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An organocatalytic strategy for the stereoselective synthesis of C-galactosides with fluorine at the pseudoanomeric carbon

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Abstract

The α -fluorination of α - and β -C-ethanals of galactose using Jørgensen catalysts and NFSI was investigated. The crude reaction products were transformed to their primary alcohol or methylenated derivatives, which are versatile precursors to biologically interesting fluorinated glycomimetics. The α -C-glycoside substrate gave moderate to high yields of fluorinated α -C-glycosides with minor amounts of β -C-glycoside analogues. The reactions on the β -C-glycoside were lower yielding but gave exclusively fluorinated β -C-glycosides. For both α - and β -C-glycoside substrates (*R*) and (*S*) catalyst showed complementary stereoselectivity. The preparation of difluorinated materials required the use of racemic catalyst as enantiomerically pure catalyst gave intractable mixtures of products. These results are in line with the results for simple achiral aldehydes, and suggest that stereochemistry in the reactions of these chiral, highly substituted, carbohydrate-derived aldehydes are controlled primarily by the chirality in the catalyst.

Graphical Abstract

The fluorination of α - and β -C-linked-galactosyl ethanals using NFSI and Jørgensen catalyst, followed by reduction or methylenation of the derived aldehydes provided versatile precursors to fluorinated glycomimetics. The (*R*) and (*S*) catalysts gave stereochemically complementary results.

[†]Electronic Supplementary Information (ESI) available: Copies of ¹H, ¹³C, 2D NMR spectra and tabulated peak assignments for **0** and **00** – **00**, X-ray data for **00**.. See DOI: 10.1039/x0xx00000x

Introduction

Fluorination of bioactive materials is an active area of research in medicinal chemistry because the electronic and conformational effects of fluorine and the stability of the C-F bond may enhance binding in receptor sites and improve drug absorption, distribution, metabolism and excretion (ADME).^{1,2,3} Accordingly a variety of strategies for introduction of fluorine into biomolecules have been developed.^{4,5,6} However, while these methods have met much success in simple substrates, their application to more highly functionalized substrates like carbohydrates is often problematic.^{7,8,9} In this context, fluorinated C-glycosides like **2** and versatile derivatives thereof that can be transformed to complex analogues like **3** and **4**, are of interest (Fig. 1).^{10,11,12,13,14,15,16,17,18,19,20,21,22,23,24} The organocatalytic α -fluorination of simple C-glycoside aldehydes **1** is an appealing route to such precursors because of the mild reaction conditions, operational simplicity and easy access to reaction substrates.^{25,26,27} Herein, we describe the application of this strategy to C-glycosides of galactose. While the proline catalyzed amination of related C-glycoside aldehydes has been studied, to the best of our knowledge the mechanistically similar α -fluorination reaction has not been reported.²⁸

Results and Discussion

The substrates for this study were the α - and β -C-linked ethanals **5a** and **5b**, which were obtained from their readily available C-allyl derivatives.^{20,21,29} Initial experiments were performed on **5a** (Scheme 1). This material was treated with two molar equivalents of NFSI in THF in the presence of 2.5 mol % of (*S*) or (*R*) Jørgensen catalyst.²⁵ Because of the instability of the fluorinated aldehyde products, the crude aldehyde was treated with NaBH₄ or methylenetriphenylphosphorane to give the primary alcohol or terminal alkene derivative.³⁰ NaBH₄ reduction of the product from the fluorination with the (*S*) catalyst afforded α -C-monofluorinated glycoside **6**, and a β -C-glycoside **7** of undetermined configuration at the fluorinated carbon, in 75 and 10% yield respectively from **5a**. The fluorination/reduction sequence with the (*R*) catalyst produced the α -C-fluorohydrin **8** as the major product, together with **6**, the β -C-fluorohydrin **9** that was epimeric to **7** at the fluorinated carbon, and the α -C-difluorinated product **10** in 68, 6, 3 and 20% yield respectively.

Wittig methylenation on the crude fluorination product with the (*S*) catalyst gave the α -C-alkene **11** and the β -C-product **12** in 73 and 9% yields, respectively (Scheme 2). The fluorination/methylenation sequence with the (*R*) catalyst afforded **13** (53%), the α -C-glycoside that was epimeric to **11** at the fluorinated carbon, and an intractable mixture of other fluorinated alkenes (31%).

Towards more stereoselective syntheses of the β -C-fluorohydrins **7** and **9**, the fluorination-reduction protocol on the β -C-ethanal **5b**, was next investigated (Scheme 3). For optimal results, these reactions were performed at higher catalyst loading and shorter reaction times, compared to the reactions on **5a**. With 30 mol% catalyst **7** and **9** were obtained as the only observed mono-fluorinated products with the (*S*) and (*R*) catalysts, in 44 and 50 % yields, respectively.

The synthesis of difluorinated derivatives was next examined more carefully (Scheme 4). Reactions on **5a** using excess NFSI, extended reaction times and increased loading of the (*R*) or (*S*) catalyst, gave complex mixtures with low yields of fluorinated materials and other unidentified products. Experiments with pyrrolidine instead of the chiral catalyst gave similar results. However when the reaction was performed with a 20 mol% of a racemic mixture of the catalyst, and the crude product treated with NaBH₄, or subjected to methylenation, the difluorinated products **10** and **14** were obtained in 83 and 40% yield, respectively. The alkene **14** was also prepared by oxidation of a purified sample of **10** with Dess-Martin periodinane followed by Wittig olefination on the crude aldehyde. The overall yield of **14** from **10** following this procedure was 66%. As for the mono-fluorination reactions, the difluorination-reduction protocol on **5b** was lower yielding compared to case of **5a**, giving a 16% yield of the difluorinated β-C-glycoside **15** with 30 mol% of the racemic catalyst.

Stereochemical assignment of the fluorinated compounds rests on X-ray analysis of alkene **11**, $J_{H,H}$ values and chemical correlation between alkene and alcohol derivatives (Fig. 2, Supporting Information, Scheme 2). The X-ray structure of **11** indicated the α-C1 orientation and the *R* configuration at the fluorinated carbon. Interestingly, the molecule adopted the ¹C₄ chair conformation. A similar result has been reported for a related mono-fluorinated α-C-galactoside. A $J_{1,2}$ of 2.4 Hz also supported the α-C1 orientation, but is ambiguous in relation to assignment of ring conformation. Similarly $J_{1,2}$ values between 0 – 4.5 and close to 10 Hz were consistent with the α-C1 in **6**, **8**, **10**, **13**, **14**, and a β-C1 in **7**, **9**, **12** and **15** respectively. The configuration at the fluorinated carbon in the α-C-glycosides **6**, **8**, **11** and **13** rests on correlation to **11**. Thus, alkene **13** was deduced as *S* on the basis of its epimeric relationship to **11**. The α-C- fluorohydrins **6** and **8** were assigned by correlating them to **11** and **13** respectively, through an ozonolysis/aldehyde reduction sequence. The configurations at the fluorinated carbons in the β-C-glycosides **7**, **9** and **12** were tentatively assigned using the stereochemical model proposed for the Jørgensen catalyst (*vide infra*). That the (*S*) v.s. (*R*) catalyzed reactions selectively gave epimeric fluorides, **6** vs. **8**, **7** vs. **9**, **11** vs. **13** as the major products provides a measure of self-consistency to this argument.

The stereoselectivity in the fluorination of α-C-ethanal **5a** is in agreement with the model proposed for the Jørgensen catalysts (Scheme 5). Thus, the (*S*) and (*R*) catalyst lead to *E* enamines **16** and **20**, which should favour fluorination on the *Re* and *Si* face respectively, which is in line with the experimental result, stereoselective formation of **6** and **8**. Similarly high stereoselectivities have been observed in the proline catalysed amination on related carbohydrate aldehydes. The apparently better performance of the (*S*) catalyst compared to the (*R*), on the α-C-glycoside substrate may represent a “matched/mismatched” scenario. Thus **16** and **20** are expected to adopt the “exo-anomeric” conformation with respect to the pseudoglyconic torsion (i.e. the aminated carbon of the enamine is gauche to O5 and H1 of the sugar ring).³¹ The facial selectivity imposed by the (*R*) catalyst (i.e. on **20**), is opposed by steric hindrance from O5 of the sugar. For **16**, the enamine from the (*S*) catalyst, the facial selectivity of the catalyst is matched to that imposed by the sugar residue. Therefore, fluorination on **20** may be less stereoselective and side reactions more likely compared to **16**.

The results for the difluorination reactions also appear to be consistent with the Jørgensen model. Thus, for a given enantiomer of the catalyst, α -deprotonation on the mono-fluorination iminium ion (i.e. **17**, **21**), is sterically hindered by the bis-arylcarbinol side chain in the catalyst residue.²⁵ A racemic mixture of catalyst allows for amine exchange on **17** and **21** to give diastereomeric iminium ions in which α -deprotonation is not sterically blocked, thereby facilitating formation of the di-fluorinated product. As in the initial fluorination step, conformational effects may also impact on difluorination and explain why the (S) catalyst apparently shows a lesser propensity for difluorination compared to the (R) catalyst. Thus, “exo-anomeric” conformations for **17** and **21**, suggests that deprotonation in **17** (the favored mono-fluorinated iminium ion with the (S) catalyst), is hindered by both the sugar ring and the catalyst moiety, whereas the hindrance to deprotonation on **21**, the corresponding species with the (R) catalyst, comes from the catalyst moiety only.

Without definitive assignment of the configuration at the fluorinated carbons in the β -C-glycosides, it is impossible to tell whether the Jørgensen model extends to the fluorination of **5b**. However, the finding that enantiomeric catalysts exhibit opposite stereoselectivity in the fluorination of **5b**, supports this notion. This observation is also relevant to the mechanism by which the fluorinated β -glycosides **7** and **12** are formed from the α -C-glycoside **5a**. A pathway in which **5a** or **16** first epimerizes to **5b** or the β -C-glycoside of **16** respectively (via an retro-Michael-Michael mechanism), followed by fluorination, seems more likely than epimerization of a first formed α -C-fluorinated species.³² In the latter scenario, the individual reactions of **5a** with the enantiomeric catalysts might have been expected to give mixtures of the β -C-glycosides **7** and **12**.

Conclusions

The α -fluorination of C-linked galactosyl ethanals using the Jørgensen catalyst and NFSI is a promising route to versatile precursors for complex fluorinated glycomimetics. Enantiomeric catalysts influenced complementary facial selectivity in line with the results for simple achiral aldehydes. The high diastereoselectivity observed for these stereochemically complex substrates is similar to that observed for the organocatalyzed amination on related substrates, and point to the powerful stereodirecting effects of these proline-based catalysts in enamine reactions. These findings provide insight on the effects of conformation and remote substituents in such complex substrates, and provide the basis for the wider application of this chemistry in carbohydrate synthesis.

Experimental section

Solvents were purified by standard procedures or used from commercial sources as appropriate. Petroleum ether refers to the fraction of petroleum ether boiling between 40 and 60 °C. Ether refers to diethyl ether. Unless otherwise stated thin layer chromatography (TLC) was done on 0.25 mm thick precoated silica gel 60 (HF-254, Whatman) aluminium sheets and flash column chromatography (FCC) was performed using Kieselgel 60 (32–63 mesh, Scientific Adsorbents). Elution for FCC usually employed a stepwise solvent polarity gradient, correlated with TLC mobility. Chromatograms were observed under UV (short and long wavelength) light, and/or were visualized by heating plates that were dipped in a

solution of ammonium (VI) molybdate tetrahydrate (12.5 g) and cerium (IV) sulfate tetrahydrate (5.0 g) in 10% aqueous sulphuric acid (500 mL), or a solution of 20% sulfuric acid in ethanol. ^1H and ^{13}C and ^{19}F NMR spectra were recorded using Bruker Ultra Shield 400 MHz, Bruker Ultra Shield 500 MHz and Bruker Ultra Shield Plus 600 MHz instruments, in CDCl_3 or C_6D_6 solutions with residual CHCl_3 or C_6H_6 as internal standard (δ_{H} 7.27, 7.16 and δ_{C} 77.2, 128.4 ppm). Chemical shifts are quoted in ppm relative to tetramethylsilane (δ_{H} 0.00) and coupling constants (J) are given in Hertz. First order approximations are employed throughout. High resolution mass spectrometry was performed on an Ultima Micromass Q-TOF or Waters Micromass LCT Premier mass spectrometers.

General procedure for fluorination-reduction sequence on aldehydes: 2-(2', 3', 4', 6'-Tetra-O-benzyl- α -D-galactopyranosyl)-2-*R*-fluoroethanol (6) and 2-(2', 3', 4', 6'-Tetra-O-benzyl- β -D-galactopyranosyl)-2-*R*-fluoroethanol (7)

To a solution of aldehyde **5a** (250 mg, 0.44 mmol) in THF (2.2 mL) was added (*S*)-Jørgensen catalyst (7.0 mg, 0.012 mmol). The mixture was stirred at rt for 15 min at which time NFSI (275 mg, 0.87 mmol) was added, and stirring continued at rt for 6 h, or after NMR indicated disappearance of the starting material. The reaction mixture was then concentrated at rt under reduced pressure and the residue filtered over a short column of silica gel. The filtrate was evaporated *in vacuo* at rt, the residue taken up in methanol (10 mL) and NaBH_4 (250 mg, 6.8 mmol) slowly added at 0 °C to the mixture. The reaction was stirred for 30 min, then pH adjusted to 6 with 1M HCl, and the volatiles removed under reduced pressure at rt. The residue was extracted with EtOAc and the organic phase washed with saturated aqueous NaHCO_3 , and brine, dried (Na_2SO_4), and evaporated *in vacuo*. FCC of the crude product gave **6** (195 mg, 75%) and **7** (27 mg, 10%).

For **6**: white amorphous solid, m.p = 79–83 °C; R_f 0.50 (60% hexanes: 40% EtOAc); $[\alpha]_{\text{D}}^{25} + 40$ (c 1.6, CH_2Cl_2); ^1H NMR (500 MHz, CDCl_3) δ 7.38–7.16 (m, 20H), 4.65 (ddt, $J = 2.8, 8.9, 48$ Hz, 1H), 4.58–4.42 (m, 8H), 4.43 (m, partially buried, 1H), 4.33 (m, 1H), 4.20 (t, $J = 8.1$ Hz, 1H), 3.98 (dd, $J = 3.2, 6.3$ Hz, 1H), 3.95–3.78 (m, 2H), 3.75 (t, $J = 3.5$ Hz, 1H), 3.69 (m, 1H), 3.60 (dd, $J = 3.1, 11.6$ Hz, 1H), 2.91 (br s, 1H); ^{13}C NMR (125 MHz, CDCl_3) δ 138.3, 138.2, 138.0, 137.8, 128.7, 128.6, 128.4, 128.3, 128.1, 128.0, 127.9, 127.8, 90.4 (d, $J_{\text{C-F}} = 169$ Hz), 74.2, 73.7, 73.5, 73.4 (two peaks), 73.3, 73.2, 71.7, 65.5, 65.2 (d, $J_{\text{C-F}} = 32$ Hz), 63.1 (d, $J_{\text{C-F}} = 21$ Hz); ^{19}F NMR (377 MHz, CDCl_3) δ –198.5 (s); HRMS (ESI, MNa^+) m/z calc for $\text{C}_{36}\text{H}_{39}\text{FNaO}_6$ 609.2628, found 609.2618.

For **7**: colorless oil; $R_f = 0.28$ (60% hexanes: 40% EtOAc); $[\alpha]_{\text{D}}^{25} + 24$ (c 0.7, CH_2Cl_2); ^1H NMR (600 MHz, CDCl_3) δ 7.32–7.22 (m, 20H), 5.00 (m, 2H), 4.91 (dt, $J = 3.8, 51.7$ Hz, 1H), 4.78 (m, 1H), 4.82–4.65 (m, 4H), 4.45 (ABq, $J = 11.6$ Hz, $\delta = 0.06$ ppm, 2H), 4.19 (t, $J = 9.6$ Hz, 1H), 4.05 (ddd, $J = 6.2, 12.6, 19.6$ Hz, 1H), 3.97 (bs, 1H), 3.85 (m, 1H), 3.66 (dd, $J = 2.2, 9.5$ Hz, 1H), 3.61 (dd, $J = 6.3, 8.7$ Hz, 1H), 3.58 (t, $J = 6.1$ Hz, 1H), 3.48 (dd, $J = 5.8, 8.8$ Hz, 1H), 3.43 (dd, $J = 9.2, 28.4$ Hz, 1H), 2.64 (m, 1H); ^{13}C NMR (150 Mz, CDCl_3) δ 138.5, 138.2, 138.1, 137.7, 128.7, 128.4, 128.3, 128.2, 128.1, 127.9, 127.8, 127.7, 90.0 (d, $J_{\text{C-F}} = 178$ Hz), 84.6, 79.2 (d, $J_{\text{C-F}} = 19$ Hz), 77.8, 75.8, 74.4, 74.3 (d, $J_{\text{C-F}} =$

4.4 Hz), 73.8, 72.9, 72.4, 69.3, 62.9 (d, J_{C-F} = 23 Hz); ^{19}F NMR (377 MHz, CDCl_3) δ -206.9 (s); HRMS (ESI, MNa^+) m/z calc for $\text{C}_{36}\text{H}_{39}\text{FNaO}_6$ 609.2628, found 609.2625

2-(2', 3', 4', 6'-Tetra-O-benzyl- α -D-galactopyranosyl)-2-S-fluoroethanol (8), 2-(2', 3', 4', 6'-Tetra-O-benzyl- β -D-galactopyranosyl)-2-R-fluoroethanol (9) and 2-(2', 3', 4', 6'-Tetra-O-benzyl- α -D-galactopyranosyl)-2,2-difluoroethanol (10)

Aldehyde **5a** (150 mg, 0.265 mmol) was subjected to the standard fluorination-reduction procedure with (*R*)-Jørgensen catalyst, as described for the synthesis of **6** and **7**. FCC of the product after the reduction step afforded **6** (5 mg, 3%), **8** (105 mg, 68%), **9** (9 mg, 6%) and **10** (31 mg, 20%).

For **8**: colorless oil; R_f = 0.38 (40% EtOAc/60% hexanes); $[\alpha]_{\text{D}}^{25} +39$ (c 1.3, CH_2Cl_2); ^1H NMR (500 MHz, CDCl_3) δ 7.27–7.18 (m, 20H), 4.87 (dq, J = 4.3, 46.7 Hz, 1H), 4.76 (app d, J = 12.0 Hz, 1H), 4.71 (app d, J = 10.5 Hz, 1H), 4.67 (s, 2H), 4.53 (app d, J = 11.5 Hz, 1H), 4.48 (app d, J = 11.5 Hz, 1H), 4.40 (ABq, J = 12.0 Hz, δ = 0.08 ppm, 2H), 4.14 (m, 1H), 4.05 (br t, J = 7.1 Hz, 1H), 3.97 (ddd, J = 3.0, 5.9, 34.1 Hz, 1H), 3.96 (dd, J = 2.0, 9.5 Hz 1H), 3.84 (t, J = 2.6 Hz, 1H), 3.81–3.76 (m, 2H), 3.59 (t, J = 9.5 Hz, 1H), 3.31 (dd, J = 4.4, 10.2 Hz, 1H), 2.43 (br s, 1H); ^{13}C NMR (125 MHz, CDCl_3) δ 138.7, 138.6, 138.3, 138.2, 128.7, 128.6, 128.5, 128.3, 128.1, 128.0, 127.8, 93.5 (d, J_{C-F} = 178 Hz), 79.1, 75.7, 75.0 (d, J_{C-F} = 4 Hz), 74.6, 74.2, 73.5 (d, J_{C-F} = 35 Hz), 69.2, 63.1 (d, J_{C-F} = 25 Hz); ^{19}F NMR (377 MHz, CDCl_3) δ -198.5 (s); HRMS (ESI, MNa^+) m/z calc for $\text{C}_{36}\text{H}_{39}\text{FNaO}_6$ 609.2628, found 609.2618.

For **9**: colorless oil; R_f = 0.48 (40% EtOAc/60% hexanes); $[\alpha]_{\text{D}}^{25} +32$ (c 0.1, CH_2Cl_2); ^1H NMR (500 MHz, C_6D_6) δ 7.03–7.38 (m, 20H), 4.94 (m, 2H), 4.88 (dt, J = 4.5, 45 Hz, 1H partially buried under m at 4.94 ppm), 4.55 (m, 2H), 4.30 (ABq, J = 11.8 Hz, δ = 0.06 ppm, 2H), 4.24 (ABq, J = 11.8 Hz, δ = 0.05, 2H), 4.08 (t, J = 9.6 Hz, 1H), 3.85 (m, 3H), 3.70 (ddd, J = 1.8, 10.0, 11.8 Hz, 1H), 3.63 (t, J = 8.1 Hz, 1H), 3.52 (dd, J = 5.4, 8.9 Hz, 1H), 3.36 (m, 1H), 3.33 (dd, J = 2.7, 8.2 Hz, 1H); ^{13}C NMR (125 MHz, C_6D_6) δ 139.7, 138.2, 139.2, 139.1, 129.0 (two peaks), 128.9, 128.5 128.3 (two peaks), 128.1, 128.0, 93.4 (d, J_{C-F} = 174 Hz), 85.7, 80.5 (d, J_{C-F} = 21 Hz), 77.5, 75.5 (d, J_{C-F} = 8 Hz), 75.4, 75.3, 74.1, 73.9, 72.4, 69.1, 62.3 (d, J_{C-F} = 25 Hz); ^{19}F NMR (377 MHz, CDCl_3) δ -196.5 (s); HRMS (ESI, MNa^+) m/z calc for $\text{C}_{36}\text{H}_{39}\text{FNaO}_6$ 609.2628, found 609.2626.

For **10**: colorless oil; R_f = 0.63 (40% EtOAc/60% hexanes); $[\alpha]_{\text{D}}^{26} +44$ (c 0.6, CH_2Cl_2); ^1H NMR (500 MHz, C_6D_6) δ 7.22–7.07 (m, 20H), 4.62 (ddd, J = 2.1, 8.9, 18.0 Hz, 1H), 4.52 (apparent d, J = 11.5 Hz, 1H), 4.47 (m, 1H), 4.38 (br m, 5H), 4.18 (ABq, J = 11.8 Hz, δ = .05 ppm, 1H), 4.10 (br m, 1H), 4.05 (dd, J = 2.2, 5.0 Hz, 1H), 3.99 (dd, J = 3.1, 5.3 Hz, 1H), 3.93 (bm, 1H), 3.80 (dd, J = 3.1, 5.0 Hz, 1H), 3.71 (dd, J = 2.9, 11.2 Hz, 1H), 3.27 (t, J = 7.5 Hz, 1H); ^{13}C NMR (125 MHz, C_6D_6) δ 139.0, 138.8, 138.7, 132.4, 132.2, 131.9, 131.7, 128.8, 128.1, 127.9, 127.8, 127.7, 127.6, 127.5, 127.4, 127.2, 125.6, 125.2, 123.6 (t, J_{C-F} = 243 Hz), 75.6, 75.0, 74.9, 74.3, 73.9, 73.8, 73.7, 72.6, 68.3 (dd, J_{C-F} = 25, 34 Hz), 66.6, 64.1 (dd, J_{C-F} = 28, 34 Hz); ^{19}F NMR (377 MHz, CDCl_3) δ -114.6 (ABq, J = 256 Hz, δ = 9.8 ppm, 2F); HRMS (ESI, MNa^+) m/z calc for $\text{C}_{36}\text{H}_{38}\text{F}_2\text{NaO}_6$ 627.2534, found 627.2523.

Synthesis of **7** from **5b**

Aldehyde **5b** (46 mg, 0.081 mmol) was subjected to the standard fluorination procedure with NFSI (51 mg, 0.16 mmol) and (*S*)-Jørgensen catalyst (15 mg, 0.024 mmol) for 1.5 h. Reduction of the crude product with NaBH₄ following the standard procedure and FCC of the product afforded **7** (21 mg, 44%).

Synthesis of **9** from **5b**

Aldehyde **5b** (46 mg, 0.081 mmol) was subjected to the standard fluorination procedure with NFSI (51 mg, 0.16 mmol) and (*R*)-Jørgensen catalyst (15 mg, 0.024 mmol) for 1.5 h. Reduction of the crude product with NaBH₄ following the standard procedure and FCC of the product afforded **9** (24 mg, 50%).

Synthesis of **10** using racemic catalyst

Aldehyde **5a** (330 mg, 0.58 mmol) was subjected to the fluorination procedure described for **6**, with NFSI (730 mg, 2.30 mmol), (*R*)-Jørgensen catalyst (35 mg, 0.058 mmol) and (*S*)-Jørgensen catalyst (35 mg, 0.058 mmol) for 18 h. Reduction of the crude product with NaBH₄ following the standard procedure and FCC of the product afforded **10** (290 mg, 83%).

General procedure for fluorination-Wittig methylenation sequence on aldehydes: **3-(2', 3', 4', 6'-Tetra-O-benzyl- α -D-galactopyranosyl)-3-*R*-fluoropropene (11)** and **3-(2', 3', 4', 6'-Tetra-O-benzyl- β -D-galactopyranosyl)-3-*R*-fluoropropene (12)**

To a solution of aldehyde **5a** (490 mg, 0.87 mmol) in THF (5.0 mL) was added (*S*)-Jørgensen catalyst (25 mg, 0.022 mmol). The mixture was stirred at rt for 15 min at which time NFSI (540 mg, 1.7 mmol) was added, and stirring continued at rt for 6 h. The reaction was monitored and processed as described for the synthesis of **6**. The crude fluoro-aldehyde was used directly in the next step. To a solution of Ph₃PCH₃Br (932 mg, 2.6 mmol) in anhydrous THF (10 mL) was added 2M NaHMDS in THF (1.3 mL, 2.6 mmol) at -78 °C. The resulting bright orange solution was stirred for 1 h at this temperature and then a solution of the product from the previous step in THF (10 mL) was added. The reaction mixture was warmed to rt over 2 h, then poured into saturated aqueous NH₄Cl and extracted with EtOAc. The organic phase was washed with brine, dried (Na₂SO₄), filtered, and evaporated under reduced pressure. FCC of the residue afforded **11** (423 mg, 73%) and **12** (51 mg, 9%).

For **11**: White needles (ether:methanol); m.p = 97–98 °C; *R_f* = 0.35 (20 % ethyl ether: 80% hexanes); [α]_D²⁵ +53 (c 0.5, CH₂Cl₂); ¹HNMR (500 MHz, CDCl₃) δ 7.27–7.10 (m, 20H), 6.05 (m, 1H), 5.43 (br d, *J* = 17.5 Hz, 1H), 5.28 (d, *J* = 11.0 Hz, 1H), 5.02 (ddd, *J* = 5.5, 7.0, 46.8 Hz, 1H), 4.40–4.59 (m, 8H), 4.33 (t, *J* = 7.2 Hz, 1H), 4.07 (dd, *J* = 8.5, 11.5 Hz, 1H), 3.99 (dd, *J* = 2.4, 7.0 Hz, 1H), 3.72 (m, 3H); ¹³CNMR (125 MHz, CDCl₃) δ 138.8, 138.5, 138.3, 138.0, 134.8 (d, *J*_{C-F} = 17 Hz), 128.6, 128.5, 128.4, 128.2, 127.9, 127.8, 127.7, 118.0 (d, *J*_{C-F} = 13 Hz), 90.2 (d, *J*_{C-F} = 168 Hz), 75.3, 74.7, 74.0, 73.6, 73.5, 73.3, 73.2, 71.8, 69.6 (d, *J*_{C-F} = 29 Hz), 65.8; ¹⁹FNMR (377 MHz, CDCl₃) δ -190.5 (s); HRMS (ESI, MNa⁺) *m/z* calc for C₃₇H₃₉FN₆O₆ 605.2679, found 605.2668.

For **12**: colorless oil; $R_f = 0.27$ (20 % ethyl ether: 80% hexanes); $[\alpha]_D^{26} + 30$ (c 1.1, CH_2Cl_2); (500 MHz, CDCl_3) δ 7.30–7.15 (m, 20H), 6.01 (m, 1H), 5.32 (ddd, $J = 1.0, 2.5, 17.0$ Hz, 1H), 5.21 (d, $J = 10.5$ Hz, 1H), 5.09 (dd, $J = 8.0, 47.0$ Hz, 1H), 4.91 (app d, $J = 10.5, 1\text{H}$), 4.87 (app d, $J = 11.5, 1\text{H}$), 4.70–4.55 (m, 4H), 4.37 (ABq, $J = 11.5, \delta = 0.05$ ppm, 2H), 4.08 (t, $J = 9.5$ Hz, 1H), 3.90 (d, $J = 2.0$ Hz, 1H), 3.53 (m, 3H), 3.46 (t, $J = 6.3$ Hz, 1H), 3.20 (ddd, $J = 1.7, 9.7, 27.7$ Hz, 1H); $^{13}\text{CNMR}$ (125 MHz, CDCl_3) δ 138.9, 138.5 (two peaks), 138.2, 133.6 (d, $J_{C-F} = 20$ Hz), 128.7, 128.6, 128.5, 128.4, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8, 127.7, 118.9 (d, $J_{C-F} = 12$ Hz), 90.7 (d, $J_{C-F} = 174$ Hz), 85.0, 80.9 (d, $J_{C-F} = 20$ Hz), 78.0, 75.7, 74.9 (d, $J_{C-F} = 4$ Hz), 74.6, 73.8, 73.5, 72.5, 69.2; $^{19}\text{FNMR}$ (377 MHz, CDCl_3) δ –193.7 (s); HRMS (ESI, MNa^+) m/z calc for $\text{C}_{37}\text{H}_{39}\text{FNaO}_6$ 605.2679, found 605.2679.

3-(2', 3', 4', 6'-Tetra-O-benzyl- α -D-galactopyranosyl)-3-(S)-fluoropropene (**13**)

Aldehyde **5a** (540 mg, 0.95 mmol) was subjected to the standard fluorination-Wittig methylenation procedure with (*R*)-Jørgensen catalyst, as described for the synthesis of **11** and **12**. FCC of the product after the methylenation step afforded **13** (309 mg, 53 %) and an unseparated mixture of fluorinated products (180 mg, ca 31%).

For **13**: colorless oil; $R_f = 0.06$ (CHCl_3); $[\alpha]_D^{25} + 19$ (c 5.5, CH_2Cl_2); $^1\text{HNMR}$ (500 MHz, CDCl_3) δ 7.25–7.17 (m, 20H), 5.82 (m, 1H), 5.30 (dt, $J = 1.6, 18.6$ Hz, 1H), 5.20 (d, $J = 10.7$ Hz, 1H), 5.19 (dt, $J = 5.0, 47.8$ Hz, 1H), 4.69–4.40 (m, 8H), 4.19 (m, 1H), 3.92 (m, 3H), 3.82 (dt, $J = 4.5, 29.0$ Hz, 1H), 3.70 (dd, $J = 7.2, 10.3$ Hz, 1H), 3.52 (dd, $J = 5.1, 10.3$ Hz, 1H); $^{13}\text{CNMR}$ (125 MHz, CDCl_3) δ 138.8, 138.7, 138.6, 138.4, 133.2 (d, $J_{C-F} = 19$ Hz), 128.7, 128.6, 128.5, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8, 127.7, 119.0 (d, $J_{C-F} = 11$ Hz), 94.1 (d, $J_{C-F} = 174$ Hz), 77.9, 76.2, 74.9, 74.5, 73.8, 73.6, 73.5, 73.4, 73.3 (d, $J_{C-F} = 14$ Hz), 73.2, 67.9; $^{19}\text{FNMR}$ (377 MHz, CDCl_3) δ –184.7 (s); HRMS (ESI, MNa^+) m/z calc for $\text{C}_{37}\text{H}_{39}\text{FNaO}_6$ 605.2679, found 605.2671.

3-(2', 3', 4', 6'-Tetra-O-benzyl- α -D-galactopyranosyl)-3,3-difluoropropene (**14**)

To a solution of aldehyde **5a** (5.30 g, 9.35 mmol) in THF (50 mL) was added (*S*)-Jørgensen (250 mg, 0.47 mmol) and (*R*)-Jørgensen catalyst (250 mg, 0.47 mmol). The mixture was stirred at rt for 15 min at which time NFSI (7.00 g, 22.4 mmol) was added, and stirring continued at rt for 12 h. The reaction mixture was concentrated at rt under reduced pressure and the residue filtered over a short column of silica gel. The filtrate was evaporated in *vacuo* at rt, and the crude aldehyde was used in the next step without further purification. To a solution of $\text{Ph}_3\text{PCH}_2\text{Br}$ (15.0 g, 37.4 mmol) in anhydrous THF (90 mL) was added 2M NaHMDS in THF (23 mL, 46.0 mmol). The resulting bright orange solution was stirred for 1 h at this temperature at which time a solution of the crude aldehyde in THF (30 mL) was added. The reaction mixture was warmed to rt over 2 h, then processed following the general procedure for the Wittig reaction, as described for the synthesis of **11** and **12**. FCC of the crude product afforded **14** (2.23 g, 40%): colorless oil; $R_f = 0.17$ (CHCl_3); $[\alpha]_D^{25} + 37$ (c 0.4, CH_2Cl_2); $^1\text{HNMR}$ (600 MHz, C_6D_6) δ 7.04–7.38 (m, 20H), 6.36 (m, 1H), 5.71 (bd, $J = 17.5$ Hz, 1H), 5.10 (d, $J = 11.2$ Hz, 1H), 4.57 (m, 1H), 4.35–4.50 (m, 6H), 4.31 (ABq, $J = 11.0$ Hz, $\delta = 0.03$ ppm, 2H), 4.22 (app d, $J = 11.5$ Hz, 1H), 4.20 (dd, $J = 7.9, 11.3$ Hz, 1H), 4.06 (dd, $J = 2.4, 5.3$ Hz, 1H), 4.00 (m, 1H), 3.97 (dd, $J = 3.5, 11.3$ Hz, 1H), 3.82 (dd, $J = 2.8, 4.4$

Hz, 1H); ^{13}C NMR (150 MHz, C_6D_6) δ 139.7, 139.2 (two peaks), 138.8, 132.9 (t, $J_{\text{C-F}} = 24$ Hz), 129.3, 129.2, 129.1, 129.0, 128.9, 128.8, 128.7, 128.5, 128.1, 128.0, 127.9, 127.8, 120.9 (t, $J_{\text{C-F}} = 242$ Hz), 119.3 (t, $J_{\text{C-F}} = 10$ Hz), 76.1 (two peaks), 76.0, 75.9, 74.5, 73.7 (two peaks), 72.9, 72.3 (t, $J_{\text{C-F}} = 31$ Hz), 67.0; ^{19}F NMR (377 MHz, CDCl_3) δ -103.2 (ABq, $J = 264$ Hz, $\delta = 7.3$ ppm, 2F); HRMS (ESI, MNa^+) m/z calc for $\text{C}_{37}\text{H}_{38}\text{F}_2\text{NaO}_6$ 623.2585, found 623.2571.

Synthesis of difluoroalkene **14** from alcohol **10**

To a solution of alcohol **10** (40 mg, 0.066 mmol) in dry DCM (1 mL) was added Dess-Martin periodinane solution (0.1 mL of a 15% wt/vol in DCM, 0.035 mmol). The reaction mixture was stirred at rt for 20 min, then concentrated at rt under reduced pressure. Next, the residue was filtered over a short column of silica gel, and the filtrate evaporated in *vacuo* at rt. The crude aldehyde product was used in the next step without further purification. To a solution of $\text{Ph}_3\text{PCH}_2\text{Br}$ (400 mg, 1.1 mmol) in anhydrous THF (1 mL) was added 2M NaHMDS in THF (0.7 mL, 1.4 mmol). The resulting bright orange solution was stirred for 1 h at this temperature at which time a solution of the crude aldehyde in THF (1.0 mL) was added. The reaction mixture was warmed to rt over 2 h, then processed following the standard procedure for Wittig reactions. FCC of the crude product afforded **14** (32 mg, 80%).

2-(2', 3', 4', 6'-Tetra-O-benzyl- β -D-galactopyranosyl)-2,2-difluoroethanol (**15**)

Aldehyde **5b** (65 mg, 0.115 mmol) was subjected to the fluorination procedure described for **6**, with NFSI (181 mg, 0.570 mmol), (*R*)-Jorgensen catalyst (10 mg, 0.017 mmol) and (*S*)-Jorgensen catalyst (10 mg, 0.017 mmol) for 18 h. Reduction of the crude product with NaBH_4 following the standard procedure and FCC of the product afforded **15** (11 mg, 16%): colorless oil; $R_f = 0.60$ (5% acetone: 95% CH_2Cl_2); $[\alpha]_{\text{D}}^{20} +20$ (c 0.1, CHCl_3); ^1H NMR (600 MHz, C_6D_6) δ 7.40–7.05 (m, 20H), 4.84 (ABq, $J = 10.7$ Hz, $\delta = 0.14$ ppm, 2H), 4.74 (ABq, $J = 11.6$ Hz, $\delta = 0.40$ ppm, 2H), 4.45 (t, $J = 9.4$ Hz, 1H), 4.41 (ABq, $J = 11.8$ Hz, $\delta = 0.02$ ppm, 2H), 4.38 (br m, 5H), 4.22 (ABq, $J = 11.8$ Hz, $\delta = 0.10$ ppm, 2H), 3.91 (br m, 1H), 3.78 (br m, 2H), 3.70 (d, $J = 1.90$ Hz, 1H), 3.52 (dd, $J = 6.3, 9.2$ Hz, 1H), 3.47 (dd, $J = 6.6, 9.2$ Hz, 1H), 3.34 (dd, $J = 2.8, 9.3$ Hz, 1H), 3.30 (t, $J = 6.5$ Hz, 1H), 2.15 (t, $J = 6.5$ Hz, 1H); ^{13}C NMR (150 MHz, C_6D_6) δ 139.6, 139.5, 139.2, 138.9, 130.0–127.0 (several lines buried under C_6D_6 triplet), 121.9 (t, $J_{\text{C-F}} = 245$ Hz), 85.2, 78.2 (t, $J_{\text{C-F}} = 28$ Hz), 77.9, 75.5, 75.3, 75.1 (d, $J_{\text{C-F}} = 3$ Hz), 74.3, 74.0, 73.0, 69.5, 64.2 (t, $J_{\text{C-F}} = 32$ Hz); ^{19}F NMR (377 MHz, CDCl_3) δ -114.3 (ABq, $J = 264$ Hz, $\delta = 6.6$ ppm, 2F); HRMS (ESI, MNa^+) m/z calc for $\text{C}_{36}\text{H}_{38}\text{F}_2\text{NaO}_6$ 627.2534, found 627.2521.

General procedure for correlation of allylic fluorides and fluorohydrin derivatives: Transformation of **11** to **6**

A stream of O_3/O_2 was bubbled through a solution of **11** (50 mg, 0.09 mmol) in DCM 20 mL at -78°C , for 5 min. PPh_3 (50 mg, 0.19 mol) was then added to the mixture and stirring continued at rt for an additional 2 h. The volatiles were evaporated in *vacuo*, and NaBH_4 (30 mg, 0.8 mmol) was then slowly added at 0°C to a solution of the residue in MeOH (2 mL).

The reaction mixture was processed following the standard procedure for NaBH₄ reduction. FCC of the crude product provided **6** (31 mg, 62%).

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Notes and references

1. Hagmann WK. *J Med Chem.* 2008; 51:4359–4369. [PubMed: 18570365]
2. O'Hagan D. *Chem Soc Rev.* 2008; 37:308–319. [PubMed: 18197347]
3. Purser S, Moore PR, Swallow S, Gouverneur V. *Chem Soc Rev.* 2008; 37:320–330. [PubMed: 18197348]
4. Champagne PA, Desroches J, Hamel J-D, Vandamme M, Paquin J-F. *Chem Rev.* 2015; 115 ASAP. doi:10.2102/cr500706a.
5. Yang X, Wu T, Phipps RJ, Toste DF. *Chem Rev.* 2015; 115:826–870. [PubMed: 25337896]
6. Valero G, Companyó X, Rios R. *Chem Eur J.* 2011; 17, 2018–2037.
7. Schedler D, Baker DC. *Carbohydr Res.* 2004; 339:1585–1595. [PubMed: 15183732]
8. Cheng Y, Guo A-L, Guo D-S. *Cur Org Chem.* 2010; 14:977–999.
9. Leclerc E, Pannecoucke X, Ethève-Queiquejeu M, Sollogoub M. *Chem Soc Rev.* 2013; 42:4270–4283. [PubMed: 23212149]
10. Plantier-Royon R, Portella C. *Carbohydr Res.* 2000; 327:119–146. [PubMed: 10968679]
11. Jiménez-Barbero J, Demange R, Schenk K, Vogel P. *J Org Chem.* 2001; 66:5132–5138. [PubMed: 11463266]
12. Marcotte S, D'Hooge F, Ramadas S, Feasson C, Pannecoucke X, Quirion J-C. *Tetrahedron Lett.* 2001; 42:5879–5882.
13. Picard J, Lubin-Germain JN, Uziei J, Augé J. *Synthesis.* 2006:979–982.
14. Tony KA, Denton RW, Dilhas A, Jiménez-Barbero J, Mootoo DR. *Org Lett.* 2007; 9:1441–1444. [PubMed: 17358072]
15. Pérez-Castells J, Hernández-Gay JJ, Denton RW, Tony KA, Mootoo DR, DR, Jiménez-Barbero J. *Org Biomol Chem.* 2007; 5:1087–1092. [PubMed: 17377662]
16. Poulain F, Serre A-L, Lalot J, Leclerc E, Quirion J-C. *J Org Chem.* 2008; 73:2435–2438. [PubMed: 18275216]
17. Colombel S, Van Hijfte N, Poisson T, Leclerc E, Pannecoucke X. *Chem Eur J.* 2013; 19:12778–12787. [PubMed: 23955898]
18. Colombel S, Van Hijfte N, Poisson T, Pannecoucke X, Monneaux F, Leclerc E. *J Fluorine Chem.* 2015; 173:84–91.
19. For examples of general strategies for transforming C- glycoside alkenes and aldehydes to more complex C-glycosides, Gustafsson T, Saxin M, Kihlberg J. *J Org Chem.* 2003; 68:2506–2509. [PubMed: 12636428] , and ref. 19–23.
20. Wipf P, Pierce JG. *Org Lett.* 2006; 8:3375–3378. [PubMed: 16836409]
21. Pu J, Franck RW. *Tetrahedron.* 2008; 64:8618–8629. [PubMed: 19746154]

22. Thota VN, Gervay-Hague J, Kulkarni SS. *Org Biomol Chem*. 2012; 10:8132–8139. [PubMed: 22961309]
23. Thota VN, Brahmaiah M, Kulkarni SS. *J Org Chem*. 2013; 78:12082–12089. [PubMed: 24199837]
24. Altiti AS, Mootoo DR. *Org Lett*. 2014; 16:1466–1469. [PubMed: 24559301]
25. Marigo M, Fielenbach D, Braunton A, Kjærsgaard A, Jørgensen KA. *Angew Chem Int Ed*. 2005; 44:3703–3706.
26. Steiner DD, Mase N, Barbas CF III. *Angew Chem Int Ed*. 2005; 44:3706–3710.
27. Beeson TD, MacMillan DWC. *J Am Chem Soc*. 2005; 127:8826–8828. [PubMed: 15954790]
28. Nuzzi A, Massi A, Dondoni A. *Org Lett*. 2008; 10:4485–4488. [PubMed: 18788740]
29. Nolen EG, Watts MM, Fowler DJ. *Org Lett*. 2002; 4:3963–3965. [PubMed: 12599503]
30. For related one-pot reactions on simple aldehydes using the Jørgensen catalyst, Jiang H, Falcicchio A, Jensen KL, Paixão MW, Bertelsen S, Jørgensen KA. *J Am Chem Soc*. 2009; 131:7153–7157. [PubMed: 19419172]
31. Jimenez-Barbero J, Espinosa JF, Asensio JL, Cañada FJ, Poveda A. *Adv Carbohydr Chem Biochem*. 2001; 56:235–284. [PubMed: 11039113]
32. Massi A, Nuzzi A, Dondoni A. *J Org Chem*. 2007; 72:10279–10282. [PubMed: 18004870]

Figure 1.
Fluorinated precursors for complex C-glycosides

Figure 2.
X-ray for **11**

Scheme 1.
Fluorination-reduction on α -C-ethanal **5a**

Scheme 2.
Fluorination-methylenation on α -C-ethanal **5a**

Scheme 3.
Fluorination-reduction on β -C-ethanal **5b**

Scheme 4.
Synthesis of difluorinated derivatives

Scheme 5.
Mechanism for fluorination