Pause mechanism (putting action on temporary hold), even if later mechanisms are essential for complete cancellation. We speculate that these later mechanisms include not only the arky pallidal suppression of striatum described above (§6), but even later phases involved in terminating the cortical and thalamic activity patterns that drive the striatal Go process.

An advantage of the Pause-then-Cancel model is that it makes precise predictions about where and when manipulations should affect RT distributions and stopping performance. For example, we predicted that interfering with the PPN-STN-SNr pathway just at the time of Stop cue onset should decrease stopping performance [53] but

particularly affect those trials which are 'close calls'—as trials in which the Go process was especially slow should be successful even without the Pause mechanism. The result should be a broadening of the Failed Stop RT distribution. Conversely, interference with arky pallidal neurons should affect the efficacy of stopping more generally, without as marked an effect on the RT distribution. Testing such predictions presents some technical challenges—for example, we currently lack an effective means of selectively and briefly inhibiting arky pallidal neurons, and while optogenetic manipulations have the required temporal precision, unless illumination is carefully controlled, it can itself engage the fast STN-SNr pathway.

We should re-emphasize that not all aspects of behavioural control can be reduced to performance in the SST [10]. Furthermore, months of SST training may lead rats (and monkeys) to make use of neural pathways that may not exactly map onto those used by minimally trained humans. Despite these limitations, combining the SST with multi-site basal ganglia neurophysiology has been valuable both for our understanding of basal ganglia operations, and in demonstrating the need to update long-standing psychological models to include multiple component processes of stopping.

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1. Bari A, Robbins TW. 2013 Inhibition and impulsivity: behavioral and neural basis of response control. *Prog. Neurobiol.* **108**, 44–79. (doi:10.1016/j.pneurobio.2013.06.005)
2. Eagle DM, Baunez C, Hutcheson DM, Lehmann O, Shah AP, Robbins TW. 2008 Stop-signal reaction-time task performance: role of prefrontal cortex and subthalamic nucleus. *Cereb. Cortex* **18**, 178–188. (doi:10.1093/cercor/bhm044)
3. Jin X, Tecuapetla F, Costa RM. 2014 Basal ganglia subcircuits distinctively encode the parsing and concatenation of action sequences. *Nat. Neurosci.* **17**, 423–430. (doi:10.1038/nn.3632)
4. Roseberry TK, Lee AM, Lalive AL, Wilbrecht L, Bonci A, Kreitzer AC. 2016 Cell-type-specific control of brainstem locomotor circuits by basal ganglia. *Cell* **164**, 526–537. (doi:10.1016/j.cell.2015.12.037)
5. Ersche KD, Jones PS, Williams GB, Turton AJ, Robbins TW, Bullmore ET. 2012 Abnormal brain structure implicated in stimulant drug addiction. *Science* **335**, 601–604. (doi:10.1126/science.1214463)
6. Neill DB. 1976 Frontal-striatal control of behavioral inhibition in the rat. *Brain Res.* **105**, 89–103. (doi:10.1126/science.1214463)
7. Aron AR, Poldrack RA. 2006 Cortical and subcortical contributions to Stop signal response inhibition: role of the subthalamic nucleus. *J. Neurosci.* **26**, 2424–2433. (doi:10.1523/JNEUROSCI.4682-05.2006)
8. Albin RL, Young AB, Penney JB. 1989 The functional anatomy of basal ganglia disorders. *Trends Neurosci.* **12**, 366–375. (doi:10.1016/0166-2236(89)90074-X)
9. Paré M, Hanes DP. 2003 Controlled movement processing: superior colliculus activity associated with countermanded saccades. *J. Neurosci.* **23**, 6480–6489.
10. Aron AR. 2011 From reactive to proactive and selective control: developing a richer model for stopping inappropriate responses. *Biol. Psychiatry* **69**, 55–68. (doi:10.1016/j.biopsych.2010.07.024)
11. Jahanshahi M, Obeso I, Rothwell JC, Obeso JA. 2015 A fronto-striato-subthalamic-pallidal network for goal-directed and habitual inhibition. *Nat. Rev. Neurosci.* **16**, 719–732. (doi:10.1038/nrn4038)
12. Logan GD, Cowan WB. 1984 On the ability to inhibit thought and action: a theory of an act of control. *Psychol. Rev.* **91**, 295–327. (doi:10.1037/0033-295X.91.3.295)
13. Noorani I. 2017 Towards a unifying mechanism for cancelling movements. *Phil. Trans. R. Soc. B* **372**, 20160191. (doi:10.1098/rstb.2016.0191)
14. Schall JD, Palmeri TJ, Logan GD. 2017 Models of inhibitory control. *Phil. Trans. R. Soc. B* **372**, 20160193. (doi:10.1098/rstb.2016.0193)
15. Boucher L, Palmeri TJ, Logan GD, Schall JD. 2007 Inhibitory control in mind and brain: an interactive race model of countermanding saccades. *Psychol. Rev.* **114**, 376–397. (doi:10.1037/0033-295X.114.2.376)
16. Logan GD, Yamaguchi M, Schall JD, Palmeri TJ. 2015 Inhibitory control in mind and brain 2.0: blocked-input models of saccadic countermanding. *Psychol. Rev.* **122**, 115–147. (doi:10.1037/a0038893)
17. Leventhal DK, Gage GJ, Schmidt R, Pettibone JR, Case AC, Berke JD. 2012 Basal ganglia beta oscillations accompany cue utilization. *Neuron* **73**, 523–536. (doi:10.1016/j.neuron.2011.11.032)
18. Hanes DP, Schall JD. 1996 Neural control of voluntary movement initiation. *Science* **274**, 427–430. (doi:10.1126/science.274.5286.427)
19. Hanes DP, Patterson WF, Schall JD. 1998 Role of frontal eye fields in countermanding saccades: visual, movement, and fixation activity. *J. Neurophysiol.* **79**, 817–834.
20. Brown JW, Hanes DP, Schall JD, Stuphorn V. 2008 Relation of frontal eye field activity to saccade initiation during a countermanding task. *Exp. Brain Res.* **190**, 135–151. (doi:10.1007/s00221-008-1455-0)
21. Schall JD, Godlove DC. 2012 Current advances and pressing problems in studies of stopping. *Curr. Opin. Neurobiol.* **22**, 1012–1021. (doi:10.1016/j.conb.2012.06.002)
22. Schmidt R, Leventhal DK, Mallet N, Chen F, Berke JD. 2013 Canceling actions involves a race between basal ganglia pathways. *Nat. Neurosci.* **16**, 1118–1124. (doi:10.1038/nn.3456)
23. Mallet N, Schmidt R, Leventhal D, Chen F, Amer N, Boraud T, Berke JD. 2016 Arkypallidal cells send a stop signal to striatum. *Neuron* **89**, 308–316. (doi:10.1016/j.neuron.2015.12.017)
24. Hikosaka O, Wurtz RH. 1985 Modification of saccadic eye movements by GABA-related substances. I. Effect of muscimol and bicuculline in monkey superior colliculus. *J. Neurophysiol.* **53**, 266–291.
25. Chevalier G, Vacher S, Deniau JM, Desban M. 1985 Disinhibition as a basic process in the expression of striatal functions. I. The striato-nigral influence on tecto-spinal/tecto-diencephalic neurons. *Brain Res.* **20**, 215–226. (doi:10.1016/0006-8993(85)90213-6)
26. Redgrave P, Marrow L, Dean P. 1992 Topographical organization of the nigrotectal projection in rat: evidence for segregated channels. *Neuroscience* **50**, 571–595. (doi:10.1016/0306-4522(92)90448-B)
27. Kravitz AV, Freeze BS, Parker PRL, Kay K, Thwin MT, Deisseroth K, Kreitzer AC. 2010 Regulation of parkinsonian motor behaviours by optogenetic control of basal ganglia circuitry. *Nature* **29**, 622–626. (doi:10.1038/nature09159)
28. Freeze BS, Kravitz AV, Hammack N, Berke JD, Kreitzer AC. 2013 Control of basal ganglia output by direct and indirect pathway projection neurons. *J. Neurosci.* **33**, 18 531–18 539.
29. Goldberg JH, Farries MA, Fee MS. 2013 Basal ganglia output to the thalamus: still a paradox. *Trends Neurosci.* **36**, 695–705. (doi:10.1523/JNEUROSCI.1278-13.2013)
30. Oldenburg IA, Sabatini BL. 2015 Antagonistic but not symmetric regulation of primary motor cortex by basal ganglia direct and indirect pathways. *Neuron* **86**, 1174–1181. (doi:10.1016/j.neuron.2015.05.008)
31. Redgrave P, Prescott TJ, Gurney K. 1999 The basal ganglia: a vertebrate solution to the selection problem? *Neuroscience* **89**, 1009–1023. (doi:10.1016/S0306-4522(98)00319-4)
32. Turner RS, Desmurget M. 2010 Basal ganglia contributions to motor control: a vigorous tutor. *Curr. Opin. Neurobiol.* **20**, 704–716. (doi:10.1016/j.conb.2010.08.022)
33. Dudman JT, Krakauer JW. 2016 The basal ganglia: from motor commands to the control of vigor. *Curr. Opin. Neurobiol.* **37**, 158–166. (doi:10.1016/j.conb.2016.02.005)
34. Schultz W, Romo R, Ljungberg T, Mirenovic J, Hollerman JR, Dickson A. 1995 Reward-related signals carried by dopamine neurons. In *Models of information processing in the basal ganglia* (eds JC Houk, JL Davis, DG Beiser), pp. 233–248. Cambridge, MA: MIT Press.
35. Lo C-C, Wang X-J. 2006 Cortico–basal ganglia circuit mechanism for a decision threshold in reaction time tasks. *Nat. Neurosci.* **9**, 956–963. (doi:10.1038/nn1722)
36. Angulo-García D, Berke JD, Torcini A. 2016 Cell assembly dynamics of sparsely-connected inhibitory networks: a simple model for the collective activity of striatal projection neurons. *PLoS Comput. Biol.* **12**, e1004778. (doi:10.1371/journal.pcbi.1004778)
37. Bahuguna J, Aertsen A, Kumar A. 2015 Existence and control of Go/No-Go decision transition threshold in the striatum. *PLoS Comput. Biol.* **11**, e1004233. (doi:10.1371/journal.pcbi.1004233)
38. Churchland MM, Cunningham JP, Kaufman MT, Ryu SI, Shenoy KV. 2010 Cortical preparatory activity: representation of movement or first cog in a dynamical machine? *Neuron* **68**, 387–400. (doi:10.1016/j.neuron.2010.09.015)
39. Ding L, Gold JJ. 2010 Caudate encodes multiple computations for perceptual decisions. *J. Neurosci.* **30**, 15 747–15 759. (doi:10.1523/JNEUROSCI.2894-10.2010)
40. Reimer J, Hatsopoulos NG. 2010 Periodicity and evoked responses in motor cortex. *J. Neurosci.* **30**, 11 506–11 515. (doi:10.1523/JNEUROSCI.5947-09.2010)
41. Feingold J, Gibson DJ, DePasquale B, Graybiel AM. 2015 Bursts of beta oscillation differentiate postperformance activity in the striatum and motor cortex of monkeys performing movement tasks. *Proc. Natl Acad. Sci. USA* **112**, 13 687–13 692. (doi:10.1073/pnas.1517629112)
42. Pogosyan A, Gaynor LD, Eusebio A, Brown P. 2009 Boosting cortical activity at beta-band frequencies slows movement in humans. *Curr. Biol.* **19**, 1637–1641. (doi:10.1016/j.cub.2009.07.074)

43. Misra G, Ofori E, Chung JW, Coombes SA. In press. Pain-related suppression of beta oscillations facilitates voluntary movement. *Cereb. Cortex*. (doi:10.1093/cercor/bhw061)
44. Sharott A, Magill PJ, Harnack D, Kupsch A, Meissner W, Brown P. 2005 Dopamine depletion increases the power and coherence of beta-oscillations in the cerebral cortex and subthalamic nucleus of the awake rat. *Eur. J. Neurosci.* **21**, 1413–1422. (doi:10.1111/j.1460-9568.2005.03973.x)
45. Berke JD. 2009 Fast oscillations in cortical-striatal networks switch frequency following rewarding events and stimulant drugs. *Eur. J. Neurosci.* **30**, 848–859. (doi:10.1111/j.1460-9568.2009.06843.x)
46. Brittain J-S, Brown P. 2014 Oscillations and the basal ganglia: motor control and beyond. *Neuroimage* **85**, 637–647. (doi:10.1016/j.neuroimage.2013.05.084)
47. Levy R, Ashby P, Hutchison WD, Lang AE, Lozano AM, Dostrovsky JO. 2002 Dependence of subthalamic nucleus oscillations on movement and dopamine in Parkinson's disease. *Brain* **125**, 1196–1209. (doi:10.1093/brain/awf128)
48. Engel AK, Fries P. 2010 Beta-band oscillations—signalling the status quo? *Curr. Opin. Neurobiol.* **20**, 156–165. (doi:10.1016/j.conb.2010.02.015)
49. Tan H *et al.* 2015 Subthalamic nucleus local field potential activity helps encode motor effort rather than force in parkinsonism. *J. Neurosci.* **35**, 5941–5949. (doi:10.1523/JNEUROSCI.4609-14.2015)
50. Swann N, Tandon N, Canolty R, Ellmore TM, McEvoy LK, Dreyer S, DiSano M, Aron AR. 2009 Intracranial EEG reveals a time- and frequency-specific role for the right inferior frontal gyrus and primary motor cortex in stopping initiated responses. *J. Neurosci.* **29**, 12 675–12 685. (doi:10.1523/JNEUROSCI.3359-09.2009)
51. Polley DB, Read HL, Storace DA, Merzenich MM. 2007 Multiparametric auditory receptive field organization across five cortical fields in the albino rat. *J. Neurophysiol.* **97**, 3621–3638. (doi:10.1152/jn.01298.2006)
52. Pan W-X, Hyland BI. 2005 Pedunculo-pontine tegmental nucleus controls conditioned responses of midbrain dopamine neurons in behaving rats. *J. Neurosci.* **25**, 4725–4732. (doi:10.1523/JNEUROSCI.0277-05.2005)
53. Chen F, Schmidt R, Mallet NM, Berke JD. 2014 Fast sensory responses in the pedunculo-pontine nucleus help pause actions. In *Society for Neuroscience Meeting, Washington DC, 15–19 November 2014*, poster 442. 16/KK5. Washington, DC: Society for Neuroscience.
54. Sharp DJ, Bonnelle V, De Boissezon X, Beckmann CF, James SG, Patel MC, Mehta MA. 2010 Distinct frontal systems for response inhibition, attentional capture, and error processing. *Proc. Natl Acad. Sci. USA* **107**, 6106–6111. (doi:10.1073/pnas.1000175107)
55. Wessel JR, Jenkinson N, Brittain J-S, Voets S, HEM, Aziz TZ, Aron AR. 2016 Surprise disrupts cognition via a fronto-basal ganglia suppressive mechanism. *Nat. Commun.* **7**, 11195. (doi:10.1038/ncomms11195)
56. Frank MJ. 2006 Hold your horses: a dynamic computational role for the subthalamic nucleus in decision making. *Neural Netw.* **19**, 1120–1136. (doi:10.1016/j.neunet.2006.03.006)
57. Mallet N, Micklem BR, Henny P, Brown MT, Williams C, Bolam JP, Nakamura KC, Magill PJ. 2012 Dichotomous organization of the external globus pallidus. *Neuron* **74**, 1075–1086. (doi:10.1016/j.neuron.2012.04.027)
58. Trimmer PC, Houston AI, Marshall JA, Bogacz R, Paul ES, Mendl MT, McNamara JM. 2008 Mammalian choices: combining fast-but-inaccurate and slow-but-accurate decision-making systems. *Proc. R. Soc. B* **275**, 2353–2361. (doi:10.1098/rspb.2008.0417)
59. Schultz W. 2016 Dopamine reward prediction-error signalling: a two-component response. *Nat. Rev. Neurosci.* **17**, 183–195. (doi:10.1038/nrn.2015.26)
60. Watabe-Uchida M, Zhu L, Ogawa SK, Vamanrao A, Uchida N. 2012 Whole-brain mapping of direct inputs to midbrain dopamine neurons. *Neuron* **74**, 858–873. (doi:10.1016/j.neuron.2012.03.017)
61. Naito A, Kita H. 1994 The cortico-pallidal projection in the rat: an anterograde tracing study with biotinylated dextran amine. *Brain Res.* **653**, 251–257. (doi:10.1016/0006-8993(94)90397-2)
62. Milardi D *et al.* 2015 Basal ganglia network by constrained spherical deconvolution: a possible cortico-pallidal pathway? *Mov. Disord.* **30**, 342–349. (doi:10.1002/mds.25995)
63. Nachev P, Wydell H, O'Neill K, Husain M, Kennard C. 2007 The role of the pre-supplementary motor area in the control of action. *Neuroimage* **36**, T155–T163. (doi:10.1016/j.neuroimage.2007.03.034)
64. Cai W, George JS, Verbruggen F, Chambers CD, Aron AR. 2012 The role of the right presupplementary motor area in stopping action: two studies with event-related transcranial magnetic stimulation. *J. Neurophysiol.* **108**, 380–389. (doi:10.1152/jn.00132.2012)
65. Stuphorn V, Taylor TL, Schall JD. 2000 Performance monitoring by the supplementary eye field. *Nature* **408**, 857–860. (doi:10.1038/35048576)