Effect of cooking temperatures on protein hydrolysates and sensory quality in crucian carp (*Carassius auratus*) soup

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Abstract Cooking methods have a significant impact on flavour compounds in fish soup. The effects of cooking temperatures (55, 65, 75, 85, 95, and 100 °C) on sensory properties and protein hydrolysates were studied in crucian carp (*Carassius auratus*) soup. The results showed that the soup prepared at 85 °C had the best sensory quality in color, flavour, amour, and soup pattern. Cooking temperature had significant influence on the hydrolysis of proteins in the soup showed by SDS-PAGE result. The contents of water soluble nitrogen (WSN) and non-protein nitrogen (NPN) increased with the cooking temperature, but the highest contents of total peptides and total free amino acids (FAA) were obtained at the cooking temperature of 85 °C. The highest contents of umami-taste active amino acid and branched-chain amino acids were also observed in the 85 °C sample. In conclusion, a cooking temperature of 85 °C was preferred for more excellent flavor and higher nutritional value of crucian carp soup.

Keywords Crucian carp · Cooking temperature · Protein · Hydrolysates · Flavour

Introduction

Crucian carp (*Carassius auratus*), native to China, Russia, and Korea, is one of the most important fresh water fish species (Tang et al. 2008). It is rich in protein, carbohydrates, Vitamin A, B, minerals and so on (Bastić et al. 2002). Crucian carp is usually prepared to make fish soup in many Asian-Pacific countries (Xia and Xu 2005), because the fish has a small figure but delicious taste. It is generally recognized that crucian carp soup is rich in nutrients and also has the function of regulating menstruation, lactogenesis and help the recovery of puerperants.

Flavour is one of the most important characteristics of food quality, particularly for soup/broth products. In general, most flavour components are water-soluble including nucleotides, amino acids, peptides, organic acids, organic bases, and inorganic ions. It is well known that peptides and amino acids are the primary ingredients that contribute to the taste of fish and meat products (Biswas et al. 2005, Shah et al. 2010a). Cambero et al. (2000) analyzed the relationship of compounds with the flavour of beef soup developed at different cooking temperatures. A significant rise ($P<0.05$) in the concentration of nitrogen in HPPN (higher than 600 Da peptidic nitrogen), SNAN (small non-amino acid nitrogen compounds), creatinine, IMP (inosine 5'-monophosphate) and AMP (adenosine 5'-monophosphate) occurred with the increase of cooking temperature. The results also suggested the reduction of PN (protein nitrogen) and increase of SNAN were important to the beef soup flavour. Some other studies on fish soup have been reported. de Godoy et al. (2010) evaluated the acceptance of soups prepared with different kinds of smoked fish carcasses (Nile Tilapia, carp, and pacu). Shah et al. (2010b) isolated flavour-enhancing compounds,

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creatine and creatinine from water-soluble extracts of dried herring fillet.

Fish protein is a very important food protein to human as it provides an excellent balance of amino acids. The hydrolysates of fish protein include flavour peptides, umami-taste amino acids, branched-chain amino acids (BCAA) etc. The BCAAs have been known in reducing fatigue in both anaerobic and endurance sports, which also have been used clinically to aid the recovery of burn victims (Shimomura et al. 2006). Thermal processes for fish products can significantly influence the composition of protein and protein hydrolysates, especially the peptides and free amino acids, ultimately influence the quality of the fish food (Cambero et al. 1998, Pandey et al. 2008).

To our knowledge, there was no literature report on the relationship of the cooking methods and the nutrition and taste components in fresh water fish soup. The objective of this study was to evaluate the effects of cooking temperature on the sensory properties and contents of protein hydrolysates of crucian carp soup, to provide useful information for the preparation of crucian carp soup, both for family cooking and industrialized production.

Materials and methods

Soup preparation Flesh crucian carp (*Carassius auratus*) with mean body weight of 300 ± 50 g were obtained from Century Mart Hypermarkets in Hangzhou City. Thirty individual fish were gutted and washed with tap water. After draining, two pieces of whole fish (460 ± 75 g) were cooked in a reflux with a 0.5% NaCl solution in a proportion of 6:1 (v/w) at temperatures of 55, 65, 75, 85, 95 and 100 °C for 2 h, respectively. The NaCl solutions were heated to the desired temperature before cooking. The cooked soup were cooled down to room temperature and clarified by sieving through a 0.2-mm-mesh sieve. The crucian carp soups obtained were then used in the following analysis.

Sensory evaluation The sensory analysis was carried out following the Morita (Morita et al. 2003) method with some modifications. The samples were randomly distributed to 14 trained panelists (seven females and seven males) and quantitative descriptive analysis was performed. The color, aroma, taste and soup pattern were used as descriptive index and evaluated by using a 15 cm line scale. Each index has its highest score of 30 and the total score was 120. The sample with the best flavour was assigned as the highest score. All samples were evaluated once.

Protein and SDS-PAGE analysis Protein was extracted according to the methods of Bechtel (Bechtel and Parrish

1983). The protein concentration was measured by the Biuret method (Robinson and Hodgen 1940) and adjusted with deionised water to a final concentration of 4 mg/mL. SDS-PAGE was performed according to the procedure of Laemmli (Laemmli 1970), using broad molecular weight standards, and the gels were stained with Coomassie Brilliant Blue.

Nitrogen fraction determination Determinations of nitrogen fraction in crucian carp soup were carried out according to the method reported by Careri et al. (1993). Aliquots (10 mL) of crucian carp soup were mixed with the same volume of 15% phosphotungstic acid solutions to obtain non-protein nitrogen (NPN). Mixtures were kept at room temperature for 60 min, and centrifuged at 5000 rpm for 10 min at 4 °C. The supernatants were obtained by filtration through Whatman No.4 paper. The WSN (water-soluble nitrogen) and NPN were determined by the Kjeldahl method using Kjeldahl equipment (Foss 2300, Denmark). The WSN was considered as the total nitrogen in the present work.

Peptides analysis Peptides were determined following the method described by Liu et al. (2007). A 10 g sample was deproteinized with 0.6 mol/L perchloric acid by homogenizing for 30 s. The homogenates were centrifuged at 15,000 g for 10 min and filtered through Whatman No. 54. After the pH being adjusted to 6 with 30% potassium hydroxide, the filtrates were filtered again. Twenty microlitres of filtrate were used for the HPLC analysis, using a Supelcosil LC-18 containing octadecyldimethylsilyl, 25 cm×4.6 mm (5 µm particle size) from Supelco (Bellafonte, PA, USA) and a UV detector (214 nm). The eluents used were (A) water (HPLC grade) and (B) acetonitrile containing 0.1% of trifluoroacetic acid. The flow rate was 1 ml/min and the following gradient was employed: initial of 3.2% B; 0.5 min linear change to 4.5% B; 5 min linear change to 8.5% B; 10 min linear change to 11.5% B; 22 min linear change to 99% B and kept for 12 min, then equilibrated at 3.2% for 10 min.

Free amino acids analysis The amino acids composition was analyzed according to the report of Norziah and Ching (2000), using the Waters Associates AccQ•Tag method, pre-column derivatization of samples with AccQ•fluor reagent and analysis by reverse phase HPLC. Identification of the amino acids in the sample was carried out by comparing their retention times with the standards.

Statistical analysis Data were analyzed by Duncan's multiple range tests using statistical package statistica V 5.5 software. A significant level was defined as a

probability of 0.05 or less. Determinations were carried out in triplicate.

Results and discussion

Sensory evaluation Figure 1 showed the results of sensory evaluation of crucian carp soups obtained at different cooking temperatures. The scores of color, aroma, flavour and pattern of the crucian carp soups had no linear relationship with the cooking temperatures. The crucian carp soup obtained at 85 °C had the most favorable appearance: the soup color was as white as milk. The soup obtained at 75 °C got a similar good appearance, and its score was next to the samples of 85 °C. The 55 °C one showed the lowest scored in the item of color as the soup looked as clear as water. The 75 °C and 85 °C ones got the highest grade of aroma scores (24.43 and 24.36, respectively), especially its specific fish soup aroma. The samples obtained at 95 °C and 100 °C also got relatively high values in aroma evaluation test, with 19.29 and 18.86 scores, respectively. But, the soups under these two temperatures contained many impurity substances, such as suspended meat mince and fishbone. Flavour is another important index for the sensory quality of fish soup. The strongest delicate flavour was obtained in the soup cooked at 85 °C,

while the lowest flavour score was for 55 °C. A fishy odour was detected in the 65 °C and 55 °C soups, especially in the 55 °C one (Fig. 1).

Cooking temperature had a remarkable influence on the quality of crucian carp soup. When the soups were prepared at high temperatures (95 °C and 100 °C), the proteins on the surface of fish flesh coagulated, leading to the decrease of flavour and taste compounds extracted from flesh. As a result, the soup did not taste well. Furthermore, an extended cooking time caused a breakdown of protein structure, which brought to a bad soup pattern and color of the soups. On the other hand, when the cooking temperatures were lower (55 and 65 °C), the denaturation of muscle protein in the fish flesh might not complete and the soluble protein hydrolysates had not been dissolved sufficiently, thus the flavour and taste compounds were also lower in the soups. In contrary, the soup cooked at 85 °C, had the best quality in color, aroma, flavour and soup pattern.

SDS-PAGE The electrophoretic patterns of proteins in crucian carp soups obtained at different cooking temperatures were shown in Fig. 2. The protein patterns of 55 °C and 65 °C soups were found to be similar with each other, which indicated that the two soup samples have the similar protein fractions. The increase of cooking temperature (75 °C and 85 °C) resulted in the dropping of the intensity of the

Fig. 1 Mean sensory evaluation of the crucian carp (*Carassius auratus*) soup obtained at different cooking temperatures. ($n=3$)

Fig. 2 Electrophoresis pattern of proteins of crucian carp (*Carassius auratus*) soup obtained at different cooking temperatures. (M: Marker; a 55 °C; b 65 °C; c 75 °C; d 85 °C; e 95 °C; d 100 °C)

myosin heavy chain (MHC) and molecular weight of 66–110 kDa bands. This was accompanied by an appearance of low molecular weight bands (14–45 kDa). The degradation of proteins during the cooking process was responsible for the density changes of protein bands. It is well known that high temperature promotes the heat denaturation of proteins. However, the low molecular weight bands did not increase significantly in the higher temperatures. In the present study, it was shown that when the cooking temperatures was 95 °C and 100 °C, the intensity of the heavy molecular weight (HMW) bands (>200 kDa) increased compared to that of 55–85 °C. It could be explained that the formation of protein aggregation occurred in the samples obtained at high temperatures due to protein cross-linking. Furthermore, the content of MHC decreased and some low molecular weights bands occurred as a result of the degradation of proteins in the 95 °C and 100 °C soups. Consequently, the solubility of proteins was reduced, which made contributions to the combination of intermolecular proteins and proteins with other compounds. These resulted in a relative reduction of flavour compounds from fish flesh into the soup. According to the previous research, the color of the soup had a positive correlation with the contents of proteins and lipids, and the milk-white-like soup was caused by high concentration of proteins and lipids (Chidanandaiah et al. 2002, Yamazawa and Koshino 2010). From the above sensory evaluation results, the 85 °C soup had a better milk white color than other samples, which suggested that the concentrations of proteins and lipids in this soup might be higher than those in other ones, and was confirmed by the SDS-PAGE analysis.

Nitrogen fractions analysis Figure 3 showed the changes of nitrogen fractions in the soups cooked at different temperatures. The content of WSN in all soups ranged from 0.38 to 0.57 g/100 mL. An increase in WSN was observed when the temperature was increased. The composition of NPN in the crucian carp soup was the nitrogen of free amino acids, nucleotides, and peptides, which can directly contribute to the formation of flavour or as flavour precursors (Madruga et al. 2010). In the present study, the content of NPN in soups also increased with the increase of cooking temperature. The behaviour of WSN and NPN coincided with that of the SDS-PAGE pattern, which reflected the protein degradation and the release of low-molecular-weight nitrogen compounds (Sun et al. 2010). Similar results were also reported for shrimp soup (Cambero, et al. 1998), which indicated that cooking temperature played a more important role than cooking time on WSN and NPN concentrations.

Peptides analysis Table 1 showed the peptides peaks in different soups by using reverse-phase HPLC. An increase in the chromatography area of the total peptides was observed when the cooking temperature was increased up to 85 °C. However, the total peptides tended to slightly reduce over 85 °C. The increases of total peptides in soups might be attributed to two reasons. One might be the flowing out of original peptides from fish meat into the aqueous soup during cooking. On the other hand, an increase of oligopeptides took place because of the degradation of proteins at higher temperature (Liu and Zhao 2010). However, the total peptides did not continue to increase when the cooking temperature was over 85 °C. As a matter of fact, a decrease of the total peptides occurred in the soups when cooked at 95 °C and 100 °C, implying that the rate of degradation of peptides was higher than that of peptide formation at higher temperatures.

Fig. 3 Concentration (g/100 mL) of water-soluble nitrogen (WSN), non-protein nitrogen (NPN) of crucian carp (*Carassius auratus*) soup obtained at different cooking temperatures. (n=3)

Table 1 Peptide concentrations in crucian carp (*Carassius auratus*) soup cooked at different temperatures

	55 °C	65 °C	75 °C	85 °C	95 °C	100 °C
Peak 1	10.3±0.23c	11.7±0.72c	15.6±0.39b	17.9±1.03a	14.5±0.48b	11.4±0.55c
Peak 2	3.1±0.03 d	4.2±0.11	6.3±0.17b	8.4±0.35a	5.2±0.61c	3.9±0.08 d
Peak 3	1.9±0.05a	1.8±0.02ab	1.7±0.35b	2.1±0.03a	2.0±0.09a	1.9±0.16ab
Peak 4	47.3±1.16e	65.8±1.03c	89.3±1.21b	102.2±3.49a	69.7±1.46c	57.1±1.32 d
Peak 5	5.7±0.22a	4.6±0.02ab	4.1±0.23b	3.2±1.07c	2.8±0.11c	2.6±0.04 d
Peak 6	30.5±1.81c	38.3±0.31b	44.7±0.56a	42.9±2.18ab	31.4±2.2c	29.0±1.1 d
Peak 7	0.9±0.03a	0.78±0.02a	0.64±0.01b	0.57±0.02b	0.49±0.01b	0.26±0.01c
Peak 8	4.2±0.42a	3.29±0.05c	3.9±0.32ab	3.9±0.15a	3.7±0.11b	3.2±0.05c
Peak 9	10.4±0.14a	9.2±0.01a	6.5±0.03c	7.1±1.02b	6.8±0.23b	4.9±0.08 d
Peak 10	7.14±0.92c	10.7±0.03a	9.4±0.15ab	11.5±0.51a	8.8±0.17b	6.4±0.12 d
Peak 11	0.85±0.02 d	1.5±0.07c	3.3±0.02a	2.8±0.06a	2.2±0.02b	1.6±0.18c
Peak 12	0.28±0.01c	0.72±0.02a	0.54±0.01b	0.31±0.01c	0.49±0.01b	0.19±0.01 d
Peak 13	11.3±0.13 d	15.9±0.02c	20.7±0.78b	22.4±1.04a	19.4±0.49b	16.2±1.39c
Total	133.8±2.71 d	168.6±1.98c	206.3±5.82b	225.3±4.37a	167.5±3.97c	138.4±2.64 d

Contents of peptides were in area 100 mL⁻¹ soup and expressed as means±standard error ($n=3$). Means followed by different lowercase letters in the same row are significantly different ($P\leq 0.05$)

The changes of individual peptide areas were different from the changes of total peptides. It was clear that most peptides (Peak 1, 2, 4, 6, 10 and 13) tended to increase as the cooking temperature increased in the range of 55–85 °C (Table 1), yet a decrease was observed above 85 °C. This was in agreement with the results of nitrogen fractions mentioned in section ‘*Nitrogen fractions analysis*’, which demonstrated that a remarkable increase of formation of these peptides due to the proteolytic breakdown in this temperature range. However, some peptides (Peak 5 and 7) underwent a reducing with the increase of temperatures, suggesting the degradation of these peptides was much more severe. Some peptides (Peak 3, 8, 9 and 12) did not have a significantly correlation with the heating temperatures. The slight changes suggested that a balance between degradation and formation of these peptides was existed simultaneously in the soups under different temperatures.

Peptides and FAA are the main flavour compounds and the precursors of flavour compounds (Sentandeu et al. 2003). The free amino acids are the final products of proteolysis, while the peptides are intermediate products (Hashim et al. 1998). Peptides have the buffer capacity due to the amino-group and carboxyl-group, which can not only contribute to the basic flavors directly, but can also help to improve or change food flavors though the interaction with other flavour compounds (Hansen et al. 2000, Toldrá et al. 1997).

Free amino acids analysis Like peptides, free amino acids (FAA) are another important flavour component in fish products. The free amino acids concentrations of crucian carp soups cooked at different temperatures were deter-

mined. As shown in Table 2, the predominant free amino acids in the crucian carp soup were Arg (27%–40%), Glu (5–17%) and Ala (10–12%), followed by Asp (3–10%), Gly (5–8%), Leu (2–4%) and Lys (2–4%). The remaining FAA of Phe, Thr, Tyr, Val, Ile, Cys2, Met, His and Met accounted to less than 3% of the total FAA.

The Glu, Asp, Phe, Ala and Gly, presented in relatively large quantities in soups, belonged to the umami-taste active amino acids, and were very important to the flavour of food (Itou et al. 2006). These amino acids have been considered as the main contributors to the flavour of a soup (Randhawa and Ranote 2004, Shiau et al. 1997). Furthermore, their sodium salts have a synergistic effect on the flavour with nucleotides (Kayim et al. 2011). Table 2 showed the total tasty amino acids in crucian carp soup accounted for more than 50% of the total FAA. The decreases of some amino acids (Glu and Ala) could be explained by the reaction between these amino acids and other compounds. On the other hand, Gly and Phe concentrations increased stably until the cooking temperature increased up to 85 °C; followed by a slight reduction above 85 °C. The highest total tasty amino acids concentration took place in the soup obtained at 85 °C, suggesting that the strongest flavour was obtained in the 85 °C soup. This result was in agreement with that of the sensory evaluation. The Leu, Ile and Val compose the Branched-Chain Amino Acids (BCAAs) which has special physiological functions, such as strengthen the immunity (Wakshlag et al. 2006). The concentration of BCAA accounted about 6–8% of the total FAA (Table 2). The content of BCAA increased when the cooking temperatures increased up to 85 °C. Thereafter, it showed a decreasing tendency in the soups cooked at a

Table 2 Concentration ($\mu\text{mol}/100\text{ mL}$) of free amino acids (FAA) in crucial carp (*Carassius auratus*) soup cooked at different temperatures. ($\mu\text{mol}/100\text{ mL}$)

AA	Temperatures/ $^{\circ}\text{C}$					
	55	65	75	85	95	100
Asp	60.5 $\pm$ 1.31 d	80.3 $\pm$ 5.02 cd	131.6 $\pm$ 3.18b	98.1 $\pm$ 4.33c	197.4 $\pm$ 2.69a	120.4 $\pm$ 8.94b
Ser	48.5 $\pm$ 2.26c	66.3 $\pm$ 3.15a	54.7 $\pm$ 2.99bc	59.8 $\pm$ 1.86b	52.6 $\pm$ 2.11bc	28.5 $\pm$ 0.08 d
Glu	209.1 $\pm$ 7.01a	189.0 $\pm$ 8.43b	181.7 $\pm$ 6.86b	180.3 $\pm$ 5.83b	141.1 $\pm$ 4.26c	72.7 $\pm$ 1.32 d
Gly	107.6 $\pm$ 1.04c	111.3 $\pm$ 1.95c	128.7 $\pm$ 2.88b	138.8 $\pm$ 4.09a	122.5 $\pm$ 6.45b	107.6 $\pm$ 2.87c
His	33.4 $\pm$ 0.06b	19.3 $\pm$ 0.11 d	43.3 $\pm$ 0.37ab	48.3 $\pm$ 1.02a	34.4 $\pm$ 0.06b	29.8 $\pm$ 0.15c
Arg	523.7 $\pm$ 8.93c	648.33 $\pm$ 4.76b	810.7 $\pm$ 7.11a	884.9 $\pm$ 9.93a	839.9 $\pm$ 11.97a	693.3 $\pm$ 4.98b
Thr	30.4 $\pm$ 1.34bc	45.30 $\pm$ 1.08a	35.3 $\pm$ 1.00b	29.3 $\pm$ 0.02bc	21.4 $\pm$ 0.01c	16.9 $\pm$ 0.31 d
Ala	402.8 $\pm$ 3.55a	422.7 $\pm$ 9.83a	291.8 $\pm$ 7.16c	336.0 $\pm$ 6.81b	264.1 $\pm$ 1.06c	138.8 $\pm$ 3.84 d
Pro	27.8 $\pm$ 1.76c	38.5 $\pm$ 1.20ab	34.3 $\pm$ 0.85b	46.1 $\pm$ 0.16a	44.8 $\pm$ 3.76a	25.9 $\pm$ 1.07c
Cys2	1.7 $\pm$ 0.06 cd	1.3 $\pm$ 0.01 cd	13.5 $\pm$ 0.12b	31.1 $\pm$ 0.64a	5.2 $\pm$ 1.07c	0.89 $\pm$ 0.03 d
Tyr	25.0 $\pm$ 0.5bc	35.5 $\pm$ 0.02a	24.5 $\pm$ 1.05bc	30.7 $\pm$ 0.67b	22.4 $\pm$ 1.63c	11.3 $\pm$ 4.42 d
Val	25.6 $\pm$ 2.89c	47.4 $\pm$ 6.07b	47.1 $\pm$ 5.32b	71.7 $\pm$ 3.88a	48.5 $\pm$ 1.46b	26.4 $\pm$ 5.00c
Met	40.3 $\pm$ 0.73c	82.3 $\pm$ 2.89b	102.3 $\pm$ 4.93a	108.5 $\pm$ 5.91a	31.6 $\pm$ 6.11 d	10.3 $\pm$ 0.22e
Lys	37.8 $\pm$ 0.79 d	49.6 $\pm$ 3.30 cd	71.7 $\pm$ 2.58b	75.4 $\pm$ 1.16ab	86.6 $\pm$ 5.20a	58.3 $\pm$ 2.04c
Ile	27.6 $\pm$ 0.12 cd	22.8 $\pm$ 2.06 d	33.9 $\pm$ 2.21c	51.6 $\pm$ 3.04a	43.7 $\pm$ 4.11b	18.5 $\pm$ 0.97 d
Leu	50.1 $\pm$ 2.07b	63.2 $\pm$ 1.99a	53.8 $\pm$ 3.75b	66.8 $\pm$ 2.00a	55.3 $\pm$ 2.98b	26.9 $\pm$ 0.01c
Phe	36.5 $\pm$ 3.95c	26.1 $\pm$ 4.00 d	52.8 $\pm$ 0.91b	67.0 $\pm$ 1.57a	23.7 $\pm$ 3.97 d	15.8 $\pm$ 0.01e
ΣUTAA	816.6 $\pm$ 4.09a	829.3 $\pm$ 2.01a	786.5 $\pm$ 3.01b	831.0 $\pm$ 1.25a	748.7 $\pm$ 3.08c	455.3 $\pm$ 2.15 d
ΣBCAAs	103.4 $\pm$ 1.05c	133.4 $\pm$ 2.81bc	134.8 $\pm$ 1.32bc	190.0 $\pm$ 1.18a	147.6 $\pm$ 0.97b	71.9 $\pm$ 0.54 d
ΣFAA	1688.7 $\pm$ 9.07 d	1949.1 $\pm$ 7.12c	2111.6 $\pm$ 3.99b	2335.1 $\pm$ 5.80a	2035.1 $\pm$ 9.67bc	1402.4 $\pm$ 5.96 d

Means followed by different letters in a row are significantly different ($P\leq 0.05$) ($n=3$). *FAA* free amino acids, *UTAA* Unami-Taste amino acids, *BCAAs* Branched-Chain Amino Acids

higher than 85 $^{\circ}\text{C}$. It can be concluded that the highest content of BCAA was obtained in the 85 $^{\circ}\text{C}$ soup.

Although the effect of heating temperature on the contents of each type of the amino acids was quite different, the total FAA concentration had a clear correlation with cooking temperatures, the concentration of total FAA increased continuously until the temperature reached 85 $^{\circ}\text{C}$, thereafter, the concentration decreased. There were two possible ways to constitute source of FAA in the soup, one was the original FAA flew into soup from the fish flesh and the other the degradation of proteins and peptides due to the aminopeptidase (Buscailhon et al. 1994). Therefore, the concentration of total FAA was determined by the ratio of formation and decomposition of FAA in the food. Previous researchers observed that some amino acids involved in the Maillard reaction with reducing sugars to produce hexaldehydes, heptaldehydes and so on (Chawla and Sahu 2007). Also, a small part of degradation of amino acids generated esters, ketones via through the Strecker degradation pathway. These reactions provided flavour compounds to food through the pathway of interaction of amino acids with compounds from lipid oxidation (Ardö, 2006).

Conclusion

The data obtained suggested that the best quality soup (in color, aroma, flavour and soup pattern) was obtained by heating crucian carp in a 0.5% NaCl solution (1:6, w/v) at 85 $^{\circ}\text{C}$ for 2 h. From the analysis of variance of the sensory data, temperature plays a more important role than cooking time in the generation of the sensory properties in crucian carp soup. The heating temperature played a crucial role on the proteolysis and solubility of protein in the fish soup, the highest contents of total peptides, total FAA, tasty amino acids and BCAA in crucian carp soup were obtained at the cooking temperature of 85 $^{\circ}\text{C}$. In conclusion, a cooking temperature of 85 $^{\circ}\text{C}$ was preferred for more excellent flavor and higher nutritional value of crucian carp soup.

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