Video Article

A CO₂ Concentration Gradient Facility for Testing CO₂ Enrichment and Soil Effects on Grassland Ecosystem Function

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Abstract

Continuing increases in atmospheric carbon dioxide concentrations (C_A) mandate techniques for examining impacts on terrestrial ecosystems. Most experiments examine only two or a few levels of C_A concentration and a single soil type, but if C_A can be varied as a gradient from subambient to superambient concentrations on multiple soils, we can discern whether past ecosystem responses may continue linearly in the future and whether responses may vary across the landscape. The Lysimeter Carbon Dioxide Gradient Facility applies a 250 to 500 μL⁻¹ C_A gradient to Blackland prairie plant communities established on lysimeters containing clay, silty clay, and sandy soils. The gradient is created as photosynthesis by vegetation enclosed in in temperature-controlled chambers progressively depletes carbon dioxide from air flowing directionally through the chambers. Maintaining proper air flow rate, adequate photosynthetic capacity, and temperature control are critical to overcome the main limitations of the system, which are declining photosynthetic rates and increased water stress during summer. The facility is an economical alternative to other techniques of C_A enrichment, successfully discerns the shape of ecosystem responses to subambient to superambient C_A enrichment, and can be adapted to test for interactions of carbon dioxide with other greenhouse gases such as methane or ozone.

Video Link

The video component of this article can be found at <http://www.jove.com/video/53151/>

Introduction

Atmospheric carbon dioxide concentration (C_A) has recently increased past 400 μL⁻¹ from approximately 270 μL⁻¹ prior to the Industrial Revolution. C_A is forecast to reach at least 550 μL⁻¹ by 2100¹. This rate of increase surpasses any C_A changes observed over the last 500,000 years. The unprecedented rate of change in C_A raises the possibility of non-linear or threshold responses of ecosystems to increasing C_A. Most ecosystem-scale C_A enrichment experiments apply only two treatments, a single level of enriched C_A and a control. These experiments have greatly expanded our understanding of the ecosystem impacts of C_A enrichment. However, an alternate approach that can reveal the presence of non-linear ecosystem responses to increasing C_A is to study ecosystems across a continuous range of subambient to superambient C_A. Subambient C_A is difficult to maintain in the field, and has most often been studied using growth chambers². Superambient C_A has been studied using growth chambers, open-top chambers, and free-air enrichment techniques^{3,4}.

C_A enrichment occurs across landscapes containing many soil types. Soils properties can strongly affect ecosystem responses to C_A enrichment. For example, soil texture determines the retention of water and nutrients in the soil profile⁵, their availability to plants⁶, and the amount and quality of organic matter⁷⁻⁹. The availability of soil moisture is a crucial mediator of ecosystem responses to C_A enrichment in water limited systems, including most grasslands¹⁰. Past field C_A enrichment experiments have typically examined only one soil type, and controlled tests of continuously varying C_A enrichment over several soil types are lacking. If effects of C_A enrichment on ecosystem processes differ with soil type, there is strong reason to expect spatial variation in ecosystem responses to C_A enrichment and ensuing changes in climate^{11,12}.

The Lysimeter Carbon Dioxide Gradient (LYCOG) facility was designed to address questions of spatial variation in non-linear and threshold responses of ecosystems to C_A levels ranging from ~ 250 to 500 μL⁻¹. LYCOG creates the prescribed gradient of C_A on perennial grassland plant communities growing on soils representing the broad range of texture, N and C contents, and hydrologic properties of grasslands in the southern portion of the U.S. Central Plains. Specific soils series used in the facility are Houston Black clay (32 monoliths), a Vertisol (Udic Haplustert) typical of lowlands; Austin (32 monoliths), a high carbonate, silty clay Mollisol (Udorthentic Haplustol) typical of uplands; and Bastisil (16 monoliths), an alluvial sandy loam Alfisol (Udic Paleustalf).

The operational principle employed in LYCOG is to harness the photosynthetic capacity of plants to deplete C_A from parcels of air moved directionally through the enclosed chambers. The treatment objective is to maintain a constant linear daytime gradient in C_A from 500 to 250 μL⁻¹. To accomplish this, LYCOG consists of two linear chambers, a superambient chamber maintaining the portion of the gradient from 500 to 390 (ambient) μL⁻¹ C_A, and a subambient chamber maintaining the 390 to 250 μL⁻¹ portion of the gradient. The two chambers are located side

by side, oriented on a north-south axis. The C_A gradient is maintained during the portion of the year when vegetation photosynthetic capacity is adequate; typically from late April to early November.

The chambers contain sensors and instrumentation needed to regulate the C_A gradient, control air temperature (T_A) near ambient values, and apply uniform precipitation amounts to all soils. Soils are intact monoliths collected from nearby Blackland prairie installed in hydrologically-isolated weighing lysimeters instrumented to determine all components of the water budget. Water is applied in events of volume and timing that approximate the seasonality of rain events and amounts during an average precipitation year. Thus, LYCOG is capable of evaluating the long-term effects of subambient to superambient C_A and soil type on grassland ecosystem function including water and carbon budgets.

LYCOG is the third generation of C_A gradient experiments conducted by USDA ARS Grassland Soil and Water Research Laboratory. The first generation was a prototype subambient to ambient gradient that established the viability of the gradient approach¹³ and advanced our understanding of leaf-level physiological responses of plants to subambient variation in C_A ¹⁴⁻²⁰. The second generation was a field-scale application of the concept to perennial C_4 grassland, with the gradient extended to 200 to 550 $\mu\text{L L}^{-1}$ ²¹. This field-scale experiment provided the first evidence that grassland productivity increases with C_A enrichment may saturate near current ambient concentrations²⁰, in part because nitrogen availability may limit plant productivity at superambient C_A ²². LYCOG extends this second generation experiment by incorporating replicated soils of varying texture, allowing robust testing for interactive effects of soils on the C_A response of grassland communities.

Protocol

1. Collect Soil Monoliths to be used as Weighing Lysimeters

1. Construct open-ended steel boxes 1 x 1 m square by 1.5 m deep from 8 mm thick steel.
2. Press the open-ended boxes vertically into the soil, using hydraulic presses mounted on helical anchors drilled 3 m deep into the soil.
3. Excavate the encased monolith using a backhoe or similar equipment.
4. Place a fiberglass wick in contact with soil at the base of the monolith. Pass the wick through the steel base into a 10 L reservoir to drain the monolith, and then weld the steel base onto the bottom of the box.
5. Kill existing vegetation on the monoliths by applying a non-residual herbicide, such as glyphosate.

2. Establish Plant Communities on Soil Monoliths

1. Plant the monoliths with eight seedlings each of seven species of tallgrass prairie grasses and forbs, for a total density of 56 plants per m^2 .
 1. Plant the following Grasses: *Bouteloua curtipendula* (side-oats grama), *Schizachyrium scoparium* (little bluestem), *Sorghastrum nutans* (Indiangrass), *Tridens albens* (white tridens)].
 2. Plant the following Forbs: *Salvia azurea* (pitcher sage), *Solidago canadensis* (Canada goldenrod), *Desmanthus illinoensis* (Illinois bundleflower, a legume).
2. Plant seedlings in a Latin Square design, re-randomized for each monolith.
3. Water the transplants for approximately 2 months following planting. The goal is to minimize water stress during initial establishment. Use any convenient method such as a hand wand or garden sprinkler. The frequency of watering depends on local climate and weather, particularly the occurrence of ambient rainfall.
4. Following the initial transplant establishment phase, maintain the transplants under ambient rainfall for as long as necessary while chambers (Section 3) are constructed. Remove unwanted species that emerge in the monoliths during establishment by hand-weeding.

3. Chamber Design

1. Construct two chambers each 1.2 m wide, 1.5 m tall, and 60 m long, divided into ten 5 m long sections. Construct sections from heavy steel of dimensions 5 m x 1.2 m x 1.6 m deep, buried to 1.5 m.
 1. Install four monoliths in each section, two monoliths each of two of the soil types, in random order. Install each monolith atop a 4,540 kg capacity balance.
 2. Include Bastil monoliths in the pairings in even numbered sections.
2. Join adjacent sections aboveground with a 1 m long x 1 m wide x 0.3 m tall sheet-metal duct to provide a pathway for airflow.
 1. Supply coolant at 10 °C from a 161.4 kW refrigeration unit to a cooling coil inside each duct.
 2. Enclose the vegetation with clear greenhouse film (thickness 0.006"/.15 mm), such as used in other climate manipulation experiments²³.
 3. Fit each cover with a zippered opening backed by a draft flap to allow access to the monoliths for sampling.
 4. Remove the polyethylene covers at the end of the growing season.

4. CO₂ and Air Temperature Measurement; Temperature Control

1. Sample entry and exit C_A on both chambers every 2 min through filtered air sample lines located at the entry and exit of superambient and subambient chambers. These data inform CO₂ injection and fan speed control.
 1. Sample C_A and water vapor content, and measure air temperature (T_A) at the entry and exit of each 5 m section at 20 min intervals.
 2. Measure all air samples for CO₂ and water vapor content in real time using infrared gas analyzers according to manufacturer's protocol.

3. Measure T_A at the entry, midpoint, and exit of each section with shielded fine wire thermocouples.
2. Regulate the flow of coolant through the cooling coil at the entrance of each section to maintain a consistent mean (mid-section) T_A from section to section near the ambient T_A .
3. Position a quantum sensor to have an unobstructed view of the sky and measure photosynthetic photon flux density according to manufacturer's protocol. Light level is an input to the blower control algorithm.

5. C_A Treatment Application

1. Daytime
 1. Mix pure carbon dioxide (CO_2) with incoming ambient air to $500 \mu L^{-1} C_A$, using a mass flow controller in the entrance duct of the superambient leg. See Section 4 for C_A measurement details.
 2. Advect the enriched air through the chambers using blower fans at the entrance to section 1 and in downstream sections.
 3. Maintain the desired exit C_A of $390 \mu L^{-1}$ (ambient air) by adjusting the blower speed.
 1. Increase the blower speed if the exit C_A is below the set point. This allows less time for plant uptake of CO_2 , resulting in higher exit C_A .
 2. Decrease the blower speed if exit C_A is above the set point.
 4. Use the same approach in the subambient chamber except introduce ambient air and control to achieve exit C_A of $250 \mu L^{-1}$.
2. Nighttime
 1. Reverse the direction of air flow.
 2. Inject CO_2 into the daytime exit end of the superambient chamber to achieve $530 \mu L^{-1} C_A$, and control advection rates to maintain $640 \mu L^{-1}$ at the nighttime exit (daytime entrance).
 3. Introduce ambient air at $\sim 390 \mu L^{-1} CO_2$ into the nighttime entrance (daytime exit) of the subambient chamber and control advection rate to maintain $530 \mu L^{-1}$ at the nighttime exit.

6. Precipitation Inputs

1. Apply the mean growing season rainfall amount to each monolith.
 1. Supply water to each monolith from a domestic water source through a drip irrigation system. Schedule the irrigation events and application amounts to approximate the seasonal rainfall pattern for the experiment location. The exact schedule depends on local climate.
2. Control application timing with a data logger and measure application volumes with flowmeters.

7. Sampling

1. Measure vertical profiles of volumetric soil water content (vSWC) weekly during the period of CO_2 control, with a neutron attenuation gauge or other appropriate probe.
 1. Recommended profile increments are 20 cm depth increments to 1 m depth, and one 50 cm increment below a 1 m.
2. Measure monolith aboveground net primary productivity (ANPP) by harvesting all standing aboveground biomass at the end of the growing season.
 1. All aboveground biomass is removed each year, consequently standing biomass represents current primary production.
 2. Sort the sampled biomass by species, dry to constant mass, and weigh.
 3. Use biomass of individual species to quantify plant species contributions to ANPP.

Representative Results

The superambient and subambient portions of the gradient are maintained in separate chambers (**Figure 1**). However, over seven years of operation (2007 – 2013), the chambers maintained a linear gradient in C_A concentration from 500 to $250 \mu L^{-1}$ (**Figure 2**) with only a small discontinuity in C_A between the exit of the enriched chambers (Monolith 40) and the entrance of the subambient portion of the gradient (Monolith 41).

Air temperature and vapor pressure deficit remained constant from section to section in both the superambient and subambient chambers, except in section 10 of the superambient chamber, and sections 19 and 20 of the subambient chamber, where air temperature averaged $\sim 3^\circ C$ warmer than other sections (**Figure 2**). However there can be pronounced temperature increases of $5 - 7^\circ C$ within each section, and corresponding increases in vapor pressure deficit.

Averaged over the 2007 – 2013 growing seasons, vSWC varied linearly along the C_A gradient on two of the three soils (**Figure 3**). vSWC in the top 20 cm of the soil profile increased by 3.1% per $100 \mu L^{-1}$ increase in C_A on the sandy loam (Bastisil series) soil ($R^2 = 0.34$, $p = 0.01$), and by 1.7% per $100 \mu L^{-1} C_A$ on the clay soil (Houston series). However there was no change in 0-20 vSWC in the silty clay (Austin series) soil ($p = 0.13$).

Plant productivity also varied linearly with by C_A , and the magnitude of the C_A response differed among soils. ANPP (**Figure 4A**) of monoliths with Blackland prairie plant communities had the smallest response to C_A on the clay soil, increasing by 59 g m^{-2} per $100 \mu\text{L L}^{-1}$ increase in C_A ($R^2 = 0.22$, $p = 0.02$). The ANPP response to C_A enrichment was intermediate on the silty clay soil, increasing by 76 g m^{-2} per $100 \mu\text{L L}^{-1}$ of CO_2 ($R^2 = 0.22$, $p = 0.02$), and greatest on the sandy loam soil, where ANPP gained 131 g m^{-2} per $100 \mu\text{L L}^{-1}$ of CO_2 ($R^2 = 0.55$, $p < 0.001$).

These soil-specific responses of ANPP to C_A corresponded closely to the soil-specific responses of a mesic C_4 tallgrass, *Sorghastrum nutans*, the most abundant grass species in the experimental plant communities. Aboveground biomass of *S. nutans* increased most strongly with increased C_A on the sandy loam soil, gaining over 200 g m^{-2} for every $100 \mu\text{L L}^{-1}$ increase in C_A ($R^2 = 0.40$, $p = 0.005$). In contrast, *S. nutans* gained only 100 g m^{-2} per $100 \mu\text{L L}^{-1}$ increase in C_A on the silty clay soil ($R^2 = 0.50$, $p < 0.0001$), while *S. nutans* responded marginally to C_A on the clay soil ($R^2 = 0.12$, $p = 0.07$; **Figure 4B**).

The soil-specific increase in ANPP with C_A enrichment occurred despite decreases in productivity on the two more responsive soils by the xeric C_4 mid-grass *Bouteloua curtipendula* (**Figure 4C**). *B. curtipendula* was the second most abundant species in the experimental communities. On the silty-clay soil, *B. curtipendula* was the dominant grass at subambient C_A concentrations but decreased most strongly with C_A enrichment on the silty clay soil (69 g m^{-2} , per $100 \mu\text{L L}^{-1}$ increase in C_A ; $R^2 = 0.36$, $p < 0.008$), decreased less strongly on the sandy loam soil (44 g m^{-2} , per $100 \mu\text{L L}^{-1}$ increase in C_A ; $R^2 = 0.36$, $p = 0.008$), and did not vary with C_A enrichment on the clay soil ($p = 0.46$).

Figure 1. Arrangement of chambers and soils. The two linear sequences of chambers containing grassland vegetation growing on intact soil monoliths (photo), and schematic of the distribution of the three soil types along the CO_2 gradient. Plot numbers 1 - 40 are located along the $500 - 380 \mu\text{L L}^{-1}$ portion of the gradient, and numbers 41 - 80 on the $380 - 250 \mu\text{L L}^{-1}$ portion. Photo: Philip Fay.

Figure 2. Microclimate along the C_A gradient. Daytime growing season carbon dioxide (CO₂) concentration, air temperature, and vapor pressure deficit for the 80 monoliths in the enriched and subambient chambers. Values are measured at the air entry and exit of each section, and estimated from linear interpolation for other positions. Data points represent means for the 2007 through 2013 growing seasons. Error bars omitted for clarity; mean standard errors were 3.5 for CO₂, 0.82 for air temperature, and 0.18 for vapor pressure deficit.

Figure 3. Soil moisture on each soil type along the CO₂ gradient. Growing season volumetric soil water contents (vSWC) for 0 – 20 cm in the soil profile for each soil type, plotted by position along the CO₂ concentration gradient. Linear regressions are plotted for soils with significant relationships of vSWC to CO₂ concentration. Data points represent means of 2007 through 2013 growing seasons. Error bars omitted for clarity; mean standard errors on the three soils ranged from 0.74 to 0.99.

Figure 4. Plant productivity on each soil type along the CO₂ gradient. (A) Mean aboveground net primary productivity (ANPP), the sum of the current-year biomass of all species in the 60 monoliths with Blackland Prairie plant communities; and the current year biomass of (B) the mesic C₄ tallgrass, *Sorghastrum nutans*, and (C) the xeric C₄ midgrass *Bouteloua curtipendula* plotted by position along the CO₂ concentration gradient. Linear regressions are plotted for soils with significant relationships of ANPP or species biomass to CO₂ concentration. Data points represent means of 2007 through 2013 growing seasons. Error bars omitted for clarity; mean standard errors on the three soils ranged from 34.9 to 42.5 for ANPP, 21.8 to 34.4 for *S. nutans*, and 7.4 - 24.8 for *B. curtipendula*.

Discussion

The LYCOG facility achieves its operational goal of maintaining a 250 to 500 $\mu\text{L L}^{-1}$ continuous gradient of C_A concentrations on experimental grassland communities established on three soil types. The change in C_A is linear over the prescribed range. Air temperature increased within each section, but was reset by the between-section cooling coils in most sections. As a result, the operational goal of maintaining a consistent mean temperature from section to section was met over most of the gradient. Temperature and C_A control are easily maintained during the spring and early summer when soil moisture is relatively high and plants are at their highest photosynthetic capacity.

Critical Steps in the Protocol

Control of blower speed is the most critical aspect of maintaining the prescribed CO₂ gradient. Control is based on a combination of feedback and feed-forward techniques to match air flow to the vegetation carbon uptake. The feedback technique adjusts fan speed based on the difference between measured and target exit CO₂ concentration. Feed-forward control anticipates changes in photosynthetic rate and rapidly (5 sec response time) adjusts fan speed, based on changes in photosynthetically active radiation measured with the quantum sensor. Feed-forward control considerably improves control over that achieved by feedback control alone. The maximum airflow rate through the chambers is on the order of 1 m sec⁻¹, or about 3.6 km hr⁻¹, which is on the low-end of wind speeds these plants see in the field. Thus, varying fan speed is unlikely to affect plant responses.

Another critical aspect of maintaining the CO₂ gradient is the presence of adequate photosynthetic capacity. The steeper the gradient, the greater the canopy photosynthetic capacity required to draw down the CO₂ concentration. Species or communities with more leaf area, higher photosynthetic rates, or longer chamber length all increase the CO₂ draw-down that can be achieved. Care should also be taken the monolith volume and depth is chosen to provide a realistic rooting volume for the established plant communities. The species used here have rooting depths of 1 – 1.5 m, but other species may be shallower or deeper, and monolith volume should be adjusted accordingly. The final critical aspect is the importance of reliably supplying and controlling the flow of chilled water to the cooling coils between each section, in order to match chamber temperatures to outside diurnal and seasonal variation in outside ambient temperature.

Modifications to the Technique

The first year of operation revealed that the prairie vegetation was marginally capable of adequate CO₂ drawdown. This was remedied by converting a total of 20 monoliths chosen from the Houston and Austin soil series to monocultures of switchgrass, *Panicum virgatum*. Switchgrass is a highly productive native tallgrass, and is well-watered throughout the growing season, which insures adequate C uptake capacity along the gradient even during the hot summer months. The first year also revealed greater than anticipated aerodynamic resistance in the chambers, which degraded flow rates in downstream chambers, leading to overheating. This issue was remedied by installation of additional downstream blower fans to boost the flow rates. We recommend installing new polyethylene covers each growing season to maintain maximum light transmittance.

Limitations of the Technique

The system poses certain operational issues that create both opportunities and limitations on the research questions the facility can support. Control of the gradient becomes more difficult from mid-summer through the end of the growing season, because higher summer temperatures lower soil moisture, increasing plant water stress and lowering photosynthetic capacity. This in turn requires slower air flow rates to achieve the C_A draw-down needed to meet the target C_A concentrations, which in turn further raises temperatures. This dynamic illustrates the limited ability of this system for studies of drought interactions with CO₂ concentration. Temperature increases within each 5 meter section are unavoidable because of the linear flow design of the experiment. Long-wave energy accumulates within each chamber until air passes through the cooling coil and entering the next chamber. Within-section temperature increases are of similar magnitude to some of the higher estimates for future temperature increases expected with some climate change scenarios. Thus, the within-section temperature variation represents an opportunity to analyze grassland responses to interactions between C_A and warming. Finally, the dimensions of the chamber limit the vegetation to a maximum height of about one meter, and the monolith area limits the vegetation to herbaceous species with smaller basal areas. The use of tree species, for example to study woody encroachment into grassland, would be impractical beyond the seedling stage.

Significance compared to Other Techniques

LYCOG is considerably more economical to operate compared to techniques such as FACE and OTC. LYCOG uses approximately 3,700 L per month of CO₂, which is greater than the CO₂ use in MiniFACE systems²⁴, but much less than the CO₂ consumption of FACE and OTC approaches^{3,12}. The major expense of maintaining the experiment comes from temperature control, which costs approximately \$30,000 per year, comparable to estimates of the CO₂ expense for open top chamber C_A enrichment costs but still much less than that of the CO₂ expense of Free Air CO₂ Enrichment systems³. The economic advantages come in addition to the unique capability of supporting studies at subambient CO₂ and along a continuous CO₂ gradient.

Current and Future Applications

Current research is examining ecosystem responses other than ANPP, including soil CO₂ efflux, and evapotranspiration, which will expand our understanding of soil-specific variation in C_A effects on grassland carbon and water cycling. Future possibilities for research include combining temperature and CO₂ treatments, for example by operating both chambers as superambient but maintaining one chamber at a warmer temperature differential with respect to ambient. Current vegetation can easily be replaced with other species or communities to study how variation in community structure influences CO₂ effects on ecosystem function. Other ecologically important atmospheric constituents such as methane or ozone could be added to test for interactions with CO₂.

Disclosures

The authors have nothing to disclose.

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