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Rat Model of Chronic Midthoracic Lateral Hemisection

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Abstract

Although most spinal cord injuries (SCI) are anatomically incomplete, only limited functional recovery has been observed in people and rats with partial lesions. To address why surviving fibers cannot mediate more complete recovery, it is important to evaluate the physiological and anatomical status of spared fibers. These experiments require use of animal models. Here we describe a midthoracic unilateral spinal cord hemisection (HX; corresponds to Brown-Sequard lesion in humans) in adult rats. This is a useful animal model for partial injuries because there is a clear lesion of one entire side of the cord with intact fibers remaining on the contralateral side. This model allows the study and comparison of how acute and chronic trauma affect function of the surviving fibers.

Materials and Reagents

1. Adult (~210 g) female Sprague-Dawley rats
2. 1.5% isoflurane
3. Heated workstation with gas evacuation system and a face mask for induction and maintenance of anesthesia (ProStation Kit) (MIP/Anesthesia Technology, catalog number: AS-01-0491)
4. Antibiotic (Baytril)
5. Analgesic (Buprenorphine)
6. Sterile lactated Ringer solution
7. Cotton swap
8. Buprenorphine
9. Petrolatum ophthalmic ointment (Dechra Veterinary Products)
10. 4-0 monocryl (Ethicon)
11. Wound clips
12. Anatomical tracers

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Equipment

1. Isoflurane induction chamber (1 L)
2. Water circulating heating pad
3. Surgical microscope
4. Iridectomy scissors, faucets, blades, other tools for small animal surgery

Procedure

Note: All procedures were performed on adult female Sprague Dawley rats (~200 g) in compliance with the Institutional Animal Care and Use Committee at SUNY-Stony Brook and Northport VAMC.

1. After pre-training on the behavioral tasks, rats were deeply anesthetized with 3% isoflurane in 100% O₂ in an induction chamber (1 L).
2. Anesthesia was maintained by administering 1.5% isoflurane in 100% O₂ through a face mask. A water circulating heating pad was used to maintain body temperature at 36.5–37 °C during surgeries.
3. Before surgery, animals received a subcutaneous injection of analgesic Buprenorphine (0.01 mg/kg) to reduce post-operative pain. Petrolatum ophthalmic ointment was applied to the eyes to prevent desiccation.
4. Dorsal laminectomy (*i.e.* partial vertebral laminectomy) was performed to expose T10 spinal segment. The spinal level was confirmed by using a vein at T5-T6 as a landmark. The meningeal layer at T10 was slit (1 mm) at the midline longitudinally.
5. A complete transection of the left hemicord at T10 was carried out with the tip of iridectomy scissors, as follows:
 - a. First, while holding the dura and lifting the spinal cord slightly, one tip of the scissors was passed through the entire thickness of the spinal cord dorsal to ventral at the midline;
 - b. The left dorsal and ventral columns were then cut from lateral to the midline by closing other tip of scissors;
 - c. Finally, while keeping the cord elevated, one tip of the scissors was placed under the ventral surface of the spinal cord (up to the midline) and any uncut tissue in the left dorsal and ventral columns was cut ventral to dorsal up to the midline.
6. After surgery, the muscles were closed with 4-0 monocryl suture and skin was closed with wound clips.
7. Antibiotic (5 mg/kg, sc) and 5 ml of lactated Ringer's solution were administered subcutaneously. Bladder function was not compromised by this surgery. Injections

of antibiotic, analgesic and Ringer's solution were administered for 3 days post injury.

8. Horizontal or transverse sections of the spinal cord were used for reconstruction of injury (Figure 1).
9. As a result of HX SCI, there are clear behavioral impairments revealed by challenging motor tasks and automated Catwalk gait analysis; electrophysiological experiments allow evaluation of the conduction through fibers contralateral to the lesion and the possibility of establishing a functional detour around the lesion following administration of various treatments; moreover, unilateral injections of the anatomical tracers permit visualization of anterogradely labeled midline crossing fibers and retrogradely labeled neurons (Arvanian *et al.*, 2009; Hunanyan *et al.*, 2010; Schnell *et al.*, 2011; Garcia-alias *et al.*, 2011; Hunanyan *et al.*, 2011; Petrosyan *et al.*, 2013).

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Figure 1. Images of spinal cord following dorsal laminectomy and lateral HX lesion of the spinal cord

A Image of the exposed spinal cord (at arrow) following a dorsal laminectomy procedure. B Image of rat brain and spinal cord isolated from the rat that received HX spinal cord injury (at arrow) 6 weeks earlier.

Figure 2. Lateral HX spinal cord injury

A Horizontal section of the rat spinal cord prepared immediately after HX. B Transverse section of the cord at SCI epicenter prepared 6 weeks after HX; highlighted is area of spared white matter. Scale bar, 100 μ m. (*Adopted and modified from Garcia-alias et al., 2011*).