Type 2 Diabetes Prevention: Implications of Hemoglobin A1c Genetics

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Manuscript submitted May 3, 2015; resubmitted June 29, 2015; accepted July 22, 2015

■ Abstract

Hemoglobin A1c (HbA1c) is a biomarker used for population-level screening of type 2 diabetes (T2D) and risk stratification. Large-scale, genome-wide association studies have identified multiple genomic loci influencing HbA1c. We discuss the challenges of classifying these genomic loci as influencing HbA1c through glycemic or nonglycemic pathways, based on their probable biology and pleiotropic associations with erythrocyte traits. We show that putative nonglycemic genetic variants have a measurable, albeit small, impact on the classification of T2D status by HbA1c in white and Asian populations. Accounting for their effect on HbA1c may be relevant when screening populations with higher frequencies of nonglycemic HbA1c-altering alleles.

As carriers of such HbA1c-altering alleles have HbA1c levels that may not accurately reflect overall glycemia, we describe how accounting for genotype may improve the performance of HbA1c in T2D prediction models and risk stratification, allowing for lifestyle intervention strategies to be directed towards those who are truly at elevated risk for developing T2D. In a Mendelian randomization framework, genetic variants can be used as instrumental variables to estimate causal relationships between HbA1c and T2D-related complications. This approach may help to support or refute HbA1c as an appropriate biomarker for long-term health outcomes in the general population.

Keywords: type 2 diabetes · HbA1c · fasting glucose · GWAS · glycemic pathway · glycemic trait · index SNP

1. Introduction

In the era of precision medicine, we witness a growing emphasis on personalized healthcare. While precision medicine has often been associated with targeted pharmacologic therapies, its overarching goal extends to other aspects of healthcare, e.g., customized prevention strategies, appropriate diagnostic testing, and the use of biomarkers for disease prediction and health screening [1, 2]. Hemoglobin A1c (HbA1c) is a biomarker that has been used for type 2 diabetes (T2D) diagnosis and prediction [3]. The HbA1c test measures the proportion of glycated hemoglobin in the blood, an irreversible chemical modification of the hemoglobin molecule by blood glucose, which reflects ambient glycemia averaged over the preceding 2-3 months [4], the average life span of a red blood cell

(RBC). HbA1c levels are in part genetically determined. Studies on nondiabetic twins reported heritability estimates for HbA1c levels of around 57-75%, whereas the heritability of fasting glucose (FG) was less, 38-66% [5, 6]. While previous cross-sectional examinations have shown generally good agreement between HbA1c and FG levels for diagnosing T2D in varying populations, some ethnic differences exist [7-10]. It also remains unclear to what extent the genetic factors that influence HbA1c and FG are shared [5, 6], and whether these shared genetic factors are important in T2D prediction.

HbA1c is the only test for T2D that is not directly a measurement of blood glucose [3]. While there is strong evidence supporting the use of HbA1c to identify individuals with early dysglycemia, nonglycemic factors can influence HbA1c

Abbreviations:

ABCB11	ATP-binding cassette, subfamily B, member 11
2hrG	2-hour glucose
AGEN	Asian Genetics Epidemiology Network
DCCT	Diabetes Control and Complications Trial
DIAGRAM	Diabetes Genetics Replications and Meta-Analysis
ENCODE	Encyclopedia of DNA Elements
FG	fasting glucose
FEV1	forced expiratory volume in 1 second
FN3K	fructosamine-3-kinase
GCK	glucokinase
GWAS	genome-wide association study
GRASP	Genome-Wide Repository of Associations Between SNPs and Phenotypes
HbA1c	hemoglobin A1c
HFE	human hemochromatosis protein
LD	linkage disequilibrium
MCHC	mean corpuscular hemoglobin concentration
MR	Mendelian randomization
MAGIC	Meta-Analysis of Glycemic and Insulin-Related Traits Consortium
NGSP	National Glycohemoglobin Standardization Program
PI	proinsulin
RBC	red blood cell
SLC30A8	solute carrier family 30 (zinc transporter), member 8
SNP	single nucleotide polymorphism
T2D	type 2 diabetes
TCF7L2	transcription factor 7 like 2
UKPDS	United Kingdom Prospective Diabetes Study
UCSC	University of California, Santa Cruz

variability in populations. These include hematologic conditions that affect RBC turnover or shorten RBC intravascular lifespan (e.g. hereditary hemolytic anemia and blood loss), and falsely lower HbA1c levels [11], iron deficiency anemia, and iron deficiency that have been associated with higher HbA1c readings [12, 13]. Chemical modifications of hemoglobin (e.g., carbamylated hemoglobin in renal failure), hemoglobin variants (e.g. HbS, HbC, HbD, HbE), and elevated fetal hemoglobin can interfere with some laboratory assay methods and profoundly affect measurement accuracy [14, 15]. Notably, some of these nonglycemic factors affecting measured HbA1c levels are genetically determined.

This review has two sequential parts. Firstly, we summarize the main genetic discoveries from pivotal genome-wide association studies (GWAS) on HbA1c in the last 8 years, and discuss mechanisms by which these GWAS genomic loci may influence HbA1c levels. Secondly, we highlight the potential public health and clinical implications of HbA1c genetics with respect to T2D population screening and prediction.

2. Discovery of HbA1c-related genomic loci

2.1 HbA1c discovery GWAS

To date, discovery GWAS have identified 17 HbA1c-related genomic loci. In 2008, Pare and colleagues conducted a GWAS on 14,618 nondiabetic participants of European ancestry in the Women's Genome Health Study [16]. They reported that HbA1c was associated with genetic variation at four genomic loci (*GCK*, rs730497, $p = 2.8 \times 10^{-12}$; *SLC30A8*, rs13266634, $p = 9.8 \times 10^{-8}$; *G6PC2/ABCB11*, rs1402837, $p = 6.8 \times 10^{-10}$; *HK*, rs7072268; $p = 6.4 \times 10^{-9}$). In 2010, Soranzo and colleagues conducted a larger meta-analysis of GWAS on 46,368 nondiabetic adults of European ancestry in the Meta-analysis of Glycemic and Insulin-related traits Consortium (MAGIC). They reported 10 genomic loci associated with HbA1c, six of which were novel, *FN3K* (rs1046896, $p = 1.6 \times 10^{-26}$), *HFE* (rs1800562, $p = 2.6 \times 10^{-20}$), *TMPRSS6* (rs855791, $p = 2.7 \times 10^{-14}$), *ANK1* (rs4737009, $p = 6.1 \times 10^{-12}$), *ATP11A/TUBGCP3* (rs7998202, $p = 5.2 \times 10^{-9}$), *SPTA1* (rs2779116, $p = 2.8 \times 10^{-9}$), and four loci that had been reported to be associated with HbA1c (*HK1*, *GCK*, *G6PC2/ABCB11*, and *MTNR1B*) [17].

This meta-analysis of GWAS was followed closely by a smaller GWAS ($n = 1,782$) conducted by Franklin and colleagues. The authors reported a variant in an intron of *TCF7L2* associated with HbA1c at subthreshold GW significance (rs7903146, $p = 1.48 \times 10^{-7}$) [18]. In 2012, Ryu and Lee conducted a GWAS in an Asian population ($n = 8,057$) within the Korean Association Study, and identified novel HbA1c-associated genetic variants in *CDKAL1*; the strongest signal being an intronic variant, rs7747752 ($p = 1.0 \times 10^{-11}$) [19]. In 2014, Chen and colleagues published the second-largest meta-analysis of HbA1c GWAS on 21,026 individuals from East Asian cohorts in the consortium of the Asian Genetics Epidemiology Network (AGEN). This study reported nine variants at genome-wide significance, four of which were novel (*TMEN79*, rs6684514, $p = 1.3 \times 10^{-23}$; *HB21L/MYB*, rs9399137, $p = 8.5 \times 10^{-15}$; *MYO9B*, rs11667918, $p = 9.0 \times 10^{-12}$, and *CYBA*, rs9933309, $p = 1.1 \times 10^{-28}$), one in an intron of *CDKAL1*, rs7772603, a locus previous implicated in the Korean study, and four loci (*G6PC2/ABCB11*, *GCK*, *ANK1*, and *FN3K*) which were previously identified by MAGIC [20]. This result demonstrated that HbA1c variants carried by Europeans are generally transferable to East Asians. It is anticipated

that larger trans-ethnic meta-analyses of GWAS may be required to boost the statistical power for discovering novel HbA1c-related loci, and detecting genetic variants at lower frequencies.

2.2 Glycemic and nonglycemic pathways: genetic architecture of HbA1c in relation to that of T2D and related traits

Using a combination of cross-trait association analyses, mediation analyses, and known biological function of nearby genes, previous GWAS have attempted to classify 17 discovered HbA1c-related variants as possibly “glycemic” or “nonglycemic” [17, 20]. **Figure 1** depicts overlapping sets of HbA1c-related loci with T2D, FG, proinsulin (PI), and 2-hr glucose (2hrG) GWAS loci; six were considered glycemic as they were associated at genome-wide significance with either FG or T2D, or both, three of which were additionally associated with fasting PI or 2hrG. Notably, none of the HbA1c-related loci were associated with fasting insulin (FI) at genome-wide significance [21-23]. Index single nucleotide polymorphisms (SNPs) within the six glycemic loci were as follows:

1. rs7903146: an intronic variant of *TCF7L2*, a gene encoding a transcription factor involved in the Wnt signaling pathway and the proliferation of pancreatic beta-cells [24].
2. rs13266634: a missense mutation in *SLC30A8*, encoding a zinc transporter associated with beta-cell function [25, 26] which led to the discovery of rare protein-truncating *SLC30A8* variants, conferring T2D protection [27].
3. rs1387153: a variant 29 kb from the 5' end of *MTNR1B*, encoding the receptor for melatonin, the principle hormone secreted by the pineal gland that may coordinate circadian patterns and insulin secretion [28, 29].
4. rs1799884: a variant 45 bp from the 5' end of *GCK*, encoding glucokinase, a hexokinase that phosphorylates glucose, the initial step in glucose metabolism pathways primarily in the liver and pancreas [30].
5. rs7772603: an intronic SNP in *CDKAL1*, encoding a transfer ribonucleic acid (tRNA) modification enzyme that may influence insulin secretion [31].
6. rs552976: an intronic variant in *ABCB11*, encoding an ABC transporter, bile salt export pump, involved in hepatic bile production [32]. This particular SNP is in close linkage disequilibrium ($LD \geq 0.8$) with other

SNPs in the intergenic region near *G6PC2*, which encodes a key enzyme in the terminal step of gluconeogenic and glycogenolytic pathways [33].

These genetic variants implicate diverse biochemical pathways underlying T2D risk, glucose regulation, and subsequent glycemia quantified by elevated HbA1c values.

The remaining 11 HbA1c-related genomic loci have been classified as possibly “nonglycemic”. Seven (*FN3K*, *HFE*, *TMPRSS6*, *ANK1*, *SPTA1*, *ATP11A*, and *HK1*) were first reported in European populations [16, 17], and the remaining four (*TMEN79*, *HB21L/MYB*, *MYO9B*, and *CYBA*) were reported in Asian populations [20]. These genomic loci were either not strongly associated with FG, remained associated with HbA1c even after adjustment for FG in mediation analyses, or were strongly associated with red blood cell traits (e.g. hemoglobin levels, mean corpuscular volume, and mean corpuscular hemoglobin concentration) in cross-trait look-ups [17, 20].

While some genomic loci were easily classified as possibly “glycemic” or “nonglycemic”, it was more challenging in the case of others to assign a potential mechanistic relationship to HbA1c levels. For instance, *HK1*, encoding hexokinase, was considered “glycemic” when it was first reported by Pare *et al.* in 2008 (rs7072268, intron in *HK1*, $p = 6.4 \times 10^{-9}$) [16]. Hexokinase is an enzyme that phosphorylates glucose to produce glucose-6-phosphate, the initial step in glucose metabolism. While hexokinase-1 is ubiquitously expressed, its presence is important in RBCs as they depend exclusively on glycolysis for ATP generation and energy production. Several other lines of evidence indicate that *HK1* may influence RBC biology:

1. Mutations in *HK1* are associated with hexokinase deficiency and hereditary nonspherocytic hemolytic anemia [34, 35].
2. In Soranzo *et al.*, the association between the lead SNP in *HK1* (rs16926246-C, $p = 3.1 \times 10^{-54}$) and HbA1c did not attenuate substantially with adjustment for FG (unadjusted $\beta = 0.073$ (SE 0.007) % points, $p = 4.8 \times 10^{-26}$ vs. FG-adjusted $\beta = 0.069$ (0.006), $p = 6.4 \times 10^{-30}$) [17].
3. *HK1* was associated with hemoglobin levels in population-based cohorts that contributed to this meta-analysis of GWAS ($p = 1.1 \times 10^{-3}$, $n = 7,534$) [36]. It is possible that variations in *HK1* may affect HbA1c via pleiotropic mechanisms, involving both glucose

Figure 1. Venn diagram of genomic loci associated with HbA1c, fasting glucose, fasting proinsulin, 2-hour glucose, and type 2 diabetes detected by GWAS. This Venn diagram depicts overlapping sets of HbA1c-related genomic loci which are also associated with type 2 diabetes (T2D, Mahajan *et al.* 2014, DIABetes Genetics Replication and Meta-analysis (DIAGRAM) consortium), fasting glucose (FG), fasting insulin (FI), 2-hour glucose (2hrG, Scott *et al.*, 2012, Meta-Analysis of Glucose and Insulin related traits Consortium (MAGIC)), and fasting proinsulin (PI, Strawbridge *et al.*, 2011, MAGIC) in GWAS [21-23]. PI, a precursor of insulin in secretory granules of the pancreatic beta-cell, has minimal insulin activity until it has been processed to insulin. Elevated circulating levels of PI are associated with impaired beta-cell function, decreased insulin processing or secretion, or “beta-cell stress” resulting from insulin resistance [87]. PI, similarly to HbA1c, FG, 2hrG, and FI, is a predictor of future T2D [88]. Six HbA1c GWAS loci are glycemic: one (*G6PC2/ABCB11*) is associated with FG only, and five are associated with both T2D and FG GWAS loci, with three of these five (*TCF7L2*, *GCK/YKT6*, and *SLC30A8*) also associating with either 2hrG or PI. The HbA1c index SNP in *ANK1* was considered “nonglycemic” when it was first discovered, although a T2D index SNP within *ANK1* has been found in later GWAS. None of the HbA1c GWAS loci overlap with FI.

metabolism and RBC biology, e.g., impaired intra-cellular glucose metabolism that impact RBC survival.

Another genomic locus that did not classify cleanly is *ANK1*, a gene encoding ankyrin that connects integral membrane proteins to underlying spectrin-actin cytoskeleton [37]. Mutations in erythrocytic ankyrin 1 result in ankyrin-deficient, hereditary spherocytosis [38]. In 2010, Soranzo *et al.* identified two independent HbA1c GWAS signals ($r^2 = 0.009$) residing in introns of *ANK1*,

rs6474359-T (risk allele frequency = 0.97, $\beta = 0.06$ (0.04-0.08), $p = 1.2 \times 10^{-8}$) and rs4737009-A (risk allele frequency = 0.24, $\beta = 0.03$ (0.02-0.04), $p = 6.1 \times 10^{-12}$) [17]. Mediation analyses indicated that *ANK1* likely influenced HbA1c independently of FG. Given the known biological function of *ANK1*, the authors concluded that this genomic locus likely influenced HbA1c levels through mechanisms involving RBC biology and not glucose regulation. Using the Genome-Wide Repository of Associations Between SNPs and Phenotypes (GRASP) database, SNPs in *ANK1* was also found to be associated with mean corpuscular hemoglobin concentration (MCHC; rs4737009; $p = 4.9 \times 10^{-11}$; $n = 135,367$) in a large meta-analysis of GWAS on RBC phenotypes [39].

In 2014, a trans-ethnic meta-analysis of GWAS in T2D by the Diabetes Genetics Replications and Meta-Analysis (DIAGRAM) consortium reported a T2D lead SNP, rs516946-C (risk allele frequency = 0.77, OR 1.10 (1.06-1.15), $p = 2 \times 10^{-7}$), residing in the same *ANK1* locus, specifically in an intron closer to the protein-coding exon. This SNP may be regarded as a third independent signal ($r^2 = 0.004$). These findings challenge the notion that *ANK1* influences HbA1c via RBC biology only. Incidentally, another independent SNP, rs7006290 ($r^2 = 0.05$), in the *ANK1* locus has been associated with pulmonary function decline (forced expiratory volume in 1 second (FEV1) decline) in asthmatics ($p = 5.2 \times 10^{-6}$, $n = 4,118$). Indeed, variations in *ANK1* may have pleiotropic effects on different diseases and phenotypic traits. **Figure 2** features a UCSC (University of California, Santa Cruz) browser snapshot of the *ANK1* locus, chr8:41,460,000-41,760,000 (300 kb), containing GWAS SNPs for T2D, HbA1c, FEV1 decline, and MCHC. The HbA1c-related SNP, rs4737009, falls on a putative enhancer (H3KMe1 histone mark), a DNase 1 hypersensitivity area, and 12 transcription factor binding sites derived

UCSC browser snapshot of the *ANK1* genomic locus

Figure 2. Genetic variants in the *ANK1* genomic locus and their associated phenotypes. This figure is a UCSC browser snapshot of the *ANK1* genomic locus, chr8:41,460,000-41,760,000 (300 kb), illustrating the relative positions of GWAS index SNPs for type 2 diabetes (T2D), hemoglobin A1c (HbA1c), forced expiratory volume in 1 second (FEV1) decline, and mean corpuscular hemoglobin concentration (MCHC) residing in introns of *ANK1*. The figure also shows Encyclopedia of DNA Elements (ENCODE) functional elements located near the HbA1c index SNP, rs4737009. These include an H3KMe1 histone mark, which is suggestive of a putative enhancer, and a DNase 1 hypersensitivity site, indicating a DNase-sensitive regulatory region. The zoomed-in plot shows that this HbA1c index SNP falls within 12 transcription factor binding sites (TFBS) derived from ChIP-seq experiments. These are demarcated by gray boxes. Three of these TFBS have canonical motifs for the corresponding factor (green highlight), further suggesting that the region has regulatory function impacting gene expression.

from ChIP-seq experiments, 3 of which have canonical motifs for the corresponding factor. Detailed annotation of the genomic region in close proximity to the HbA1c index SNP illuminated functional elements that potentially regulate gene expression. Laboratory experiments designed to test whether genetic variation in this locus affects transcript expression in various tissue types may offer deeper insight into the pleiotropic effects exerted by *ANK1*.

HbA1c genetics provide an additional line of evidence for the genetic determinants of dysglycemia, allowing us to map out better the genetic architecture of T2D and related traits. Risk alleles may be present in higher frequencies in certain populations [40-43], which may partly explain

some of the ethnic disparities in T2D risk and population distribution of HbA1c. Prioritizing genomic loci that are associated with multiple T2D-related traits for targeted sequencing and detailed laboratory experimentation with a focus on tissue-specific transcript expression may be a fruitful strategy for identifying causal variants. Findings may suggest mechanisms that mediate genetic effects in T2D-susceptible persons, potentially unveiling novel therapeutic targets for personalized medicine. While glycemic HbA1c variants may be particularly relevant for revealing biological pathways related to T2D susceptibility, nonglycemic HbA1c variants contribute to the inaccuracy of HbA1c as a population-level screening tool for T2D detection and risk stratification.

Table 1. Reclassification table of measured HbA1c and nonglycemic HbA1c SNP-adjusted HbA1c to estimate the population-level impact of nonglycemic HbA1c SNPs on T2D screening in European and Asian cohorts

Population	Measured FG	Measured HbA1c	Nonglycemic SNP-adjusted HbA1c		Total number
			<6.5%	≥6.5%	
European	≥7 mmol/l	<6.5%	358(60.4%)	1(0.2%)	359(60.5%)
		≥6.5%	13(2.2%)	221(37.3%)	234(39.5%)
		Total	371(62.6%)	222(37.4%)	593(100%)
	<7 mmol/l	<6.5%	9,404(98.8%)	1(0.01%)	9,405(98.8%)
Asian	≥7 mmol/l	<6.5%	57(31.5%)	1(0.6%)	58(32.0%)
		≥6.5%	3(1.7%)	120(66.3%)	123(68.0%)
		Total	60(33.1%)	121(66.9%)	181(100%)
	<7 mmol/l	<6.5%	13,813(96.4%)	40(0.3%)	13,853(96.7%)
		≥6.5%	172(1.2%)	304(2.1%)	476(3.3%)
		Total	13,985(97.6%)	344(2.4%)	14,329(100%)

3. Implications for T2D diagnosis and population screening

In 2010, the American Diabetes Association (ADA) recommended the use of HbA1c as one of three biochemical criteria for diagnosing clinical diabetes. Since then, 6.5% has been widely adopted as the HbA1c threshold for clinical diagnosis of T2D [3]. Putative nonglycemic HbA1c genetic variants may result in the misclassification of individuals around this HbA1c threshold for T2D diagnosis. **Table 1** shows two net reclassification analyses to estimate the population-level impact of putative nonglycemic HbA1c loci on screening for undiagnosed T2D; the first analysis was by Soranzo *et al.* in European samples, the second by Chen *et al.* in Asian samples [17, 20].

Putative nonglycemic HbA1c-associated SNPs affect erythrocyte turnover or abnormal glycation of hemoglobin, which may contribute to elevated or reduced HbA1c relative to glycemia. To determine the impact of these SNPs on T2D classification, Soranzo and colleagues calculated the net reclassification around the 6.5% threshold for T2D diagnosis attributable to the effects of seven putative nonglycemic HbA1c loci (FN3K, HFE, TMPRSS6, ANK1, SPTA1, ATP11A, and HK1) compared to undiagnosed T2D, defined by FG ≥ 7 mmol/l, in 10,110 participants from the Framingham Heart Study and the Atherosclerosis Risk in Communities Study [17]. The investigators first excluded individuals who either had a physician-diagnosed T2D or who were taking anti-diabetic medication. To identify the proportion of individuals whose

T2D diagnosis by measured HbA1c ≥ 6.5% were reclassified after adjusting for the contribution of the seven nonglycemic genetic variants, the authors calculated a SNP-adjusted HbA1c by increasing or decreasing the measured HbA1c by the difference between the number of HbA1c-raising alleles (0, 1, or 2) and 2 times the allelic frequency in the sample, multiplied by the effect size of each nonglycemic SNP. Subsequently, they constructed two-by-two tables representing the proportion of individuals whose T2D diagnoses were discordant by measured HbA1c and SNP-adjusted HbA1c (**Table 1**). Of 593 individuals (5.9% of total sample) with undiagnosed T2D (FG ≥ 7 mmol/l), 234 (39.5%) had measured HbA1c ≥ 6.5%, and

222 (37.4%) had a SNP-adjusted HbA1c ≥ 6.5%. This means that 2% of those with FG ≥ 7 mmol/l had a measured HbA1c ≥ 6.5%, which was adjusted to < 6.5% after accounting for the contribution of the seven putative non-glycemic SNPs (**Figure 3**). While FG was used as the reference standard in this study, we acknowledge that FG has greater biological variability than HbA1c, and so it remains unclear whether a single measurement of FG is a better screening test than measured HbA1c or SNP-adjusted HbA1c.

Chen and colleagues addressed the population impact of six putative nonglycemic HbA1c SNPs (*TMEN79*, *HB21L/MYB*, *MYO9B*, *CYBA*, *ANK1*, and *FN3K*) in cohorts of Asian ancestry (n = 15,150) using a similar reclassification analysis. In this study, the proportion of undiagnosed T2D based on FG ≥ 7mmol/l was smaller than in the European study (1.2% (n = 181) in Asians vs. 5.9% in Europeans). About two in three individuals with undiagnosed T2D (n = 123 (68.0%)) in this Asian study had measured HbA1c ≥ 6.5%. Adjusting for the putative nonglycemic loci did not result in much net reclassification. However, there was marked reclassification among the 14,329 individuals without T2D (i.e. FG < 7mmol/l), with 476 (3.3%) nondiabetic individuals classified as having T2D according to measured HbA1c ≥ 6.5%, but only 344 (2.4%) after adjusting for putative nonglycemic HbA1c SNPs. The equivalent comparison in Europeans was 1.2% vs. 1.0%. Thus, reclassification of non-T2D individuals in Asian populations was much larger than in European populations, implying that the population impact

Figure 3. Reclassification of non-T2D (A) and undiagnosed T2D individuals (B) as T2D based on measured HbA1c and HbA1c adjusted for nonglycemic SNPs around the HbA1c threshold of 6.5% in European and Asian cohorts. A: Reclassification of non-T2D individuals, defined by $FG < 7 \text{ mmol/l}$, around the HbA1c threshold of 6.5%, attributable to nonglycemic SNPs. Reclassification was more evident in Asians, where 3.3% of non-T2D individuals were classified as T2D based on measured HbA1c, but only 2.4% after accounting for nonglycemic SNPs. The equivalent comparison in Europeans was minimal - 1.2% vs. 1.0%. **B:** Reclassification of undiagnosed T2D individuals, defined by $FG \geq 7 \text{ mmol/l}$ without treatment, around the HbA1c threshold of 6.5% attributable to nonglycemic SNPs. Here, reclassification was subtle for both Asian and European cohorts. The diagrams were adapted from Soranzo *et al.* [17] (A) and Chen *et al.* [20] (B).

of putative nonglycemic genetic factors on reclassification around the HbA1c threshold of 6.5% may differ by ancestry. Similar analyses are needed in populations of non-European ancestry, such as Africans, where selective pressure by infectious diseases on RBC lifespan, polymorphism, or hemoglobin variants [44-46] may have leveraged HbA1c-influencing, RBC-related alleles to reach greater frequency.

Aggregating data from populations of different ancestry in large-sample, trans-ethnic meta-analyses of GWAS will provide power to detect additional HbA1c-related genomic loci, allowing a deeper examination of the transferability of HbA1c-related genetic variants to different ethnicities [21]. It is noteworthy that an investigation in African Americans has shown that the relative contribution of demographic and metabolic factors to measured HbA1c is considerably larger than the contribution of genetic ancestry. Adjusting for genetic ancestry, after accounting for fasting glucose, has minimal impact on T2D classification by HbA1c, suggesting that ethnicity-specific, diagnostic thresholds are not warranted for population screening [47]. Nevertheless, in populations with a higher prevalence of nonglycemic, HbA1c-altering alleles, employing genotype-specific, diagnostic

thresholds, or calibrating measured HbA1c according to an individual's genotype profile to account for the effect of nonglycemic, HbA1c-altering alleles, may improve HbA1c's ability to detect undiagnosed T2D cases. Alternatively, glucose measurements, and not HbA1c, can be used to screen individuals with many nonglycemic HbA1c-related variants for T2D, similar to the recommended procedure for individuals with renal failure (carbamylation of hemoglobin), known hemoglobinopathies [14, 15], and other serum protein variants or chemical modifications to hemoglobin that interfere with lab assays. Notably, standardization of lab assay methods across populations is crucial for:

- Estimating ethnic differences in HbA1c levels.
- Making proper inferences about shifts in the distribution of HbA1c over time.
- Estimating trends in the prevalence and control of HbA1c-defined T2D for health care policy and planning [48].

Through GWAS on HbA1c, we may be able to implicate genes that are associated with such lab assay interferences, enabling the development of newer methods to correct such inaccuracies.

4. Implications for T2D risk stratification and prediction

The ADA considers HbA1c levels between 5.7 and 6.4% as “prediabetes”, which is supposed to signal an increased risk for future T2D [3]. Indeed, individuals with moderately elevated HbA1c levels have higher incident rates of T2D over time [49-60], supporting the use of HbA1c for risk stratification in the general population. HbA1c is also able to predict future T2D in various non-European populations around the world [9, 10, 51, 53-59, 61]. In the 1990s, the Diabetes Control and Complications Trial (DCCT) [62] and United Kingdom Prospective Diabetes Study (UKPDS) [63] established clear relationships between HbA1c levels and risk of diabetes-related endpoints in both type 1 diabetes and T2D patients. However, technical variation in assay methods initially prevented the optimal use of HbA1c in clinical practice. Therefore, in 1996, the National Glycohemoglobin Standardization Program (NGSP) initiated the harmonization of HbA1c testing to standardize HbA1c results to those used in DCCT and UKPDS [64].

While implementing large-scale measures to promote healthy living in the entire community would be ideal, concentrating T2D prevention efforts on at-risk individuals may be a more cost-effective and feasible strategy. HbA1c thresholds to detect those at-risk for future T2D are, however, not consistent worldwide. The International Expert Committee of the World Health Organization recommended that individuals with HbA1c between 6.0 and 6.5% were at elevated T2D risk and may be considered for T2D prevention strategies [65]. Likewise, the Canadian Diabetes Association chose the cut-point 6.0% to define prediabetes [66], whereas the ADA opted for a lower threshold of 5.7% [3]. As ethnocultural factors, lifestyle/behavioral factors, and other non-genetic factors may modify the predictive value of HbA1c, it is unknown whether these thresholds can or should be uniformly applied in all populations around the world. A cross-sectional study of 4,325 nondiabetic participants in Harbin, China, examined the agreement between HbA1c and oral glucose tolerance for diagnosing T2D and prediabetes [67]. The authors concluded that agreement between HbA1c and OGTT was weaker among obese participants, suggesting that the HbA1c threshold for prediabetes may need to be raised for an obese population.

Cross-sectional examinations in NHANES from 2005 to 2010 indicated that 12.4% of the U.S.

population had HbA1c levels between 5.7 and 6.4% [48]. Excluding 9.6% of the U.S. population with T2D or HbA1c $\geq 6.5\%$, 13.7% (i.e., one in seven nondiabetic U.S. adults) would carry the label “prediabetes”. To yield the greatest impact on T2D risk reduction from prevention strategies, interventions should be targeted only at those who are truly at elevated risk for future T2D. Carriers of putative nonglycemic HbA1c-raising alleles with a mildly elevated HbA1c level may fall into this prediabetes category, despite having completely normal glycemia. These individuals would be less likely to benefit from prevention strategies compared to their counterparts with an elevated HbA1c from dysglycemia. Conversely, carriers of putative nonglycemic HbA1c-lowering alleles may be wrongly categorized as “low risk” and “not qualified” for such intervention programs. Population-based cohort studies that test HbA1c as a predictor for T2D, stratified by genotype, race/ethnicity, and other non-genetic factors, may provide better risk estimates in subpopulations.

HbA1c-related genetic variants can be used as genetic instruments in Mendelian randomization (MR) analyses to strengthen causal inferences for the relationship between HbA1c and disease outcomes, particularly T2D-related complications. Studies have shown that HbA1c, even in the nondiabetic range, is associated with incident cardiovascular risk [68, 69]. The association between HbA1c and microvascular complications, retinopathy, and chronic kidney disease is more compelling among individuals with T2D [70, 71]. In fact, the HbA1c cut-point for T2D diagnosis was selected based on epidemiologic evidence from cross-sectional data on different populations showing an abrupt increase in the prevalence of retinopathy above the HbA1c cut-point of 6.5% [3, 66, 71-73]. Nevertheless, some of these observational associations may be partially attributed to confounding or reverse causations. Importantly, randomized controlled trials on intensive vs. conventional therapy in diabetes patients showed that targeting a near-normal HbA1c protected against microvascular disease [74], but not necessarily macrovascular disease [75-80]. Instrumentation in an MR approach can be applied to strengthen or refute a causal relationship between HbA1c and T2D-related complications, supporting the use of HbA1c as a biomarker of disease susceptibility [81].

Applying MR methods for estimating causal associations has limitations and challenges specific to HbA1c. As HbA1c-related common genetic variants explain individually little of the variance in HbA1c, combining multiple variants in a single al-

allelic score for instrumenting HbA1c could reduce the bias from weak instruments and improve the power to detect associations [82-84]. While MR analyses may strengthen causal inference, they do not delineate biological pathways from HbA1c, or hyperglycemia, to outcomes. Accounting for pleiotropy can be challenging as biological functions of some of the HbA1c-related genetic variants are still unknown. Excluding genetic variants that contribute to heterogeneity of effects ensures that estimates from multiple instruments are all similar, but does not specifically evaluate pleiotropy nor take into consideration that instruments may affect different aspects of the intermediate phenotype which in turn have differing effects on disease [85].

A pleiotropy assessment could be performed by excluding HbA1c-related variants that are also correlated with other biomarkers or clinical measures (e.g. body mass index). However, such an approach may be overly conservative as these phenotypes may be intermediates in the causal pathways from hyperglycemia or HbA1c to T2D-related outcomes. Alternatively, carefully selecting genetic variants that are biologically relevant to a specific pathway may produce MR estimates that are more mechanistically interpretable. Notably, when measuring HbA1c, we are agnostic to the underlying glycemic and non-glycemic pathways influencing HbA1c. Therefore, allelic scores, composed of all known HbA1c-related variants that explain more of the variance, may arguably be better for

estimating the overall effect of HbA1c on disease risk, even if non-glycemic and other pleiotropic mechanisms partly account for the association. To test the robustness of MR results, sensitivity analyses can be performed using a recently developed MR method adapted from Egger regression (MR-Egger), which detects and explains directional pleiotropy in estimating causal factors [86].

5. Conclusions

In summary, HbA1c genetics can unveil new biological mechanisms in glucose metabolism, regulation, and perturbation, and T2D susceptibility. Given that HbA1c is a surrogate for overall ambient glycemia, HbA1c genetics provide complementary information to findings from genetic investigations on point-in-time measurements of glycemic traits such as FG.

Nonglycemic HbA1c genetic variants have important implications in clinical practice and public health, specifically HbA1c thresholds for T2D diagnosis, risk stratification, prediction, and the accuracy of lab assays.

The population impact of nonglycemic HbA1c genetic variants on measured HbA1c in screening for undiagnosed T2D may vary by ancestry. HbA1c genetics has challenged the traditional perspective of using HbA1c as a “one-size-fit-all” screening tool, and motivates research on individualized prevention strategies in this era of precision medicine.

Disclosures: The authors report no conflict of interests.

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