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No Effect of Acute Exposure to Coarse Particulate Matter Air Pollution in a Rural Location on High Density Lipoprotein Function

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Abstract

Context—High density lipoprotein (HDL) particles perform numerous vascular-protective functions. Animal studies demonstrate that exposure to fine or ultrafine particulate matter (PM) can promote HDL dysfunction. However, the impact of PM on humans remains unknown.

Objective—We aimed to determine the effect of exposure to coarse concentrated ambient particles (CAP) on several metrics of HDL function.

Methods—Thirty-two adults (25.9 ± 6.6 years) were exposed to coarse CAP [$76.2 \pm 51.5 \mu\text{g}\cdot\text{m}^{-3}$] in a rural location and filtered air (FA) for 2-hours in a randomized double-blind crossover study. Venous blood collected 2 and 20 hours post-exposures was measured for HDL-mediated efflux of [³H]-cholesterol from cells and 20 hours post-exposures for HDL anti-oxidant capacity by the HDL oxidation index (HOI) and paraoxonase activity. The changes [median (first, third quartiles)] between exposures among 29 subjects with available results were compared by matched Wilcoxon tests.

Results—HDL-mediated cholesterol efflux capacity did not differ between exposures at either time point [16.60% (15.17, 19.19) 2-hours post-CAP versus 17.56% (13.43, 20.98) post-FA, $p=0.768$ and 14.90% (12.47, 19.15) 20-hours post-CAP versus 17.75% (13.22, 23.95) post-FA, $p=0.216$]. HOI [0.26 (0.24, 0.35) versus 0.28 (0.25, 0.40), $p=0.198$] and paraoxonase activity [0.54

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Declaration of interest statement

The authors report no declarations of interest.

(0.39, 0.82) versus 0.60 $\mu\text{mol}\cdot\text{min}^{-1}\cdot\text{ml plasma}^{-1}$ (0.40, 0.85), $p=0.137$] did not differ 20-hours post-CAP versus FA, respectively.

Conclusions—Brief inhalation of coarse PM from a rural location did not acutely impair several facets of HDL functionality. Whether coarse PM derived from urban sites, fine particles, or longer-term PM exposures can promote HDL dysfunction warrant future investigations.

Keywords

Air pollution; atherosclerosis; high density lipoprotein

Introduction

Fine particulate matter (PM) $< 2.5 \mu\text{m}$ (PM_{2.5}) air pollution is a leading cause of global morbidity and mortality, with a substantial portion of the deaths attributable to cardiovascular (CV) causes. Supporting these associations, numerous studies have shown that PM_{2.5} exposures are capable of eliciting a variety of adverse biological responses which could potentially play a role in triggering acute CV events (Brook et al. 2010).

On the other hand, whether coarse PM (2.5–10 μm in aerodynamic diameter; PM_{10-2.5}) is also associated with adverse CV health effects is less clear given the mixed results from epidemiological observations (Brunekreef et al. 2005; Chang et al. 2011; Peng et al. 2008; Puett et al. 2009; Zanobetti et al. 2009). In addition to its larger size, coarse PM differs from PM_{2.5} in several regards including sources and components. PM_{10-2.5} itself is a heterogeneous mixture of particles typically generated from mechanical activities (e.g., crushing, suspension of ground materials), rather than from the combustion of fossil fuels as with PM_{2.5}. The sources of coarse PM range from farming, roadway dust, to construction activities. Its major constituents (metals, crustal material such as silicon, calcium, and bio-aerosols including endotoxins) also differ according to nearby activities and landscape features (Brunekreef et al. 2005). In light of the uncertainty regarding the CV health effects of coarse PM, we have recently undertaken a series of experiments investigating the potential biological effects of acute exposures.

Numerous pathways have been elucidated whereby PM could plausibly trigger acute CV events (Brook et al. 2010). Recent evidence also supports the contention that long-term exposures can even promote the development of chronic CV disease-states such as atherosclerosis (Adar et al 2013). One emerging biological pathway potentially responsible for a portion of both these acute and chronic health effects is air pollutant-mediated high-density lipoprotein (HDL) dysfunction. In addition to being the central mediator of reverse cholesterol transport (RCT), HDL plays key roles in vascular homeostasis (e.g., anti-inflammatory, anti-oxidant activities) which together serve as powerful inhibitors of the atherosclerotic process and also help to prevent plaque instability/vulnerability (Bandeali et al 2012; Eren et al. 2012; Rosenson et al. 2011; Soran et al 2009). Beyond the association between low HDL-cholesterol levels, dysfunctional HDL particles per se (in particular impaired cholesterol efflux capacity and anti-oxidant actions) are of clinical importance as they have been shown to be independent predictors of CV events and/or the presence of underlying atherosclerosis (Khera et al 2011; Li et al 2013B; Patel et al 2011). In the context

of air pollution, recent animal experiments have also demonstrated that exposure to fine or ultrafine PM ($<0.1 \mu\text{m}$) over a few weeks are capable of impairing several aspects of HDL functionality including anti-inflammatory capacity, anti-oxidant capacity and paraoxonase activity (Araujo et al 2008; Li et al 2013A; Yin et al. 2013).

However, no study has evaluated the effect of ambient PM air pollutants (of any size fraction or duration of exposure) on HDL function in humans. While disease-states that induce HDL dysfunction (e.g., diabetes) as well as the ensuing sequelae (e.g., atherosclerotic progression) are most commonly thought to be chronic processes, it is becoming increasingly clear that even acute pro-inflammatory/oxidative insults (e.g., infections) can impair HDL function within hours-to-days (G et al 2011; Eren et al. 2012; Patel et al 2011; Smith 2010). This abrupt genesis of dysfunctional HDL particles (e.g., shifting from an anti- to a pro-oxidant phenotype) could plausibly play a role in the acute instigation of plaque instability and thus the attendant CV events (Eren et al. 2012). As such, we aimed to explore if acute exposure to coarse concentrated ambient particle (CAP) in a rural location is capable of altering clinically-relevant facets of HDL function – the ability of HDL to facilitate the efflux of cholesterol from cells, antioxidant capacity (HDL oxidant index [HOI]) and paraoxonase activity (Eren et al 2012; Khera et al 2011; Patel et al 2011; Rosenson et al. 2011). In this paper we report the changes in HDL function (pre-defined secondary outcomes) induced by coarse PM from a recently completed controlled exposure study.

Methods

The Institutional Review Board of the University of Michigan approved this study. During a screening visit subjects signed an informed consent document and had blood labs drawn (fasting lipids, glucose) and a history and physical exam performed. Inclusion criteria included nonsmoking adults (no active smoking within the past year per self-report during the screening visit and living in nonsmoking households) who were from 18–50 years of age without any established CV disease or traditional CV risk factors. To be enrolled subjects were required to have screening blood pressures $< 140/90$ mm Hg and fasting glucose levels $< 126 \text{ mg}\cdot\text{dL}^{-1}$. Subjects were excluded if they were taking medications (e.g., statins) or over-the-counter pills (e.g., anti-oxidants) that might alter study outcomes.

Enrolled subjects entered into a randomized double-blind crossover study (from May 2011 to June 2012) comparing the health effects of 2-hour-long exposures to coarse CAP versus filtered air (FA). Subjects came to the research facility on each day having fasted for > 8 hours with randomized exposures occurring from 10 am to noon. Approximately 12.5 ml peripheral venous blood was drawn into sodium heparin tubes 20 hours post-exposures for later analyses of HOI and paraoxonase. Approximately 4 ml of venous blood was drawn into EDTA tubes both 2 and 20 hours post-exposures for later analyses of HDL-mediated cholesterol efflux assays. The plasma was separated from blood within 2 hours after collection and immediately stored in a -80 degree Celsius ($^{\circ}\text{C}$) freezer for later batched analyses. There was a 1–3 week washout period between exposures.

Exposure Facility and Air Pollution Measurements

Dexter Michigan was selected as the location for coarse PM exposures as it is highly influenced by rural sources (http://www.epa.gov/castnet/javaweb/site_pages/ANA115.html). This site is >10 km from freeways and >60 km west of the Detroit metropolitan area. Coarse CAP was generated by a 2-stage virtual impactor system (Demokritou et al. 2002; Moffet et al. 2004) which concentrates ambient coarse PM (predominantly from 2.5–10 μm) without substantively altering the composition or chemistry. The AIRCARE mobile air research laboratory has been described in detail elsewhere (Harkema et al. 2004). Randomized, blinded exposures were delivered to subjects who were resting seated within an air-tight chamber via a facemask with an air flow rate between 25–28 liters·minute⁻¹. Gaseous pollutants (e.g., ozone) remained at or below ambient levels for both exposures. To generate FA a high efficiency PM filter was inserted at the inlet of the concentrator (i.e., “upstream” to the series of virtual impactors). Coarse CAP mass levels were monitored during exposures downstream to the concentrator with a DataRAM monitor (Thermo Scientific, Waltham, MA). Outdoor and within chamber temperatures (maintained at approximately 24 degrees Celsius) were also monitored. Filter samples were collected immediately upstream of the exposure chamber on 47-mm Teflon filters (Pall, Ann Arbor, MI) at a flow rate of 6 L/min. The samples were analyzed gravimetrically for particle mass using a microbalance (MT-5 Mettler Toledo, Columbus, OH) in a temperature/humidity-controlled clean laboratory as described in the Federal Reference Method (USEPA. 1997. Reference method for the determination of fine particulate matter as PM_{2.5} in the atmosphere. EPA 40 CFR Pat 50. Washington DC).

HDL isolation and cholesterol efflux methods

Stored plasma samples were adjusted to $d=1.21$ g/mL with anhydrous potassium bromide powder and centrifuged (Sorvall WX-90, TFT-80.2 rotor, 429,600 g, 8 hours, 4°C). The top 200 μL from the tubes was removed and 0.45 μm filtered into chromatography vials. Next, the obtained total lipoprotein fraction was resolved on Superose 6 100/300 GL column (GE Healthcare) using fast protein liquid chromatography (Bio-Rad DuoFlow equipped with C96 autosampler with cooling). Lipoproteins were eluted with an isocratic buffer containing 10 mM phosphate buffer, 154 mM NaCl, and 0.02% sodium azide, pH 7.4, at a flow rate of 0.5 mL/min. Fractions containing HDL were collected by automatic fraction collector at 4°C, yielding albumin-free HDL at concentrations of 0.7–2.5 mg/ml. The presence and purity of HDL was confirmed by 15% PAGE electrophoresis using ApoA-I (Sigma) as a reference.

HDL-mediated cholesterol efflux experiments were performed as described previously (Li et al. 2013B), with minor modifications. RAW cells (mouse macrophages) were obtained from The American Type Culture Collection and cultured in Roswell Park Memorial Institute (RPMI) medium supplemented with 5% fetal bovine serum (FBS). Cells were plated at a density of 4×10^5 cells per well in 24-well culture plates. Cells were allowed to attach overnight after which medium was replaced with in 250 μL of Dulbecco's Modified Eagle Medium (DMEM) containing 0.2% BSA, 1 μCi of H³-cholesterol with or without 0.3 mM 8-chloroadenosine 3',5'-cyclic-monophosphate (cAMP) (Sigma). Cells then were allowed to load with H³-cholesterol for a total of 8 hours. The cells were washed three times and equilibrated for one hour in DMEM. HDL-mediated cholesterol efflux was then determined

by incubating the test HDL with H^3 -cholesterol labeled cells for 14 hours at 37°C. Radioactivity in the medium supernatants and total cell radioactivity were measured on scintillation counter.

HDL-mediated cholesterol efflux capacity was calculated as: % Efflux = $100 \times [\text{Disintegrations per minute (DPM) of Efflux Medium}] / ([\text{DPM of Efflux Medium} + \text{Cell DPM}])$; non-specific efflux (without HDL) data were subtracted from % Efflux for each sample. In the case of cAMP-stimulated cells, adenosine-5'-triphosphate-binding cassette transporter (ABC)-A1 and ABCG1-mediated efflux were calculated using following equations: a) %Efflux(ABCA1+ABCG1) with cAMP = %Efflux(+cAMP); b) %Efflux(ABCA1) = %Efflux(+cAMP) — %Efflux(without cAMP)]. Thus, cAMP-stimulated efflux represents total of ABCA1+ABCB1-mediated cholesterol efflux while efflux without cAMP stimulation represents predominately ABCG1-mediated efflux.

HDL anti-oxidant capacity and paraoxonase activity methods

HDL anti-oxidant capacity was assessed by a fluorescence assay that evaluated the ability of HDL to inhibit LDL oxidation, determined by dichlorofluorescein (DCF) fluorescence. HDL cholesterol was isolated from thawed plasma samples using the dextran-sulphate precipitation method. Briefly, 50 µl of plasma was incubated with 10 µl of LipiDirect Magnetic HDL cholesterol precipitating reagent (Reference Diagnostics, Inc, Bedford, MA) for 10 minutes at room temperature. The mixture was then centrifuged at 12000 rpm for 5 minutes at 4°C and the HDL containing supernatant was collected. HDL cholesterol concentration was determined by a colorimetric assay (Thermo Scientific, Middletown, VA). The HDL supernatant was used in the assay to evaluate the ability of HDL to inhibit LDL-induced oxidation of dihydrodichlorofluorescein (DCFH) into the fluorescent DCF as previously described (Yin et al 2013). The DCFH reagent was prepared by incubating dichlorofluorescein diacetate (DCF-DA) (Sigma-Aldrich, St. Louis, MO) at 2 mg/ml with 0.1 M NaOH. DCF-DA was hydrolyzed to DCFH and the mixture was neutralized with 10× PBS. Equal concentrations of HDL and LDL cholesterol (50 µg /ml) were mixed with Tris-HCl buffer (pH 7.4) up to a total volume of 100 µl and added to a black well flat bottom microtiter plate in triplicate. The plate was incubated at 37°C for 1 hour. Next, 20 µl of the DCFH reagent was added to each well and the plate was incubated at 37°C for 3 hours. DCF fluorescence intensity was determined using a plate reader (SynergyMx, BioTek, Vermont) at an excitation wavelength of 485 nm and emission wavelength of 530 nm. HDL anti-oxidant capacity was expressed as a HDL oxidant index (HOI), determined by the ratio of DCF fluorescence in the presence and absence of HDL cholesterol. An index < 1.0 denotes protective anti-oxidant HDL whereas an index > 1.0 indicates pro-oxidant HDL. The samples were assayed in triplicates, and the mean intra-assay coefficient of variation for all samples was 9.0%. To control for assay variability across different plates, a normal plasma HDL sample from a healthy individual was included on each plate (control sample). The inter-plate and inter-day variability calculated using this control sample was 5.7% and 15.7%, respectively. The HOI values of test samples obtained in each plate were adjusted according to the variability exhibited by the control sample on each plate to generate adjusted HOI values.

Paraoxonase-1 (PON-1) is an important HDL-associated anti-oxidant enzyme. Its activity was performed using 5 µl of plasma in the assay mixture consisting of paraoxon substrate, 2 mM CaCl₂, 2.0 M NaCl in 100 mM tris-HCl buffer (pH 8.5) as previously described (Yin et al 2013). The hydrolysis of paraoxon substrate (diethyl-*p*-nitrophenyl phosphate) to *p*-nitrophenol by PON-1 was quantified by spectrophotometry. The assay was performed at room temperature and the kinetics of *p*-nitrophenol formation was determined by recording absorbance at 405 nm every 15 seconds for 4 minutes. The enzyme activity was expressed as µmol *p*-nitrophenol formed per minute for every 1 mL of plasma.

Statistical Methods

Summary statistics were computed for continuous measures as mean ± standard deviation (SD) or median (interquartile range, IQR) if the distribution was skewed. The matched pairs of HDL function measurements after CAP versus FA exposures were compared using the matched Wilcoxon tests among the 29 subjects with all results completed. The associations between HDL function metrics were analyzed using linear mixed models. All analyses were performed using the statistical software package R (version 2.14.1).

Results

Subjects (n=32) who completed the study were healthy, as per the inclusion criteria (Table 1). PM_{10-2.5} levels were higher during coarse CAP compared to FA exposures (Table 2). There were no differences between exposures in ambient or intra-chamber temperatures (results not shown). There were no significant differences detected in cholesterol efflux capacity metrics to HDL from subjects exposed 2 hours or 20 hours following CAP versus FA exposures (Table 3; Figure 1). There was a non-significant trend for decreased PON activity (p=0.137) 20 hours following CAP as hypothesized but no differences in the HOI indices (Table 4; Figure 2). Except for the 2 metrics of cholesterol efflux, the correlations between differing HDL functional assays were not significant (Table 5).

Discussion

Brief exposure to coarse CAP from a rural location did not impair HDL function over a period of 20 hours. Three clinically-relevant markers including the HOI (i.e., the capacity of HDL to prevent LDL oxidation), paraoxonase activity (i.e., an important HDL-associated anti-oxidant enzyme), and HDL-mediated cholesterol efflux capacity metrics were not significantly altered by the inhalation of high levels of coarse PM compared to FA. As such, we did not find evidence in this short-term study to support that HDL dysfunction is likely a pertinent mechanism explaining the heightened CV risk shown to occur following acute elevations in ambient coarse PM levels in some epidemiological studies (Brunekreef et al. 2005; Chang et al. 2011; Peng et al. 2008; Puett et al. 2009; Zanobetti et al. 2009)

While a prior experiment has shown that short-term occupational exposure to welding fumes can cause an acute reduction in HDL cholesterol levels, lipoprotein functional parameters were not directly assessed (Rice et al. 2011). Therefore, this was the first study to explore the effect of any ambient air pollutant on HDL function in humans. Numerous questions thus remain unanswered by this single experiment. Whether coarse PM comprised of

differing chemical constituents (e.g., urban particles), fine or ultrafine particles, and/or longer-term exposures to PM (of any size fraction) are capable of impairing HDL function in humans warrant future investigations. Moreover, HDL particles convey a variety of pleiotropic actions normally protecting vascular health and homeostasis (Bandeali et al 2012; Eren et al. 2012; Rosenson et al. 2011; Soran et al 2009). The potential role played by HDL dysfunction in mediating several of the germane air pollution-induced adverse CV health effects (e.g., endothelial dysfunction, atherosclerosis) (Brook et al 2010) should also be explored.

Prior Animal Studies Relating Air Pollutants to HDL Function

There have been 3 published studies regarding the effects of air pollutants on HDL function. All of them were conducted over several weeks of exposures in animals and none of them evaluated coarse PM (Araujo et al 2008; Li et al 2013 (A); Yin et al. 2013). Araujo et al (2008) were the first to show that air pollutants could negatively impact HDL functionality. In this experiment, both fine and ultrafine PM exposures impaired the anti-inflammatory capacity of HDL measured by a cell-based assay. There was indirect evidence to support that this HDL dysfunction may have contributed to the concomitant increase in systemic atherosclerosis. More recent experiments have demonstrated that diesel exhaust and ultrafine PM can both impair HOI and paraoxonase activity, measured by the same (Yin et al. 2013) or similar techniques (Li et al 2013A) as in our current study. However, while diesel exhaust impaired HDL anti-oxidant and anti-inflammatory capacities, it did not induce changes in the HDL-mediated cholesterol efflux capacity (Yin et al, 2013). In these studies, diesel exhaust (Yin et al 2013) and ultrafine PM (Li et al 2013A) led to negative correlations between HOI and PON activity, suggesting that alteration in PON activity could be causally related to the HDL dysfunction. However, in the present study while we did observe a trend for decreased paraoxonase activity 20 hours following CAP versus FA as hypothesized, there were also trends for unexpected (paradoxical) decreased HOI values, which is suggestive of HDL particles with stronger antioxidant capacities. Given the statistical non-significance of both outcomes it is not possible to know if this is a true biological trend or simply chance alone. Future larger studies will be able to address this issue.

The putative mechanisms underlying the air pollution-induced HDL dysfunction in prior animal studies have not been fully elucidated. However, HDL particles can rapidly switch their phenotype into dysfunctional particles in the setting of acute pro-inflammatory or oxidative insults such as infections, illnesses, or acute coronary syndromes (G et al 2011; Eren et al. 2012; Patel et al 2011; Smith 2010). In fact, HDL can become pro-atherosclerotic in these situations over several hours-to-days. Cytokine-related alterations in acute phase proteins secreted from the liver leading to an altered protein cargo within HDL particles and/or myeloperoxidase-induced oxidation of HDL proteins (e.g., apolipoprotein-A1) after PM inhalation may be responsible. Air pollution is well-known to elicit numerous systemic pro-inflammatory and oxidative responses (Brook et al 2010). In theory, given the close proximity of inhaled pollutant particles that reach the terminal bronchioles/alveoli to HDL (either within the microvasculature or the lung interstitium), there may be direct effects of deposited air pollutant components (e.g., pro-oxidant metals or organic carbon species) upon HDL composition and subsequent function. Alternatively, lung-based cells known to

interact with PM (e.g., resident macrophages) may produce oxidants as part of an immune response that subsequently affects the nearby HDL particle. As such, prior research suggests that our hypothesis was plausible that even a 2-hour exposure to coarse CAP could impair one or more aspect of HDL function.

Study Limitations

We speculate that our present findings were negative largely because the time course of exposure was inadequate to elicit the requisite systemic responses that produce dysfunctional HDL. Alternatively, the coarse CAP derived from a rural setting may not have contained the necessary chemical constituents. The PM components are indeed likely important for the genesis of HDL dysfunction as ultrafine has been shown to cause larger changes than fine PM (Araujo et al 2008). Ongoing filter analyses of PM components during these exposures and coarse CAP exposures in a different urban setting will hopefully provide insights into these latter issues.

Unlike ultrafine and fine PM, coarse particles are more commonly deposited in larger extra-thoracic airways (Geiser and Kreyling, 2010). This may have limited the “dose” of coarse PM that actually reached subjects’ bronchioles/alveoli upon inhalation. It may be a prerequisite that particles deposit in smaller airways in order to instigate HDL dysfunction (as mechanistically speculated in the prior section). Nevertheless, it is estimated that between 15–30% of inhaled coarse PM will reach small airways (Geiser and Kreyling, 2010). It is also not known precisely where in the pulmonary tree it is required for particles to reach or interact with lung-based cells in order to promote HDL dysfunction (or any systemic CV actions for that matter) (Brook et al. 2010). Future studies evaluating the potential differences on HDL dysfunction elicited by inhaled PM of varying sizes may help elucidate this issue.

Finally, we did not measure HDL function prior to exposures. The cross-over design of our study assumes that pre-exposure values are the same between study days. This has been the case in prior experiments for other health outcomes (Brook et al 2009). It is possible that unmeasured pre-exposure differences in HDL functionality might have obscured an impact of CAP compared to FA; however, we find this unlikely given the negative results (p-values) and reasonably large sample size for a controlled human exposure study.

Conclusions

In this first study to investigate the effect of air pollution on HDL function in humans, brief inhalation of coarse PM from a rural location did not acutely impair several facets of HDL functionality. Future studies should investigate whether coarse PM of differing characteristics, fine particles, or longer-term exposures can promote HDL dysfunction.

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Figure 1. HDL-mediated cholesterol efflux activity results 2-hours (A, B [with cAMP added]) and 20-hours (C, D [with cAMP added]) post CAP versus FA exposures (n=29)

CAP, coarse concentrated ambient particles; FA, filtered air; cAMP, 0.3 mM 8-chloroadenosine 3',5'-cyclic-monophosphate

These experiments evaluated the ability of subjects' HDL particles to cause efflux of H³-cholesterol from mouse macrophages.

P values are calculated by paired Wilcoxon Signed Rank tests

Figure 2. HDL oxidation index (A), adjusted HDL oxidation index (B), and PON activity 20 hours following CAP versus FA exposures (n=29)

CAP, coarse concentrated ambient particles; FA, filtered air; PON, paraoxonase

HDL anti-oxidant capacity was assessed by a fluorescence assay that evaluated the ability of HDL to inhibit LDL oxidation, determined by dichlorofluorescein (DCF) fluorescence.

PON activity was determined as the hydrolysis of paraoxon substrate (diethyl-*p*-nitrophenyl phosphate) to *p*-nitrophenol by PON-1 by spectrophotometry.

P values are calculated by paired Wilcoxon Signed Rank tests

Table 1

Subject Characteristics (n=32; 16 female subjects)

Variable	Mean	SD	Minimum	Maximum
Age (years)	25.9	6.6	18.0	46.0
Weight (kg)	78.4	16.3	55.9	111.4
Height (m)	1.7	0.1	1.6	2.0
Body mass index (kg·m ⁻²)	26.3	5.7	18.3	43.5
Fasting glucose (mg·dL ⁻¹) *	86.9	6.9	70.0	103.0
Total cholesterol (mg·dL ⁻¹)	163.9	31.4	104.0	244.0
HDL-C (mg·dL ⁻¹)	55.4	15.9	25.0	91.0
LDL-C (mg·dL ⁻¹)	88.8	26.1	49.0	135.0
Triglycerides (mg·dL ⁻¹)	106.0	80.9	40.0	401.0

HDL-c, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; SD, standard deviation

* Missing 1 subject data point

Table 2PM_{10-2.5} concentrations during exposures (n=32)

Exposure	Mean	SD	Minimum	Maximum
Filtered air	10.1	7.1	2.6	27.4
Coarse CAP	76.2 [*]	51.5	10.3	246.5

CAP, concentrated ambient particles; SD, standard deviation

PM_{10-2.5} concentrations are in $\mu\text{g}\cdot\text{m}^{-3}$ and determined by Teflon filter-based gravimetric mass measurements for the 2-hour period of exposures.Mass levels below the detection limit ($6.8 \mu\text{g}\cdot\text{m}^{-3}$) were recorded at this value for analyses (n=15, filtered air exposures only)^{*} P values <0.01 for differences of mean or median levels between exposure types

Table 3

HDL reverse cholesterol transport activity (n=29)

Outcome	Coarse CAP Exposure		Filtered Air Exposure	
	immediate-post	20-hr post	immediate-post	20-hr post
HDL efflux capacity without cAMP (%) *	16.60 (15.17, 19.19)	14.90 (12.47, 19.15)	17.56 (13.43, 20.98)	17.75 (13.22, 23.95)
HDL efflux capacity with cAMP (%) *	18.19 (14.75, 22.11)	17.61 (13.22, 20.67)	17.71 (15.10, 22.12)	19.81 (13.32, 22.10)

Results are median (first, third quartile)

cAMP, 0.3 mM 8-chloroadenosine 3',5'-cyclic-monophosphate

* HDL efflux with cAMP (added stimulus) assessed ABCA1 and ABCG1-mediated RCT activity together, while results without cAMP predominantly assessed ABCG1-mediated RCT activity alone.

Table 4

HDL oxidation index and paraoxonase activity results after exposures (n=29)

Outcome	Coarse CAP Exposure	Filtered Air Exposure
HOI	0.26 (0.24, 0.35)	0.28 (0.25, 0.40)
Adjusted HOI*	0.29 (0.24, 0.35)	0.30 (0.24, 0.39)
Paraoxonase activity ($\mu\text{mol}\cdot\text{min}^{-1}\cdot\text{ml plasma}^{-1}$)	0.54 (0.39, 0.82)	0.60 (0.40, 0.85)

Results are median (first, third quartile)

Lab values were collected and analyzed only 20 hours post-exposures.

HOI, HDL oxidation index; CAP, concentrated ambient particles

Table 5

Correlations between metrics of HDL function (n=29)

	HDL efflux capacity with cAMP	HDL efflux capacity without cAMP	HOI	Paraoxonase
HDL efflux capacity with cAMP		0.762 (p<0.001)	-0.197 (p=0.106)	0.116 (p=0.290)
HDL efflux capacity without cAMP			-0.139 (p=0.334)	0.062 (p=0.501)
HOI				-0.065 (p=0.846)

cAMP, 0.3 mM 8-chloroadenosine 3',5'-cyclic-monophosphate; HOI, HDL oxidation index