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Application of a real-time PCR method for detecting and monitoring hookworm *Necator americanus* infections in Southern China

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PEER REVIEW

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Comments

This study developed a real-time PCR method for the hookworm *N. americanus*. The method is based on the ITS sequence and proved to be specific for this parasite species. The authors have also shown that it is more sensitive than standard PCR. It can be applied to epidemiology studies to detect hookworm prevalence and infection intensities in infected individuals.

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ABSTRACT

Objective: To develop a quantitative PCR method for detecting hookworm infection and quantification. **Methods:** A real-time PCR method was designed based on the intergenic region II of ribosomal DNA of the hookworm *Necator americanus*. The detection limit of this method was compared with the microscopy-based Kato-Katz method. The real-time PCR method was used to conduct an epidemiological survey of hookworm infection in southern Fujian Province of China. **Results:** The real-time PCR method was specific for detecting *Necator americanus* infection, and was more sensitive than conventional PCR or microscopy-based method. A preliminary survey for hookworm infection in villages of Fujian Province confirmed the high prevalence of hookworm infections in the resident populations. In addition, the infection rate in women was significantly higher than that of in men. **Conclusions:** A real-time PCR method is designed, which has increased detection sensitivity for more accurate epidemiological studies of hookworm infections, especially when intensity of the infection needs to be considered.

KEYWORDS

Hookworm, Detection method, Epidemiology, Infection rate

1. Introduction

Hookworm infection, caused by the soil-transmitted helminths *Necator americanus* (*N. americanus*) and *Ancylostoma duodenale* (*A. duodenale*), is one of the most common chronic infections among the world's poorest populations living in developing countries, with up to 576 million people currently infected^[1,2]. Of the two principal species of hookworm infecting humans, *N. americanus* is found throughout sub-Saharan Africa, tropical regions of the Americas, South China, and Southeast Asia^[2]. Although

rarely directly lethal, hookworm infection is a significant factor of morbidity, associated with chronic anemia and protein malnutrition. When measured in disability-adjusted life years, hookworm infection is the second most important parasitic infection, just behind malaria^[3]. In Southern China, hookworm is one of the three major soil-transmitted helminths^[4,5]. According to a report in 2003, there was an estimate of 39 million people being infected. The predominant hookworm species in Southern China like Hainan province is *N. americanus*^[6,7].

Diagnosis of hookworm infection is usually based on

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detection of hookworm eggs in human fecal samples[8]. However, due to similarity in morphology, hookworm eggs cannot be reliably identified to species. Although the method of coproculture for obtaining third-stage larvae allows more accurate microscopic identification of the worms, this approach is laborious, unreliable, and requires skilled personnel[9]. Given that accurate specific diagnosis of human hookworm infections is essential for understanding the epidemiology and designing targeted control strategies, molecular methods for the identification of hookworms and other strongylid nematodes have been developed[9,10]. Interspecific differences in nuclear DNA and mitochondrial DNA (mtDNA) have been employed to design PCR-based diagnostic methods. Using differences in *cox1* gene of the mtDNA, Zhan *et al.* achieved specific identification of the two major hookworm species—*N. americanus* and *A. duodenale*[11]. The differences in the cAMP-dependent protein kinase gene also allowed the development of a PCR-restriction fragment length polymorphism method for species differentiation[12]. However, methods based on the internal transcribed spacers (ITS-1 and 2) of the nuclear ribosomal DNA (rDNA) have received the widest applications because of their lower mutation rates within species and substantial differences between species[13,14]. Species-specific PCR primers have been designed to differentiate *N. americanus* from *A. duodenale*, and the method proved useful in epidemiological survey of hookworm infections[15,16]. In recent years, quantitative real-time PCR has been used in parasitology to monitor the prevalence of geohelminths[17,18]. Here, we attempt to develop a real-time PCR method to specifically identify *N. americanus* infection in humans and evaluate this method in epidemiological surveys of hookworm infection in Southern China.

2. Materials and methods

2.1. Hookworm DNA

To collect *N. americanus* hookworm samples, human stool samples were collected in a suburban district of Xiamen City, Fujian Province, China. The samples were examined using a flotation method with saturated NaCl, and helminth eggs were examined microscopically. Five grams of a positive stool sample were used for coproculture at 27 °C, and *N. americanus* third-stage larvae were identified morphologically using published keys and descriptions[19]. A total of 50 *N. americanus* third-stage larvae were collected for DNA extraction using the WizardTM DNA isolation kit (Promega, Madison, USA). For comparison, genomic DNA of *Strongyloides stercoralis* (*S. stercoralis*) and *Ancylostoma caninum* (*A. caninum*) were extracted similarly from samples maintained in our laboratory.

2.2. PCR and sequencing of ITS-2 of *N. americanus* rDNA

For PCR amplification of the ITS-2 sequence of *N. americanus* rDNA from a Southern China isolate, two primers NAF (5'-TGT TCA GCA ATT CCC GTT TA-3') and NAR (5'-GTC CTT CAC ATT GTC TCC GT-3') were designed based on a GenBank sequence (accession number Y11734). A 50 µL PCR reaction was assembled with 5 µL of 10×PCR buffer, 3 µL of 25 mmol/L MgCl₂, 4 µL of 2.5 mmol/L dNTPs, 1.5 µL of each primer (10 µmol/L), 1 U of Taq polymerase (TaKaRa), and 10 ng of nematode DNA. PCR was performed using the following conditions: initial denaturing at 95 °C for 5 min followed by 35 cycles of 30 seconds at 95 °C, 30 seconds at 55 °C, 40 seconds at 72 °C, and final extension at 72 °C for 7 min. Five microliters of the PCR reaction were electrophoresed in 15 g/L agarose gel. The band was excised from the gel and DNA was extracted using the QIAquick gel extraction kit (Qiagen, Valencia, USA). The DNA was cloned into pMD18-T and the insert was sequenced using the BigDye Terminator version 3.1 Cycle Sequencing Kit on an ABI3100 Genetic Analyzer. DNA sequence was BLAST searched in the GenBank.

2.3. Development of a real-time PCR method

A real-time PCR method of detection for hookworm *N. americanus* was developed based on the SYBR Green quantification kit (Tiangen Biotech, Beijing, China) on a Rotor Gene 3000 PCR machine. The real-time PCR program included 40 cycles of 15 seconds at 95 °C, 15 seconds at 56 °C, and 18 seconds at 72 °C. The reactions were set up in a final volume of 20 µL with 8 µL of 2.5×RealMasterMix, 0.8 µL of each primer (10 µmol/L), and 2 µL of the template DNA. Initially, we tested the recombinant plasmid DNA in 10-fold serial dilutions with concentrations ranging from 10² to 10⁸. Fluorescent PCR products were detected at 72 °C. The threshold cycle, C_t, was defined as the cycle number at which the change of fluorescence in the reaction exceeds ten standard deviations above the background fluorescence. Background fluorescence was calculated as the mean fluorescence between cycle 3 and 6. To determine the sensitivity and reproducibility of the real-time PCR method, we used a dynamic range of the template concentrations. Each concentration was tested for at least six times and covariance (standard deviation/mean) of the C_t was determined.

To further determine the detection sensitivity, *N. americanus* eggs were harvested, washed three times with physiological saline and the egg number was counted under a microscope. A total of 1, 5, 10, 50, and 100 *N. americanus* eggs were added to separate microcentrifuge tubes containing 200 mg of fecal materials from a healthy person. DNA was isolated from the mixture using the QIAamp DNA Stool Kit and subjected to real-time PCR analysis.

2.4. Epidemiological survey

In July and August of 2009, we collected 216 stool samples from residents above one year of age from two villages in Canghai District of Xiamen City. The samples were stratified by genders of the residents. Three methods were compared for their sensitivity to detect hookworm infections. First, about 2 g of each stool sample was subjected to floatation in 265 g/L NaCl, and hookworm eggs were quantified using the Kato–Katz method[20]. For molecular diagnosis, DNA was isolated from 2 g of the stool samples using the QIAamp DNA Stool Kit (Qiagen). Regular PCR was performed as described above for 35 cycles, and PCR products were electrophoresed in 15 g/L agarose gel and visualized under UV light. In parallel, the newly developed real-time PCR method was used to detect hookworm DNA in the stool samples. χ^2 test was used to determine whether the detection rates of the three methods were significantly different.

3. Results

To develop a species-specific molecular detection method, we designed a pair of primers NAF and NAR based on the rDNA ITS-2 sequence of *N. americanus* in the GenBank. The PCR specifically amplified a 206 bp fragment from DNA extracted from *N. americanus* larvae, and sequence of the PCR product completely conformed to the *N. americanus* ITS-2 sequence deposited in GenBank (Y11734). In contrast, no PCR product was detected when genomic DNA from either *S. stercoralis* or *A. caninum* was used as the template (data not shown), indicating that the primers are specific for *N. americanus*.

The same primers NAF and NAR were used to develop a SYBR Green-based real-time PCR method, which also specifically amplified the ITS-2 sequence from *N. americanus*, but not from *S. stercoralis* or *A. caninum* (not shown). The PCR product displayed a temperature of 81 °C, and the melting curve had only one peak, suggesting that the PCR primers were specific without generating non-specific PCR products or formation of primer dimers. We then used a wide range of DNA concentrations (10^2 to 10^8 copies/reaction) of the ITS-2 plasmid to determine the standard curve of the reactions. As shown in Figure 1, the C_t values and template concentrations displayed excellent linear correlation ($R^2=0.993$), indicating that the method could be applied to quantify the template DNA in a wide range of concentrations.

Figure 1. Standard curve of the real-time PCR established by using the recombinant plasmid as the template.

To determine whether the results from different concentrations of plasmid were reproducible, the reactions were replicated six times and the C_t values were determined. From a series of plasmid concentrations (10^2 to 10^8 copies/reaction), the reproducibility of the method was excellent with insignificant amount of variation (covariance below 3%) (Table 1). Using the ITS-2 plasmid, we compared the sensitivity between the real-time PCR and conventional PCR method. The real-time PCR method reliably detected ITS-2 plasmid at 10^2 copies/reaction, whereas regular PCR could only produce visible DNA bands above 10^4 copies/reaction, indicating that the real-time PCR method was at least 100 times more sensitive than the regular PCR method (Figure 2). To further confirm the increased sensitivity of the real-time PCR method, we extracted whole DNA from fecal materials from a healthy person mixed with different numbers of *N. americanus* eggs (1–100). We have confirmed that the real-time PCR method could detect as few as one egg in 200 mg of fecal materials (data not shown).

Table 1
Reproducibility of the real-time PCR for detection of *N. americanus*.

Template (copies/reaction)	C_t values	Covariance (%)
6.02×10^8	8.77 ± 0.12	1.37
6.02×10^7	12.78 ± 0.24	1.88
6.02×10^6	15.37 ± 0.17	1.15
6.02×10^5	18.62 ± 0.15	0.80
6.02×10^4	23.58 ± 0.16	0.68
6.02×10^3	27.28 ± 0.23	0.84
6.02×10^2	32.19 ± 0.35	1.09

Data are expressed in mean $\pm$ SD.

Figure 2. Comparison of sensitivity for detection of *N. americanus* between real-time and conventional PCR.

Inset: conventional PCR (35 cycles). The recombinant plasmid concentrations (copies/reaction): 1, 1×10^8 ; 2, 1×10^7 ; 3, 1×10^6 ; 4, 1×10^5 ; 5, 1×10^4 ; 6, 1×10^3 ; 7, 1×10^2 . M indicates molecular size markers.

We have performed a survey to determine hookworm prevalence in two villages of Xiamen using three detection methods: Kato–Katz method, regular PCR, and real-time PCR. In stool samples collected from 216 residents, the Kato–Katz method detected hookworm infections in 30 people (13.89%). In addition to these 30 people, regular PCR method detected infections in additional 5 people, yielding a positive rate of 16.20%. Like the regular PCR method, the real-time PCR method detected the same 35 positive

samples. Remarkably, additional 14 samples were positive with the real-time PCR method, which resulted in the detection of hookworm infections in 49 people (22.69%), which was significantly higher than the other two methods ($\chi^2=6.20$, $P<0.05$) (Figure 3). In our study, we also found that the prevalence of hookworm infections differed significantly ($\chi^2=4.60$, $P<0.05$) between the two villages. When the infection rates were compared between male and female residents, the infection rate in females (34.20%) was three times higher than that in males (10.50%) ($\chi^2=17.37$, $P<0.05$).

Figure 3. Hookworm infection rates in two villages of Xiamen (Dongfu and Xinyang), Fujian Province, China as detected by the real-time PCR method.

4. Discussion

Hookworm *N. americanus* is the most prevalent species of helminth infections in Southern China[6,7]. Here, we designed a real-time fluorescent PCR method to specifically detect hookworm infections in humans in southern China. This real-time PCR method had significantly higher sensitivity in the diagnosis of hookworm infections than the traditional PCR method or the microscopy-based Kato-Katz method. It is anticipated that the more sensitive real-time PCR method will offer higher levels of detection of hookworm infection, especially when infection intensity is low. In addition, this method can be used to perform quantitative measure of the intensity of hookworm infection in endemic areas.

The prevalence of hookworm infections differs dramatically between geographic locations, age and sex of the human populations[6,21,22]. Past surveys for hookworm infections conducted in southern provinces such as Yunnan, Hainan, and Sichuan detected high infection rates in rural populations[6,21,22]. Hookworm infection was significantly more prevalent in elderly people[6,7]. We conducted an

epidemiological survey in two villages of Xiamen and demonstrated that hookworm infection remains highly endemic in Fujian Province of Southern China. These infection rates were similar to those reported earlier in Southern China[23–26], but lower than two more recent reports[6,7,21]. Infection rates were different between the two villages, which may reflect different ecology of hookworm infections at these sites. Intriguingly, the infection rate in women was significantly higher than that of in men, which was consistent with an earlier observation in Hainan province[6]. However, the reasons for the differential infection rates in men and women remain unknown. With the development of this real-time PCR method, we hope future epidemiology of hookworm infection will not only focus on infection rates but also the intensity of infection.

Conflict of interest statement

We declare that we have no conflict of interest.

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Comments

Background

Hookworm infection is a public health problem in developing countries in the tropical and subtropical regions. In Southern China, hookworm infection is highly prevalent, especially in the countryside. Microscopy-based Kato-Katz method is the standard for diagnosis, but the detection sensitivity is low. More sensitive PCR diagnostic methods have been developed using mitochondrial DNA sequence and internal transcribed spacers of the nuclear ribosomal DNA.

Research frontiers

PCR diagnostic methods were developed for major hookworm species based on mitochondrial and nuclear ribosomal DNA sequences. The real-time PCR method is specific for *N. americanus*, the most common hookworm species in Southern China, and more sensitive than conventional PCR methods.

Related reports

There are several PCR methods developed, which allow differentiation of different hookworm species (Gasser *et al.*, 2008). This paper developed a more sensitive and

quantitative real–time PCR method for detecting and quantifying hookworm infections.

Innovations and breakthroughs

The major advantages of this real–time PCR method are its ability to allow quantification of infection intensity.

Applications

This method can be used for both detection and quantification of hookworm infection in patients.

Peer review

This study developed a real–time PCR method for the hookworm *N. americanus*. The method is based on the transcribed spacers sequence and proved to be specific for this parasite species. The authors have also shown that it is more sensitive than standard PCR. It can be applied to epidemiology studies to detect hookworm prevalence and infection intensities in infected individuals.

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