Published in final edited form as:

Science. 2013 July 26; 341(6144): 399–402. doi:10.1126/science.1234907.

A secreted PTEN phosphatase that enters cells to alter signaling and survival

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Abstract

Phosphatase and Tensin Homologue on chromosome Ten (*PTEN*) is a tumor suppressor and an antagonist of the phosphoinositide-3 kinase (PI3K) pathway. We identified a 576-amino acid translational variant of *PTEN*, termed PTEN-Long, that arises from an alternative translation start site 519 bp upstream of the ATG initiation sequence, adding 173 N-terminal amino acids to the normal PTEN open reading frame. PTEN-Long is a membrane permeable lipid phosphatase that is secreted from cells and can enter other cells. As an exogenous agent, PTEN-Long antagonized PI3K signaling and induced tumor cell death *in vitro* and *in vivo*. By providing a means to restore a functional tumor suppressor protein to tumor cells, PTEN-Long may have therapeutic uses.

PTEN (Phosphatase and tensin homologue on chromosome ten) is a tumor suppressor that is mutated in multiple types of cancer (1,2). *PTEN* encodes a dual-specificity phosphatase,

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Supplementary Materials

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Materials and Methods

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whose primary substrate is phosphatidylinositol 3,4,5 trisphosphate (PIP3) (3–7). Through this activity, PTEN antagonizes phosphoinositide 3-kinase (PI3K) signaling and thereby affects a myriad of cellular processes including growth, proliferation, and survival (8–11). In mice, loss of *Pten* in tumors or the tumor microenvironment results in neoplastic growth (12–14) indicating that *PTEN*'s tumor suppressive functions are not confined to tumor cells alone.

Inspection of the *PTEN* mRNA transcript (Fig. 1A) revealed an alternative translation initiation codon (CUG) at bp 513 that was 5' of and in-frame with the canonical translation initiation codon (AUG) at bp 1032. Alternative translation beginning at bp 513 was predicted to encode a 576-amino acid translational variant, which we termed PTEN-Long. PTEN-Long contains a 173-amino acid domain at its N-terminus followed by the classical 403 amino acids of PTEN (Fig. S1)(15). Search of the Catalogue of Somatic Mutations in Cancer (<http://www.sanger.ac.uk/genetics/CGP/cosmic/>) database found five somatic missense mutations specific for PTEN-Long in tumor samples (Fig. S1). Alignment of the predicted 173 amino acid sequence of the PTEN-Long unique region with predicted protein sequences from other species indicated that it is evolutionarily conserved (Fig. S2).

To determine if PTEN-Long is translated in cells we transfected human breast (BT549) and glioblastoma (U87MG) cells that lack PTEN with a cDNA expression vector containing the PTEN-Long and PTEN open reading frames. Immunoblots of these lysates revealed two forms of PTEN: a ~75-kD form, PTEN-Long, and a ~55-kD species that corresponded to the conventional translated protein. Expression of PTEN-Long was increased by mutating the alternate start site (CTG) to a classical start site (ATG) (16), was not diminished when the canonical ATG was mutated to ATA, and was lost if the alternatively translated region was deleted or a frame shift was introduced between the alternate start site at 513 and the classic start site at 1032 (Fig. S3). PTEN-Long showed a different enzymatic profile across the tested range of lipid substrate concentrations (soluble di-C8-phosphatidylinositol-3,4,5-trisphosphate (di-C8-PIP3)) (Fig. 1B, Fig. S4) ($p < 0.001$ by ANOVA) (17). A missense mutation in the phosphatase domain of PTEN-Long (G302R), analogous to PTEN (G129R), a tumor mutation, decreased the phosphatase activity of the protein (Fig. S5)(1, 18, 19). Immunoblots of U87MG cells overexpressing equivalent amounts of transfected PTEN, PTEN-Long, or their mutated analogues confirmed that, similar to PTEN, PTEN-Long decreased signaling through the PI3K pathway in a phosphatase-dependent manner (Fig. S6). Immunoblots of mouse embryonic stem cells and human cancer cell lines with antibodies either specific for PTEN-Long or an epitope shared by PTEN and PTEN-Long showed that a ~75 kD PTEN-Long band was present in wild-type but not cells lacking PTEN (Fig. 1C). Immunoprecipitations with PTEN or PTEN-Long antibodies confirmed that PTEN-Long contains canonical PTEN peptides and is therefore a translational variant of PTEN (Fig. 1D, Fig. S7). Amounts of PTEN-Long were reduced in primary human breast tumors compared to those of matched normal breast samples (Fig. 1E, Fig. S8, Table S1). Immunohistochemistry of brain tissue from a mouse glioblastoma caused by the deletion of *Pten*, *p53*, and overexpression of *PDGF* revealed that PTEN-Long was more abundant in the tumor microenvironment than in the tumor or normal tissue (Fig. S9)(20). PTEN-Long was similarly abundant in the tumor microenvironment in 4 of 50 samples from primary breast cancers (Fig. S10).

Computer modeling indicated that PTEN-Long contained a secretion signal sequence with a predicted cleavage site at amino acid 22 (Fig. S11)(21). Thus, we tested whether PTEN-Long might exist outside of cells. We overexpressed V5 epitope tagged PTEN, PTEN-Long, and a mutant with an altered signal sequence predicted to abolish secretion (PTEN-Long Δ A⁶) (Fig. S12) in human embryonic kidney (HEK293) cells. We concentrated the proteins from conditioned medium by immunoprecipitation with PTEN (138G6) antibody or

with heparin columns (Fig. S13). With antibodies that recognized the V5 tag or the endogenous C terminal region of PTEN and PTEN-Long, we detected PTEN-Long but not PTEN nor PTEN-Long Δ A⁶ in conditioned medium (Fig. 2A, Fig. S14). Brefeldin-A, an inhibitor that interferes with retrograde transport of vesicles in the endoplasmic reticulum, inhibited the secretion of PTEN-Long (Fig. 2B)(22). Endogenous PTEN-Long from HEK293 cells bound to concanavalin A-sepharose, a hallmark of a secreted glycoprotein (Fig. S15). Furthermore, PTEN-Long but not PTEN was present in both human plasma and serum (Fig. S16). A screen for PTEN interacting proteins identified multiple heparan-sulfate-modified cell surface proteins in the glypican and syndecan families (Fig. S17)(23). Thus PTEN-Long appears to be secreted from cells and to interact with cell surface proteins.

We noted the presence of a poly-arginine stretch in the unique region of PTEN-Long that was evolutionarily conserved (Fig. S2) and bore a resemblance to the poly-basic residues of the cell-penetrating element of the HIV transactivator of transcription (TAT) protein (24, 25). To determine whether this sequence conferred similar properties to PTEN-Long, we constructed a PTEN-Long Δ R⁶ construct in which these six arginines were deleted (Fig. S18). After treating cells with 100 nM purified Red Fluorescent Protein (RFP)-V5/His, PTEN-Long-RFP-V5/His, or PTEN-Long Δ R⁶-RFP-V5/His, we detected PTEN-Long-RFP but not RFP nor PTEN-Long Δ R⁶-RFP in the cells by fluorescence microscopy (Fig. 3A, Fig. S19). We confirmed this observation through subcellular fractionation (Fig. 3B, Fig. S20). Fusion of the PTEN-Long unique region (PL) to red fluorescent protein (RFP) enabled cellular uptake of the fusion protein (Fig. S21). Therefore the PL domain facilitates cell penetration of tethered peptide sequences. We next tested whether PTEN-Long affected cellular signaling as an exogenous agent. Treatment of cells in culture for fifteen minutes with purified PTEN-Long reduced basal phosphorylation of the protein kinase AKT on Thr 308 in the majority of cell lines (Fig. 3C, Fig. S22–23). When starved U87MG cells were incubated with PTEN-Long for 10 minutes, subsequent stimulation of phosphorylation of AKT Thr 308 by insulin or epidermal growth factor (EGF) was inhibited. Inhibition of PI3K signaling appeared to be dependent upon the poly-arginine sequence of PTEN-Long because the PTEN-Long Δ R⁶ mutant did not block Insulin or EGF-induced phosphorylation of AKT (Fig. S24). Dose response experiments in cells deprived of serum and treated for 24 hours showed that PTEN-Long decreased PI3K signaling as indicated by the decreased phosphorylation of AKT, FOXO, and PRAS40, and induced cell death as indicated by cleavage of caspase 3 (Fig. 3,D–E). We confirmed this effect on cell survival by treating U87MG and MDA-MB-468 cells grown in 0.1% serum with various doses of PTEN-Long or PTEN-Long Δ R⁶ (Fig. 3F).

Upon treatment of mice with PTEN-Long we observed in blood a transient increase in glucose concentration (Fig. S25) and PTEN-Long (Fig. S26). Similar to what we observed in culture, PTEN-Long was detected in mouse tissues, including a xenografted breast tumor (MDA-MB-468), and could alter cell signaling (Fig. S27–29). PTEN-Long derivatives lacking phosphatase or cell penetrating activity were unable to cause these signaling changes. TAT-PTEN only recapitulated some of the effects of PTEN-Long indicating that the PTEN-Long unique region may have other functions beyond the cell permeability conferred by TAT (Fig. S30).

We further explored the effect of PTEN-Long in murine tumor models. We engrafted U87MG cells subcutaneously into athymic nude mice and allowed them to grow until their tumor volume reached approximately 0.2 cm³. Animals were then injected intraperitoneally with PTEN-Long (4 mg/kg) or a control preparation. PTEN-Long caused tumor regression after four days of treatment (Fig. 4A, Fig. S31). Similar regressions were also observed in multiple other xenograft models with the exception of the human colon cancer tumor cell line HCT116 (Fig. S32–33). Although both PTEN-Long and PTEN-Long G302R were

detected within tumor tissues after five days of treatment, only the wild-type protein affected PI3K signaling or induced apoptosis (Fig. 4B). Immunohistochemistry of serial sections of tumor xenografts confirmed exogenous PTEN-Long within tumor cells and a concomitant reduction of pAKT staining. We also treated syngeneic allografts from a mouse model of glioblastoma (GBM). After treatment with PTEN-Long (4 mg/kg), tumors derived from *Pten*^{-/-}, *p53*^{-/-}, PDGF overexpressing GBM cells underwent complete tumor regression within five days, whereas treatment with PTEN-Long Δ R⁶ did not prevent tumor growth (Fig. 4C) (20). Mutant PTEN-Long proteins derived from three cancers (Fig. S1) in which PTEN-Long underwent somatic mutation all showed reduced ability to affect signaling *in vitro* and *in vivo*, suggesting there may be selective pressure to mutate PTEN-Long during tumor development (Fig. S34). Thus PTEN-Long protein appears to alter PI3K signaling *in vivo* to inhibit tumor growth and this effect is dependent on its entrance into cells.

PTEN-Long is a translational variant of *PTEN* that, like classical PTEN, acts as an antagonist of the PI3K pathway. As a secreted product of a tumor suppressor gene capable of entering cells, endogenous PTEN-Long may contribute to an organism's maintenance of signaling, tissue homeostasis and suppression of cancer. Recombinant PTEN-Long and derivative fusion proteins may be useful for delivering proteins in a variety of experimental and clinical settings.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

We thank L. Greene, T. Ludwig, D. Yamashiro, K. Olive and members of the Parsons laboratory for their assistance. Supported by NIH CA082783 NCI CA097403, NIH R01NS066955, the Avon Foundation, Octoberwomen Foundation. BH is supported by 2T32 CA09503. Columbia University has applied for patents on PTEN-Long. NM_000314.4 is curated by GenBank. Additional data are present in the supporting online material.

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Fig. 1.

PTEN-Long is a translational variant of PTEN with lipid phosphatase activity. (A) Shared PTEN/PTEN-Long mRNA (NM_000314.4; orange) with predicted coding regions of PTEN (red) and PTEN-Long (blue). (B) Phosphatase activity assay. 40 nM PTEN and PTEN-Long, assayed with di-C₈-PIP₃ for lipid phosphatase activity, error bars indicate SEM of three replicates ($p < 0.001$, ANOVA), Regression lines to Michaelis-Menten kinetics with V_{max} and 95% Confidence interval (CI) shown in inset moles PO₄/min/moles enzyme. This is a representative experiment of 3. (C) Immunoblot of whole cell lysates from wildtype and *Pten* null embryonic stem cells and cancer cell lines of known *PTEN* status. PTEN-Long (Gly-2) recognizes an epitope specific to the PTEN-Long unique region. PTEN (138G6) recognizes an epitope that is common to both PTEN and PTEN-Long. (D) Reciprocal immunoprecipitations of proteins from whole cell lysates of HEK293 cells with PTEN-Long

specific antibody, probed with an antibody that recognizes an epitope that is common to PTEN/PTEN-Long and vice versa. Input lanes are from longer exposure of the same membrane. Arrows indicate PTEN-Long. (E) Immunoblots of four, randomly selected PTEN wild-type sets of matched breast tumor (T)/normal breast (N) pairs.

Fig. 2. Secretion of PTEN-Long. (A) PTEN, PTEN-Long, and PTEN-Long ΔA^6 from serum free conditioned medium of transfected HEK293 cells eluted from a heparin column or immunoprecipitated with C-Terminal PTEN Antibody 138G6. (B) Cellular (lanes 3 and 4) or secreted PTEN-Long from culture medium (lanes 1 and 2) of transfected HEK293 cells treated with or without brefeldin A at 1 μ g/ml. Arrow indicates PTEN-Long.

Fig. 3.

Entry of PTEN-Long into cells and inhibition of PI3K signaling. (A) Direct fluorescence of U87MG cells 30 min after treatment with 100 nM of either red fluorescent protein (RFP), PTEN-Long-RFP, or PTEN-Long ΔR^6 -RFP. (B) Subcellular fractionation of MDA-MB-468 cells 1 hour after 25 nM treatment with indicated purified proteins. C and N indicate cytoplasmic and nuclear fractions, respectively. Offset, immunoblot of input protein samples used to treat the cells. (C) Effect of 25 nM PTEN-Long at 15 minutes upon pAKT(308). (D) Immunoblots of U87MG and (E) MDA-MB-468 cell lysates 24 hours after treatment with indicated doses of PTEN-Long. (F) Cell viability assay. U87MG and MDA-MB-468 cells were treated with the indicated doses of PTEN-Long or PTEN-Long ΔR^6 for 24 hours before

scoring of the fraction of cells with trypan blue staining. Assayed in quadruplicate, error bars indicate SEM.

Fig. 4.

Effects of PTEN-Long on signaling and tumor development in mice. (A) Graph of U87MG tumor volumes as measured by calipers and treated with either PTEN-Long (4 mg/kg) or an equal volume of mock purified material (N= 5 mice/treatment, error bars indicate $\pm$ SD). (B) Immunoblots of markers of PI3K signaling in U87MG xenografts treated for 5 days with PTEN-Long or phosphatase mutant PTEN-Long(G302R). (C) *Pten*^{-/-} *p53*^{-/-} PDGF overexpressing cells derived from the genetically engineered mouse model of GBM were allografted into the flank of syngeneic hosts and treated with 4 mg/kg of PTEN-Long or PTEN-Long ΔR^6 for 5 days. Tumor volume was assessed daily by caliper measurements and on days 0 and 5 with xenogen imaging system as shown. After 5 days of treatment the mice

in the PTEN-Long ΔR^6 (red arrows) cohort were then treated with PTEN-Long (green arrows) followed with caliper measurements. Blue arrows indicate initial PTEN-Long treatment (days 0–4).