Video Article

Investigating the Neural Mechanisms of Aware and Unaware Fear Memory with fMRI

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Abstract

Pavlovian fear conditioning is often used in combination with functional magnetic resonance imaging (fMRI) in humans to investigate the neural substrates of associative learning¹⁻⁵. In these studies, it is important to provide behavioral evidence of conditioning to verify that differences in brain activity are learning-related and correlated with human behavior.

Fear conditioning studies often monitor autonomic responses (e.g. skin conductance response; SCR) as an index of learning and memory⁶⁻⁸. In addition, other behavioral measures can provide valuable information about the learning process and/or other cognitive functions that influence conditioning. For example, the impact unconditioned stimulus (UCS) expectancies have on the expression of the conditioned response (CR) and unconditioned response (UCR) has been a topic of interest in several recent studies⁹⁻¹⁴. SCR and UCS expectancy measures have recently been used in conjunction with fMRI to investigate the neural substrates of aware and unaware fear learning and memory processes¹⁵. Although these cognitive processes can be evaluated to some degree following the conditioning session, post-conditioning assessments cannot measure expectations on a trial-to-trial basis and are susceptible to interference and forgetting, as well as other factors that may distort results^{16,17}.

Monitoring autonomic and behavioral responses simultaneously with fMRI provides a mechanism by which the neural substrates that mediate complex relationships between cognitive processes and behavioral/autonomic responses can be assessed. However, monitoring autonomic and behavioral responses in the MRI environment poses a number of practical problems. Specifically, 1) standard behavioral and physiological monitoring equipment is constructed of ferrous material that cannot be safely used near the MRI scanner, 2) when this equipment is placed outside of the MRI scanning chamber, the cables projecting to the subject can carry RF noise that produces artifacts in brain images, 3) artifacts can be produced within the skin conductance signal by switching gradients during scanning, 4) the fMRI signal produced by the motor demands of behavioral responses may need to be distinguished from activity related to the cognitive processes of interest. Each of these issues can be resolved with modifications to the setup of physiological monitoring equipment and additional data analysis procedures. Here we present a methodology to simultaneously monitor autonomic and behavioral responses during fMRI, and demonstrate the use of these methods to investigate aware and unaware memory processes during fear conditioning.

Video Link

The video component of this article can be found at <http://www.jove.com/details.php?id=3083>

Protocol

1. Psychophysiology

The Biopac Systems, Inc. physiological monitoring system (see Table of specific equipment) is non-standard equipment in most imaging facilities. Schedule 15-30 minutes prior to participant arrival to set up physiological monitoring and other equipment described in this protocol (Figure 1).

1. Connect a control room computer operating AcqKnowledge (Biopac Systems, Inc.) physiological monitoring software to the Biopac MP150 (MP150WSW) using a standard Ethernet crossover cable (CBLETH2).
2. Connect the Biopac Isolated Digital Interface (STP100C) to a control room computer operating Presentation (Neurobehavioral Systems, Inc; Albany, CA) software using a DB25 M/F ribbon cable.
3. Connect the Biopac GSR amplifier (EDA-100C-MRI) to the RF interference filter (MRIRFIF) within the control room using a shielded extension cable (MECMRI-3).
4. Connect the RF interference filter (MRIRFIF) to a shielded extension cable (MECMRI-1) within the MRI scanning chamber.
5. Connect the shielded extension cable to carbon fiber lead wires (LEAD 108) that attach to radio translucent electrodes (EL508). Note: Twisting the leads in a tight spiral reduces artifacts in the skin conductance data that can be created during scanning.
6. Attach radio translucent electrodes (EL508) to the distal phalanx of the middle and ring fingers of the participant's left hand.
7. Due to the nature of scanning equipment, MRI chamber room temperatures are often set below 21°C. Cover the participant with a blanket to maintain hand temperature.

2. Behavioral Responses (Joystick)

1. Connect a control room computer operating Presentation software (Neurobehavioral Systems, Inc; Albany, CA) to the joystick's FORP Interface Unit (Current Designs, Inc; Philadelphia, PA) using a USB-mini cable.
2. Connect a fiber optic cable to the FORP Interface Unit within the control room, then pass the cable through a wave guide into the MRI chamber.
3. Connect the fiber optic cable to the MR-compatible joystick.
4. Direct participants to place the joystick in a comfortable and easy to reach position.

3. Stimulus Presentation

1. Connect a control room computer operating Presentation software to the external VGA and audio ports of the IFIS-SA (Invivo Corp., Orlando, FL) control room console (Figure 1).
2. Check the fiber optic cable connections between the IFIS control room console and the IFIS Peripheral Interface Unit within the MRI chamber, as well as the connections between the Peripheral Interface Unit and the Audio/Visual Display Unit.
3. Place the Audio/Visual Display Unit behind the head-coil such that the participant can view the monitor through a mirror attached to the head-coil.
4. Connect the Audio/Visual Display Unit's acoustic interface box to the IFIS system's MR-compatible stereo headphones using vinyl tubing.
5. Calibrate the volume of auditory stimuli using a sound pressure level meter.

4. Experimental Procedure

1. Inform participants that 2 tones will be presented several times during the study, and that the volume of the tones will vary above and below their perceptual threshold (Figure 2).
2. Direct participants to push a button on the joystick box immediately upon hearing either tone, then to update their expectation of receiving the UCS by moving the joystick to control the position of a rating bar on a 0 to 100 scale (Figure 3).
3. Instruct participants to rate their UCS expectancy on a continuous scale from 0 to 100. Inform them that ratings of 0 indicate they are certain the UCS will not be presented, ratings of 50 indicate they are uncertain whether the UCS will be presented, and ratings of 100 indicate they are certain the UCS will be presented. Direct participants to use other values on the scale to indicate intermediate expectations. Then, allow participants to practice using the joystick to make ratings.
4. Expose participants to a differential fear conditioning procedure using 2 tones (700 & 1300 Hz; 10s duration; 20s ITI) as the conditioned stimuli (CS) and a loud white-noise (100dB, 500ms) as the UCS.
5. Present 60 trials of the CS+ (coterminating with the UCS) and 60 trials of the CS- (presented without the UCS) in a pseudorandom order such that no more than 2 trials of the same CS are presented consecutively.
6. Counterbalance the tones that serve as the CS+ and CS- across participants.
7. Modulate the volume of the CS+ and CS- independently. Adjust CS volume on the subsequent trial with the same CS. Decrease CS volume 5dB if a button press is made (i.e. following a perceived trial). Increase the volume 5dB if a button press is not made (i.e. following an unperceived trial).

5. Scanning Procedure

1. Collect standard high resolution T1-weighted structural images (e.g. MPRAGE) to serve as an anatomical reference for functional data.
2. Collect BOLD fMRI of the whole brain during the conditioning procedure. Thirty six, 4mm thick slices should be sufficient to cover the brain with relatively standard imaging parameters (e.g. TR = 2000ms, TE = 30ms, FOV = 24cm, 64x64 matrix). Synchronize the fMRI acquisition with stimulus presentation using an fMRI trigger box.

6. SCR Data Acquisition & Analysis

1. Sample skin conductance at 2,000 Hz using AcqKnowledge software and the MR-compatible Biopac physiological monitoring system described in section 1.
2. Apply a 1 Hz Infinite Impulse Response (IIR) low pass digital filter to the skin conductance data to reduce artifacts produced during imaging (see Figure 4).
3. Resample the skin conductance data at 250 Hz.
4. Calculate SCR as the difference in skin conductance level from response onset to response peak.
5. SCR data can be square root transformed to normalize the distribution of response amplitudes prior to statistical analysis.

7. UCS Expectancy Data Acquisition & Analysis

1. Sample (40 Hz) and record UCS expectancy data using Presentation software.
2. Calculate UCS expectancy as the average (1s sample) response during the last second of CS presentation.

8. Functional MRI Data Acquisition & Analysis

1. Complete standard preprocessing of brain imaging data (e.g. slice timing correction, image registration, spatial smoothing) using a functional imaging analysis software package (e.g. AFNI¹⁸).
2. Create standard nuisance (e.g. motion) and stimulus-based regressors for perceived and unperceived trials of the CS+ and CS-, as well as the UCS.
3. Create a motor response-based reference waveform to serve as a nuisance regressor to account for motor activity related to button press responses.
 - a. Create a stick function that codes for the timing of button press responses.
 - b. Convolve the button press stick function with the canonical hemodynamic response function (HRF).
4. Create motor response-based reference waveform to serve as a nuisance regressor to account for motor activity related to joystick responses.

- a. Create a stick function that codes for the timing of changes in the slope (e.g. slope absolute value > 10) of UCS expectancy ratings.
 - b. Convolve the joystick slope stick function with the canonical HRF.
5. Perform first level analyses using all stimulus-based and nuisance regressors.
 6. Perform a second level repeated measures ANOVA to identify regions in which activation shows a main effect of CS type, a main effect of perception, or a CS type X perception interaction.

9. Representative Results:

The methodology presented here typically results in relatively high UCS expectancy ratings during perceived CS+ trials and low ratings during perceived CS- trials (Figure 5)^{10,15,19}. Such results indicate participants are aware of CS-UCS contingencies. On unperceived trials, UCS expectancy ratings typically remain unchanged from pre-CS ratings. UCS expectancies on these unperceived CS+ and CS- trials typically fall near 50 indicating participants are unsure of whether the UCS will be presented^{10,15,19} (Figure 5). This inability to produce differential UCS expectancy ratings to the unperceived CS+ and CS- indicates that participants are unable to express their contingency awareness on unperceived conditioning trials (Figure 6). In contrast, learning-related changes in SCR have been observed during both perceived and unperceived conditioning trials^{10,15,19}. Specifically, SCRs were larger to the perceived CS+ than to the perceived CS-. Similarly, larger SCRs have been demonstrated during unperceived CS+ than unperceived CS- trials^{10,15,19} (Figure 6). Taken together, these behavioral and autonomic data demonstrate fear conditioning with contingency awareness on perceived trials, and fear conditioning without contingency awareness on unperceived trials. The functional imaging research using this methodology has demonstrated learning-related hippocampal activation on perceived, but not unperceived conditioning trials¹⁵ (Figure 7). In contrast, differential amygdala activity was observed on both perceived and unperceived conditioning trials¹⁵. These findings are consistent with the view that the hippocampus supports processes related to contingency awareness, while the amygdala supports CR expression with and without awareness.

Figure 1. Diagram of basic equipment for stimulus presentation and behavioral/psychophysiological response monitoring. Presentation software is used to present audio-visual stimuli and monitor UCS expectancy ratings made by moving a joystick with the right hand. AcqKnowledge software and Biopac equipment are used to monitor skin conductance from the left hand. Solid (Biopac), single dashed (IFIS Audio-Visual), and double dashed lines (Fiber optic joystick) depict cables for distinct stimulus presentation and response monitoring systems. Black arrows indicate direction of information flow.

Figure 2. Conditioned Stimuli. Present the CS+ and CS- in a pseudorandom order such that no more than 2 trials of the same CS are presented consecutively. Vary the volume of the CS+ and CS- independently. If a CS is perceived (indicated by a button press), decrease CS volume 5dB on the subsequent trial of the same CS. If a CS is unperceived (indicated by no button press), raise CS volume 5db on the subsequent trial with the same CS.

Figure 3. UCS expectancy rating scale. Instruct participants to rate their expectation of UCS presentation on a 0 to 100 scale. Ratings of 0 indicate certainty the UCS will not be presented, ratings of 100 indicate certainty the UCS will be presented, and ratings of 50 reflect uncertainty as to whether the UCS will be presented. Intermediate ratings should be used to indicate gradations in UCS expectancy.

Figure 4. Comparison of raw and filtered skin conductance data. a) Raw skin conductance data collected during fMRI. b) Skin conductance data after application of a 1Hz IIR low pass filter.

Figure 5. UCS expectancy ratings. -Participants typically report high UCS expectancies on perceived CS+ trials and low expectancies on perceived CS- trials. UCS expectancies on unperceived CS+ and CS- trials do not differ.

Figure 6. UCS expectancy and SCR. Differences in UCS expectancy are typically observed on perceived CS+ and CS- trials indicating participants are aware of the stimulus contingencies. On unperceived trials, UCS expectancy ratings typically do not differ indicating participants are unable to express their contingency awareness. In contrast, differences in conditioned SCRs are usually observed on both perceived and unperceived conditioning trials. Such findings reflect learned fear expression with (i.e. on perceived trials) and without (i.e. on unperceived trials) contingency awareness.

Figure 7. Functional MRI of the hippocampus and amygdala. Hippocampal responses are typically larger to the CS+ than CS- on perceived, but not unperceived conditioning trials. Differential amygdala responses are typically observed on both perceived and unperceived conditioning trials. These findings are consistent with the view that the hippocampus supports processes related to contingency awareness, while the amygdala supports fear expression with and without awareness.

Discussion

The fear conditioning methodology described here provides a means to investigate the neural mechanisms of aware and unaware fear memory processes. This method takes advantage of the simultaneous monitoring of behavioral, autonomic, and fMRI data. Monitoring behavioral (i.e. UCS expectancy) and autonomic responses (i.e. SCR) is a critical component of this method. UCS expectancy provides a means to assess contingency awareness, while SCR provides an index of CR expression. Together, these behavioral and autonomic responses can be used during the presentation of supra and subthreshold CS+ and CS- trials to investigate fear conditioning with and without contingency awareness. Functional MRI data can then be used to investigate the neural correlates of aware and unaware fear memory processes. A particular strength of this methodology is that it exposes participants to each type of conditioning trial (i.e. perceived CS+ & CS-, unperceived CS+ & CS-). Within-subject designs like the one described here are more powerful than between subject designs because of the relatively large inter-subject variability observed in both SCR and fMRI signal responses. Another strong point of this method is that the volume of CS presentation is tailored to each participant's perceptual threshold. Further, the perceptual threshold is allowed to vary over the course of the conditioning session. Prior work has typically presented stimuli at a set level below threshold^{7,20,21}. However, perceptual thresholds can vary over time reducing the ability to detect subthreshold effects²². An additional strength of this methodology is that UCS expectancy is assessed on a trial-by-trial basis during the conditioning session. Other fMRI research has assessed awareness of CS-UCS contingencies during post-conditioning evaluations²³. However, post-conditioning assessments 1) cannot assess variations in expectancy from trial-to-trial, 2) may be insensitive to subtle evidence of contingency awareness, and 3) are susceptible to issues that distort results such as forgetting and interference. Although there are a number of strengths to our methodology, monitoring UCS expectancy as described may engage attentional processes in a manner that differs from studies that do not use online expectancy measures. This is an issue that investigators should consider along with the advantages of this methodology when designing their projects.

Disclosures

No conflicts of interest declared.

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References

1. LaBar, K.S., Gatenby, J.C., Gore, J.C., LeDoux, J.E., & Phelps, E.A. Human amygdala activation during conditioned fear acquisition and extinction: a mixed-trial fMRI study. *Neuron*. **20**, 937-945, doi: S0896-6273(00)80475-4 [pii] (1998).
2. Buchel, C., Morris, J., Dolan, R.J. & Friston, K.J. Brain systems mediating aversive conditioning: an event-related fMRI study. *Neuron*. **20**, 947-957, doi:S0896-6273(00)80476-6 [pii] (1998).
3. Cheng, D.T., Knight, D.C., Smith, C.N., Stein, E.A. & Helmstetter, F.J. Functional MRI of human amygdala activity during Pavlovian fear conditioning: stimulus processing versus response expression. *Behav. Neurosci.* **117**, 3-10 (2003).
4. Knight, D.C., Smith, C.N., Stein, E.A. & Helmstetter, F.J. Functional MRI of human Pavlovian fear conditioning: patterns of activation as a function of learning. *Neuroreport*. **10**, 3665-3670 (1999).
5. Cheng, D.T., Knight, D.C., Smith, C.N. & Helmstetter, F.J. Human amygdala activity during the expression of fear responses. *Behav. Neurosci.* **120**, 1187-1195, doi:2006-22387-001 [pii] 10.1037/0735-7044.120.5.1187 (2006).
6. Balderston, N.L. & Helmstetter, F.J. Conditioning with masked stimuli affects the timecourse of skin conductance responses. *Behav. Neurosci.* **124**, 478-489, doi:2010-16138-005 [pii] 10.1037/a0019927 (2010).
7. Esteves, F., Parra, C., Dimberg, U. & Ohman, A. Nonconscious associative learning: Pavlovian conditioning of skin conductance responses to masked fear-relevant facial stimuli. *Psychophysiology*. **31**, 375-385 (1994).
8. Cheng, D.T., Richards, J. & Helmstetter, F.J. Activity in the human amygdala corresponds to early, rather than late period autonomic responses to a signal for shock. *Learn. Mem.* **14**, 485-490, doi:14/7/485 [pii] 10.1101/lm.632007 (2007).
9. Knight, D.C., Nguyen, H.T. & Bandettini, P.A. The role of the human amygdala in the production of conditioned fear responses. *Neuroimage*. **26**, 1193-1200, doi:S1053-8119(05)00184-9 [pii] 10.1016/j.neuroimage.2005.03.020 (2005).
10. Knight, D.C., Nguyen, H.T. & Bandettini, P.A. The role of awareness in delay and trace fear conditioning in humans. *Cogn. Affect. Behav. Neurosci.* **6**, 157-162 (2006).
11. Schultz, D.H. & Helmstetter, F.J. Classical conditioning of autonomic fear responses is independent of contingency awareness. *J. Exp. Psychol. Anim. Behav. Process.* **36**, 495-500, doi:2010-21866-002 [pii] 10.1037/a0020263 (2010).
12. Dunsmoor, J.E., Bandettini, P.A. & Knight, D.C. Neural correlates of unconditioned response diminution during Pavlovian conditioning. *Neuroimage*. **40**, 811-817, doi:S1053-8119(07)01090-7 [pii] 10.1016/j.neuroimage.2007.11.042 (2008).
13. Katkin, E.S., Wiens, S. & Ohman, A. Nonconscious fear conditioning, visceral perception, and the development of gut feelings. *Psychol. Sci.* **12**, 366-370 (2001).
14. Knight, D.C., Waters, N.S., King, M.K. & Bandettini, P.A. Learning-related diminution of unconditioned SCR and fMRI signal responses. *Neuroimage*. **49**, 843-848, doi:S1053-8119(09)00777-0 [pii] 10.1016/j.neuroimage.2009.07.012 (2010).
15. Knight, D.C., Waters, N.S. & Bandettini, P.A. Neural substrates of explicit and implicit fear memory. *Neuroimage*. **45**, 208-214, doi:S1053-8119(08)01217-2 [pii] 10.1016/j.neuroimage.2008.11.015 (2009).
16. Lovibond, P.F. & Shanks, D.R. The role of awareness in Pavlovian conditioning: empirical evidence and theoretical implications. *J. Exp. Psychol. Anim. Behav. Process.* **28**, 3-26 (2002).
17. LaBar, K.S. & Disterhoft, J.F. Conditioning, awareness, and the hippocampus. *Hippocampus*. **8**, 620-626, doi:10.1002/(SICI)1098-1063(1998)8:6<620::AID-HIPO4>3.0.CO;2-6 (1998).
18. Cox, R.W. AFNI: software for analysis and visualization of functional magnetic resonance neuroimages. *Comput. Biomed. Res.* **29**, 162-173, doi:S0010480996900142 [pii] (1996).
19. Knight, D.C., Nguyen, H.T. & Bandettini, P.A. Expression of conditional fear with and without awareness. *Proc. Natl. Acad. Sci. U. S. A.* **100**, 15280-15283, doi:10.1073/pnas.2535780100 2535780100 [pii] (2003).
20. Bunce, S.C., Bernat, E., Wong, P.S. & Shevrin, H. Further evidence for unconscious learning: preliminary support for the conditioning of facial EMG to subliminal stimuli. *J. Psychiatr. Res.* **33**, 341-347, doi:S0022-3956(99)00003-5 [pii] (1999).
21. Kotze, H.F. & Moller, A.T. Effect of auditory subliminal stimulation on GSR. *Psychol. Rep.* **67**, 931-934 (1990).
22. Miller, J. Threshold variability in subliminal perception experiments: fixed threshold estimates reduce power to detect subliminal effects. *J. Exp. Psychol. Hum. Percept. Perform.* **17**, 841-851 (1991).
23. Tabbert, K., Stark, R., Kirsch, P. & Vaitl, D. Dissociation of neural responses and skin conductance reactions during fear conditioning with and without awareness of stimulus contingencies. *Neuroimage*. **32**, 761-770, doi:S1053-8119(06)00221-7 [pii] 10.1016/j.neuroimage.2006.03.038 (2006).