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Extensive Aspartoacylase Expression in the Rat Central Nervous System

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Abstract

Aspartoacylase (ASP) catalyzes deacetylation of *N*-acetylaspartate (NAA) to generate acetate and aspartate. Mutations in the gene for ASP lead to reduced acetate availability in the CNS during development resulting in the fatal leukodystrophy Canavan disease. Highly specific polyclonal antibodies to ASP were used to examine CNS expression in adult rats. In white matter ASP expression was associated with oligodendrocyte cell bodies, nuclei and some processes, but showed a dissimilar distribution pattern to myelin basic protein and oligodendrocyte specific protein. Microglia expressed ASP in all CNS regions examined, as did ependymal cells of the choroid plexus. Pial and ependymal cells and some endothelial cells were ASP positive, as were unidentified cellular nuclei throughout the CNS. Astrocytes did not express ASP in their cytoplasm. In some fiber pathways and nerves, particularly in the brainstem and spinal cord, the axoplasm of many neuronal fibers expressed ASP, as did some neurons. Acetyl coenzyme A synthase immunoreactivity was also observed in the axoplasm of many of the same fiber pathways and nerves. All ASP-immunoreactive elements were unstained in brain sections from tremor rats, an ASP-null mutant. The strong expression of ASP in oligodendrocyte cell bodies is consistent with a lipogenic role in myelination. Strong ASP expression in cell nuclei is consistent with a role for NAA-derived acetate in nuclear acetylation reactions, including histone acetylation. Expression of ASP in microglia may indicate a role in lipid synthesis in these cells, whereas expression in axons suggests that some neurons can both synthesize and catabolize NAA.

Keywords

ASP; *N*-acetylaspartate; NAA; Canavan disease; oligodendrocytes; microglia; leptomeninges; myelin; acetyl coenzyme A; acetyl coenzyme A synthase; protein acetylation; histone acetylation

Introduction

Aspartoacylase (ASP, EC 3.5.1.15) is one of three aminoacylase enzymes that are responsible for the deacetylation of *N*-acetylated amino acids. In tissues such as kidney, these acetylated amino acids are derived from the catabolism of *N*-terminal acetylated proteins (reviewed in Perrier et al., 2005), and aminoacylase enzymes act to salvage the deacetylated amino acids for reuse in protein synthesis (Lindner et al., 2008).

Aminoacylases 1 and 3 have relatively broad substrate specificity and act on a number of N-acetylated amino acids. ASPA, also known as aminoacylase 2, acts to deacetylate only one acetylated amino acid, namely N-acetylaspertate (NAA) (D'Adamo, Jr. et al., 1977; Madhavarao et al., 2003). ASPA hydrolyzes NAA into free acetate and aspartate (Kaul et al., 1993; Zeng et al., 2002) and is expressed strongly in a number of tissues including the brain and kidney (Birnbaum, 1955; Madhavarao et al., 2004; Hershfield et al., 2006). In peripheral tissues ASPA may function like other aminoacylases to recycle NAA derived from the breakdown of proteins such as actin, which is acetylated at its N-terminal aspartate (Gaetjens et al., 1966; Alving et al., 1966). However, in light of the exceptionally high concentration of NAA in the brain (10 mM or higher; Tallan, 1957; Inglese et al., 2008; Miyake et al., 1981), ASPA has apparently adopted additional specialized roles in CNS metabolism beyond simple amino acid salvage associated with the turnover of N-acetylated proteins.

Mutations in genes encoding for aminoacylase enzymes lead to reduced or absent capacity for the catabolism of N-acetylated amino acids, and increased excretion of the corresponding acetylated amino acids in urine. Mutations in aminoacylase 1 lead to an inborn error of metabolism and result in macrocephaly and neurological symptoms (Sass et al., 2006; Sass et al., 2007). The most severe aminoacylase deficiency results from mutations in the gene that encodes for ASPA, leading to the development of Canavan disease during infancy (Matalon et al., 1988). Canavan disease is a progressive and fatal hereditary neurodegenerative disease characterized by macrocephaly, reduced myelination and severe vacuolation in thalamus, midbrain, brainstem and spinal cord (Adachi et al., 1973; Matalon et al., 1995; Surendran et al., 2005). Catabolism of NAA is absent in Canavan disease patients, NAA levels are increased in the brain, and affected neonates excrete excess NAA in their urine (Jakobs et al., 1991; Kelley et al., 1992).

The primary functional significance of ASPA-mediated deacetylation of NAA in the CNS is still under debate after decades of research, and there is additional uncertainty concerning the metabolic fate of the acetate derived from NAA catabolism (reviewed in Moffett et al., 2007). Acetyl coenzyme A (acetyl CoA) is a primary building block for lipids, and ASPA-mediated deacetylation of NAA has been proposed to provide some of the acetate necessary for the synthesis of acetyl CoA, and in turn myelin lipids (D'Adamo, Jr. et al., 1966; D'Adamo et al., 1968; Burri et al., 1991; Mehta et al., 1995; Namboodiri et al., 2006). Therefore, Canavan disease has been hypothesized to result from a deficiency in NAA-derived acetate, which leads to decreased myelin synthesis during early postnatal CNS development (Hagenfeldt et al., 1987; Mehta et al., 1995; Madhavarao et al., 2005; Wang et al., 2009). Other metabolic deficits that may result from ASPA deficiency include virtually all cellular functions linked to acetyl CoA supply in the CNS, particularly in oligodendrocytes. These could include protein acetylation reactions such as nuclear histone acetylation, endoplasmic reticulum protein acetylation and cytoskeletal protein acetylation (Ariyannur et al., 2010b). ASPA is localized in both the cytosolic and nuclear compartments of oligodendrocytes and is transported between the two compartments (Hershfield et al., 2006). Additionally, one of the enzymes responsible for converting free acetate derived from NAA into acetyl coenzyme A, known as acetyl coenzyme A synthase-1 (AceCS1), is also localized in the nuclei of oligodendrocytes (Ariyannur et al., 2010a). The localization pattern for these two enzymes is consistent with a role for NAA-derived acetate in nuclear histone acetylation reactions in oligodendrocytes. Histone acetylation is associated with epigenetic gene regulation, and therefore disrupted histone acetylation in Canavan disease may be a major etiological factor in the progression of the disease.

The polyclonal antibody used in the current study, which was directed against an ASPA peptide sequence, provided an expanded and more detailed expression pattern for ASPA

than reported in previous studies. Most importantly, in the current study the identity of the target epitope was confirmed to correspond exclusively to ASPA protein by demonstrating that no staining was present in brain tissue sections from a mutant rat strain, the tremor rat, in which the ASPA gene sequence is absent (Kitada et al., 2000). Here we present a detailed analysis of ASPA protein expression throughout the rat brain.

Materials and Methods

ASPA antibodies

Specific rabbit polyclonal antibodies were produced against a 19 amino acid conserved murine and human ASPA peptide sequence (residues 83-101: SEDLPYEVRRRAQEINHLFG). This antibody has been reported previously (Hershfield et al., 2006). During the course of working with the polyclonal anti-ASPA sera it was found that diffuse background staining occurred in wild type and tremor rat tissue sections. In order to remove non-specific antibodies, polyclonal antisera were diluted 1:1,000 in PBS containing 2% normal goat serum (NGS) and pre-incubated for 24 hr or longer with an excess of formalin-fixed liver and brain tissue slices from tremor rats. This process was repeated until no background immunoreactivity was observed in tremor rat brain slices.

Peroxidase immunohistochemistry

Six adult female and three adult male Sprague-Dawley rats (Charles River Laboratories, Raleigh NC, 200-250 grams) and 3 tremor rats (Wistar strain, 70 days old, bred in our facility) were anesthetized with pentobarbital (50 mg/kg) and perfused transcardially with 400 ml of 4% freshly depolymerized paraformaldehyde. Brains were removed and postfixed for several hours in paraformaldehyde before being passed through a series of 10%, 20% and 30% sucrose solutions. Brains were grossly cut in 4mm thick coronal blocks using a Jacobowitz rat brain matrix slicer (Jacobowitz, 1974) and the blocks were mounted on aluminum chucks in OCT embedding compound and frozen rapidly with compressed difluoroethane gas. Coronal sections were cut at a thickness of 20 μ m at -18°C and were collected in 10 mM phosphate buffered saline (PBS).

Avidin-biotin complex/peroxidase immunohistochemistry was done as previously described (Madhavarao et al., 2004) with some modifications. Sections were processed free-floating. Endogenous peroxidase activity was blocked by washing freshly cut sections in PBS and incubating them in a 50:50 mixture of methanol and water containing 1% H_2O_2 for 30 min. Sections were blocked against non-specific antibody binding by incubation for 20 minutes or longer in PBS containing 2% NGS and 0.1 % sodium azide.

Brain sections were incubated for 16 to 24 hr with the tremor tissue-adsorbed polyclonal ASPA antibodies at a dilution of 1:25,000 to 1:30,000. Sections were maintained in a humid chamber at room temperature with constant rotary agitation, and were washed thoroughly after incubation to remove sodium azide. Bound antibodies were visualized by the avidin-biotin complex method with horseradish peroxidase as the enzyme marker (Vectastain Elite; Vector Labs, Burlingame, CA). The sections were incubated with the biotinylated secondary antibody and avidin-peroxidase complex solutions for 90 minutes each. The secondary antibody was diluted in PBS plus 2% NGS and the ABC reagent in PBS plus 0.5% BSA, both without azide. After final washing, the sections were developed with a Ni and Co enhanced diaminobenzidine chromogen (Pierce Chemical Co, Rockford, IL). AceCS1 immunohistochemistry was performed as previously described (Ariannur et al., 2010a).

Comparative studies with antibodies to myelin basic protein (MBP) and oligodendrocyte specific protein (OSP) were done by a modified method in order to improve antibody penetration into dense myelin. Antigen retrieval was performed on free floating tissue

sections using EDTA (5 mM HEPES buffer with 1 mM EDTA and 0.05% Triton X-100 at pH 8 and 80°C for 10 min.). Polyclonal antibodies (Abcam, Cambridge, MA; MBP = #AB 2404; OSP = #AB 53041) were diluted 1:500, and 1:4,000 respectively, and purified ASPA antibodies were diluted 1:30,000. Primary antibodies were applied to tissue sections for 42 to 48 hr at room temperature in a humid chamber with constant rotary agitation. Incubation times with the biotinylated secondary antibody and avidin-peroxidase complex were extended to 2 hr each.

Stained sections were transferred to deionized water containing 0.01% BSA, and were manually mounted on treated slides (Superfrost plus, Fischer Scientific) with a fine tipped brush. Slides were dried at 45°C for 10 to 15 minutes, dehydrated in an ethanol series, cleared in xylene and sealed with cover glasses using xylene based adhesive. Images were acquired on a DIC-equipped Olympus BX51 microscope with an Olympus DP71 digital camera, and were adjusted to normalize brightness and contrast with PC based imaging software (Adobe Systems, San Jose CA). Additional software was used to correct for background illumination levels, and in some cases to produce images with extended depth of field (Media Cybernetics, Silver Spring MD).

Protein extract preparation for Western blotting

Two adult Sprague-Dawley wild type rats and two tremor rats were anesthetized, brains were removed and frozen on dry ice, and stored at -80°C until use. All subsequent steps were performed on ice. Tissues were minced and homogenates were prepared by 8-10 strokes with a Potter-Elvehjem homogenizer in isolation medium (0.25 M sucrose, 25 mM KCl, 5 mM MgCl₂, 10mM Tris-HCl pH 7.4) with protease inhibitor cocktail (15 µL/mL) and phosphatase inhibitors (Sigma-Aldrich). Following filtration, solutions were centrifuged (800g, 10 min 4°C), the supernatants were adjusted to homogenization buffer (0.5 mM DTT, 50 mM Tris-HCl, pH 8.0, 50 mM NaCl, 0.05% IGEPAL CA-630, 8% glycerol) and were centrifuged again (16,000 × g, 20 min, 4°C). The resulting supernatants were used for Western blotting. Protein concentrations were determined by Bio-Rad's DC protein assay (Hercules, CA).

SDS-PAGE and Western blot

Western blotting was done as previously described (Ariyannur et al., 2010a). Samples were diluted in loading buffer with 10mM DTT, and heated (90° C, 5 min) prior to loading onto 10% pre-cast Tris-glycine gels (Invitrogen, Carlsbad, CA). Brain proteins were applied at 60 µg per lane. Proteins were transferred to Immobilon-P PVDF membranes, which were blocked with 0.05% Tween-20 and 5% NGS. Blots were incubated overnight with anti-ASPA antibodies (see figure legend for antibody concentrations), were washed in PBS, and then incubated for 1 hr with horseradish peroxidase-conjugated secondary antibodies (1:2,500). Membranes were washed with PBS and developed with Super Fast diaminobenzidine substrate (Sigma/Aldrich).

Results

ASPA antibody specificity

In the current study we have examined the cellular localization of ASPA protein in the rat CNS using an adsorption purified polyclonal antibody. The crude polyclonal antibody was specific for ASPA, and displayed a major protein band with an apparent molecular weight of approximately 37 kDa in Western blots of the cytosolic fraction from rat brain homogenates (Fig. 1 insert). Two relatively minor immunoreactive bands were observed at other molecular weights in Western blots of protein extracts from wild type and tremor rat brains with unpurified antisera. After adsorption of the crude ASPA antisera with brain and liver

tissue from tremor rats, the non-specific immunoreactive bands were absent in Western blots, and only the 37 kDa band remained with brain extracts from wild type rats.

Using the crude antisera, we observed non-specific background staining in brain sections from wild type and tremor rats at the dilutions used for immunohistochemistry (1:25,000 to 1:30,000). The background staining was especially strong in the striatum (Figs. 1, A and B), and it was determined that this background staining could be removed by pre-incubating partially diluted antisera with formalin-fixed brain and liver tissue slices from tremor rats. Pre-incubation of the polyclonal antibodies with a large excess of tissue sections from this ASPA null animal removed the non-specific antibodies from the polyclonal mixture (Figs. 1 C, D). Using this method we were able to produce highly specific polyclonal antibody preparations with virtually no background staining, providing high-contrast images with exceptional cellular detail. This adsorption purification method also prevented the typical loss of antibody titer associated with affinity purification. Further, all immunoreactivity with the pre-adsorbed anti-ASPA antibodies was absent in tremor rat brain sections (Fig. 1D), indicating that all immunoreactivity in wild type rat brain sections was due to ASPA protein expression.

ASPA immunoreactivity overview

In general, ASPA immunoreactivity in the brain was most intense in fiber tracts, thalamus and brainstem as previously reported (Klugmann et al., 2003; Madhavarao et al., 2004). Cortical areas including isocortex, hippocampus, piriform cortex, entorhinal cortex and cerebellar cortex had generally lower expression levels for ASPA than white matter. Fiber tract ASPA expression was predominated by oligodendrocytes, whereas expression in grey matter occurred in oligodendrocytes, microglia and unidentified cellular nuclei. In addition to oligodendrocytes and microglia, pial cells were among the most strongly immunoreactive cells for ASPA throughout the brain. We also found that ASPA expression was stronger in fiber pathways than could be accounted for by oligodendrocytes and their processes. In forebrain, fiber pathways such as the corpus callosum, fornix, optic chiasm and tracts, internal capsule and medial forebrain bundle all showed light to moderate ASPA immunoreactivity in apparent axons, but this was difficult to image at the light microscopic level due to the high level of expression in oligodendrocytes and some of their processes. Axoplasmic ASPA expression was more obvious in large diameter fibers of the brainstem. When caudal fiber pathways composed of large axons were cut in cross section the axoplasm was immunoreactive, but the myelin sheaths were unstained.

Another notable aspect of ASPA expression was the clear demarcation of differential oligodendrocyte morphologies associated with fiber tracts composed of small diameter fibers versus those composed of large diameter fibers. Oligodendrocytes have been classified according to their morphology and the number and size of neuronal fibers they myelinate. Type I and II oligodendrocytes predominate in forebrain pathways such as corpus callosum, where fiber diameters rarely exceed 2 μm in diameter, whereas in hindbrain and spinal cord, type III and IV oligodendrocytes predominate where axon diameters often reach 4 μm or greater in diameter (Butt et al., 1998). Type I/II oligodendrocytes typically myelinate multiple smaller diameter axons, whereas type III/IV oligodendrocytes usually myelinate a single large diameter axon. ASPA expression clearly demonstrated two very different morphologies among oligodendrocytes. Consistent with the type I/II morphology, oligodendrocytes in many forebrain pathways such as corpus callosum, anterior commissure, internal capsule and olfactory tracts all had relatively small cell bodies and multiple proximal processes that branched. In contrast, many oligodendrocytes in more posterior pathways containing larger diameter axonal fibers had larger cell bodies, and often had one very large proximal process which did not branch, which is consistent with a Type III/IV morphology.

ASPA expression by region

In the telencephalon ASPA expression was most prominent in fiber tracts. Immunoreactivity in cortical areas was lower in comparison to white matter, as previously reported (Kaul et al., 1991; Madhavarao et al., 2004) but was still substantial. In isocortex ASPA was expressed predominantly in 3 cell types, pial cells at the surface, microglia in all cortical layers, and oligodendrocytes located most extensively in the deeper layers. At the cortical surface ASPA was expressed very strongly in cells of the pia mater (Figs. 2A, 2B). The pial cells were intensely stained in both nucleus and cytoplasm. Pia mater is comprised of pial cells, extracellular collagen fibrils, and meningeal macrophages. The strong ASPA expression in pia mater transitioned to very light expression in the tunica externa surrounding some arterioles penetrating the cortical surface (Fig. 2B). Some blood vessels throughout the brain were lightly immunoreactive for ASPA, and in many cases the endothelial cell nuclei were more strongly stained than their cytoplasm (Figs. 2C, 2D).

Isocortex

ASPA expression in isocortex was distributed in all layers (Fig. 3A). In isocortical layers I through III most ASPA expression was observed in microglia. ASPA-expressing microglia were highly ramified in morphology, typically issuing several major processes, which branched further at their distal ends forming a network of fine immunoreactive fibers throughout the neuropil. Layer I of isocortex contained scattered immunoreactive microglial cells (Fig. 3B). In addition to ramified microglia, a few strongly immunoreactive cells in layer I had the appearance of oligodendrocytes (Fig. 3B, arrow). Oligodendrocyte specific protein (OSP) immunoreactivity was observed in a network of very fine diameter oligodendrocyte fibers in layer I of isocortex (Fig. 3C). Myelin basic protein (MBP) was also observed in a network of lightly myelinated fibers in isocortical layer I, and these were of larger diameter than the fibers stained for OSP (Fig. 3D). Isocortical layers II and III contained many ASPA-positive microglia and an associated network of their immunoreactive processes. In layers IV through VI, numerous APSA-positive oligodendrocytes were the predominant immunoreactive cell type, and were intermingled with immunoreactive microglia. This increasing protein expression of ASPA from cortical surface to the underlying white matter is also reflected in ASPA enzyme assays showing increasing activity in tissue homogenates taken from superficial to deep layers of isocortex (Kaul et al., 1991). Finally, as was observed throughout the CNS, many cellular nuclei from unidentified cell types were lightly to moderately ASPA positive in all cortical layers. These ASPA expressing cell nuclei were relatively small in size (typically 4 to 7 micrometers in the longest dimension) indicating that many of them likely belonged to glial cells. Future dual labeling immunofluorescence experiments using markers for neurons and various glial cell types are warranted.

Comparison with expression of other oligodendrocyte markers

After mild antigen retrieval MBP immunoreactivity was greatly increased throughout the CNS. OSP immunoreactivity was also substantially increased, whereas ASPA immunoreactivity was more modestly improved. The staining pattern for ASPA in the external capsule and overlying cortex (Figs. 4A and 4B) was dissimilar to that of OSP (Figs. 4C and 4D) and MBP (Figs. 4E and 4F). OSP expression was observed in a dense network of very fine oligodendrocyte processes throughout grey and white matter (Fig. 4C). In layer V of isocortex OSP was observed in numerous fine diameter oligodendrocyte processes, with many processes being clustered and arranged in columns (Fig. 4D). MBP immunoreactivity was present in myelin sheaths, and accordingly was more intense in white matter than in grey matter (Fig. 4E). In isocortex MBP-immunoreactivity was also somewhat concentrated in columns (Fig. 4F). The pattern of fiber staining for MBP had a distinctive appearance from the staining patterns observed with OSP and ASPA antibodies. MBP

staining in myelin appeared as patchy immunoreactivity on hollow tubes associated with neuronal axons (Fig. 4F, inset).

Overall the staining patterns for the 3 proteins were disparate. ASPA was present in oligodendrocyte cell bodies, nuclei and some processes, especially larger diameter proximal processes. ASPA was also present in microglia and their processes. OSP was present in many more oligodendrocyte processes than seen with ASPA antibodies, but staining was mostly restricted to finer diameter terminal processes, and was only seen in oligodendrocyte cell bodies relatively rarely. The MBP staining pattern shared some similarities to that of OSP, but the diameter of the stained fibers was greater with MBP (myelinated axons) than OSP antibodies (oligodendrocyte processes).

Telencephalic nuclei

In the septum, caudate-putamen (CP), globus pallidus (GP) and ventral striatum, ASPA was expressed in numerous oligodendrocytes associated with fiber pathways such as the internal capsule, fornix and anterior commissure (ac; Fig. 5A). These forebrain pathways contained predominantly type I/II oligodendrocytes which issued several processes. Expression of ASPA was more pronounced in the globus pallidus than in the striatum due to the greater density of myelinated fibers (Figs. 5 B-D). Immunoreactive microglia were present throughout these areas, and were most apparent in grey matter. In the striatum, ASPA-expressing oligodendrocytes were associated with fiber bundles of the internal capsule (Fig. 5E). The medial septum contained numerous immunoreactive oligodendrocytes associated with the intercalated fibers of the fornix (Fig. 5F). Numerous ASPA-positive cell nuclei were observed throughout the telencephalic nuclei, but due to a lack of cytoplasmic staining their identity could not be determined.

Telencephalic fiber pathways

In the corpus callosum most if not all oligodendrocytes were immunoreactive for ASPA to some degree, ranging from very lightly to intensely stained (cc in Fig. 6A, see also Figs. 2A, 5A and 7A). The great majority of immunoreactive oligodendrocytes in the corpus callosum and external capsule were Type I/II morphologically, with more than one cellular process. ASPA was expressed in oligodendrocyte cell bodies, nuclei, proximal processes, and in some finer more distal processes, but it was not expressed throughout entire oligodendrocyte arborizations. In addition to the corpus callosum, all telencephalic fiber pathways contained numerous oligodendrocytes that expressed ASPA protein, including the fornix (Fig. 6A), fimbria (Fig. 6B), anterior commissure (Figs. 6A and 6C), lateral olfactory tract (Fig. 6D), internal capsule and stria terminalis (Fig. 6E) and cerebral peduncles (Fig. 6F). Many of the oligodendrocytes in these forebrain pathways exhibited Type I/II morphology, and were often arranged in rows or clusters with oligodendrocyte cell bodies in direct contact with one another.

The subfornical organ is a circumventricular organ that is outside the blood brain barrier. It contained numerous ASPA immunoreactive microglia with a relatively compact ramified morphology (Fig 6A, inset). The subfornical organ also contained numerous ASPA-positive cell nuclei which could not be positively identified due to a lack of cytoplasmic staining (Fig. 6A, arrow in inset). The fenestrated vasculature of the subfornical organ was invested with numerous ASPA-positive cells that included endothelia, microglia and possibly pericytes which surrounded the vessels.

Hippocampus

In the hippocampal formation ASPA expression was relatively light, and was observed predominantly in microglia and oligodendrocytes. ASPA expressing oligodendrocytes in the

hippocampus were densely packed throughout the alveus, fimbria (Figs. 6B and 7A) and fornix (Fig. 6A). Stained oligodendrocytes were also present in the stratum lacunosum moleculare, which contains myelinated axons from hippocampal pyramidal cells. The most numerous ASPA expressing cells in the hippocampal grey matter were microglia, which were present in large numbers throughout the hippocampal formation (Figs. 7B-F). Microglia in the hippocampus ranged from lightly to strongly immunoreactive and were present in all layers (Fig. 7B), and tended to be more strongly immunoreactive in their cytoplasm than in their nuclei (Fig. 7C, arrow). However, many microglia expressed ASPA in both cytoplasm and nucleus as was the case with other immunoreactive cell types. ASPA-expressing microglia in the hippocampus possessed highly ramified processes which formed a network of fine immunoreactive fibers present throughout the neuropil. The lowest density of ASPA-positive microglia in the hippocampus was observed in the granule cell layer (Fig. 7D).

Thalamus and hypothalamus

ASPA expression was strong in the thalamus and hypothalamus due to the large number of oligodendrocytes associated with afferent, efferent and en passant fiber pathways. Extensive oligodendrocyte ASPA expression was observed in many nuclear groups, as well as in fiber pathways such as the optic tracts, stria medularis thalami and medial lemniscus, external medullary lamina and posterior commissure (Fig. 8A). In the posterior commissure some oligodendrocytes had larger cell bodies than those observed in forebrain pathways such as corpus callosum, possessing large primary processes directed vertically in the commissure indicative of type III/IV oligodendrocytes. The optic tracts contained some oligodendrocytes with very large primary processes oriented perpendicular to the optic fibers (Figs. 8C). The optic chiasm contained numerous ASPA positive oligodendrocytes in a network of fine immunoreactive processes (Fig. 8D). ASPA expression immediately surrounding the III ventricle was relatively sparse, and was predominated by immunoreactive microglia and unidentified cell nuclei (Fig. 8E). Some immunoreactive microglial cells were adherent to the abluminal surface of the ependymal cell layer lining the ventricle. The ependymal cells were lightly stained in their cytoplasm, and some had moderately stained nuclei. The lateral hypothalamus contained numerous strongly immunoreactive oligodendrocytes associated with the medial forebrain bundle (Fig. 8F). At high magnification, the axoplasm of many of the fibers of the medial forebrain bundle was lightly to moderately immunoreactive for ASPA, but their myelin sheaths appeared unstained (Fig. 8F, inset). However, delicate oligodendrocyte processes were immunoreactive at the periphery of myelin sheaths, possibly in the terminal portions of the oligodendrocyte dendrites, or in paranodal oligodendrocyte cytoplasm.

Midbrain

ASPA expression in the midbrain was strongest in areas with high densities of myelinated fibers. ASPA expression in the periaqueductal grey was relatively light (Fig. 9A) with microglia being the most commonly stained elements there. Expression was also modest in the superficial grey matter layers of the superior colliculus, but was more pronounced in deeper layers (Fig. 9B). ASPA expression was also relatively low in the reticular portion of the substantia nigra (Figs. 9A and 9C). The central roots of the third cranial nerve (n III; oculomotor) were only lightly stained for ASPA (arrows in Figs. 9C and 9E), whereas the peripheral portions of the oculomotor nerve expressed ASPA strongly in Schwann cells (asterisks in Fig. 9E). In contrast, the central roots of nerve III were very strongly immunoreactive for MBP (arrows in Figs. 9D and 9F), but the peripheral portions of the nerve were completely unstained for MBP (asterisks in Fig. 9F) demonstrating the lack of congruence between ASPA and MBP expression.

Cerebellum

The rat cerebellum exhibited a wide range of ASPA expression, with relatively low levels in the cortex, and very high levels in white matter and deep nuclei (Fig. 10A). In cerebellar cortex, ASPA expression in the molecular layer occurred almost exclusively in ramified microglia (Figs. 10B, C and D). In contrast, a significant number of stained oligodendrocytes, microglial cells and their associated processes were present in the granule cell layer (Figs. 10B - F). Oligodendrocytes in the granule cell layer had type I/II morphology, and were interspersed among scattered microglia. In the Purkinje cell layer, strong expression was observed in oligodendrocytes, and faint ASPA expression was observed in cells that may have been Bergmann glial cells, but Purkinje cells were unstained (Fig. 10D). White matter contained numerous strongly stained oligodendrocytes and some immunoreactive microglia (Figs. 10B, C, E and F).

Brainstem

The brainstem of the rat exhibited some of the strongest ASPA expression in the CNS due to the great density of heavily myelinated fiber tracts (Fig 11A). Oligodendrocytes with both type I/II morphology and type III/IV morphology were observed in the anterior medullary velum at the roof of the fourth ventricle (Fig. 11B). Many of the oligodendrocytes in the brainstem and medulla had a single large process indicative of type III/IV oligodendrocytes (Figs. 11C - E). In a number of fiber pathways such as the spinal tract of the trigeminal nerve, type III/IV oligodendrocytes were associated with large diameter axons with unstained myelin sheaths, and ASPA-positive axoplasm (Fig. 11C, see inset). In cross section these appeared as light toroids (unstained myelin sheaths) with dark centers (stained axoplasm; Figs. 11C and D). Oligodendrocytes with type III/IV morphology were also observed in other fiber pathways such as the medial cerebellar peduncle and the trapezoid body (Figs. 11D and E). Microglial cells and pial cells also expressed ASPA, as in other parts of the CNS. Choroid plexus epithelial cells within the IV ventricle were unstained. However, the ependymal cells of the choroid plexus expressed ASPA at moderate levels (Fig. 11F). These ventricular macrophages, also known as Kolmer cells, are adherent to the choroid plexus epithelium and act as ventricular phagocytes.

Medulla

ASPA expression in the medulla was very strong, with a high density of immunoreactive cells in fiber pathways and a lower density of immunoreactive cells in cranial nerve nuclei and the medullary reticular formation (Fig. 12A). Oligodendrocytes in fiber pathways expressed ASPA most strongly, and exhibited both type I/II and type III/IV morphology due to the mixed population of large and small diameter nerve fibers. Nuclear groups such as the nucleus of the solitary tract and inferior olive had relatively lower levels of expression, and contained many immunoreactive cell nuclei and microglia (Figs. 12A and B). Oligodendrocytes and the axons of trigeminal neurons were stained in the spinal tract of the trigeminal nerve (Fig. 12C). The ependymal cells lining the central canal were also lightly to moderately immunoreactive for ASPA (Fig. 12D). Some neurons in the brainstem expressed ASPA at low to moderate levels, for example in the accessory inferior olive and lateral reticular nucleus (Figs. 12E and F). Many of these neurons were not stained any more strongly than their surrounding neuropil, making them difficult to image. Unlike other cells types which showed both cytoplasmic and nuclear staining, ASPA expression in neurons was typically restricted to the cytoplasm, and excluded from their nuclei. Overall, the majority of neuronal cell bodies did not express detectable levels of ASPA despite many of the axons originating from those cranial nerve nuclei being moderately stained.

ASPA and AceCS1 expression in neuronal axons

The fiber pathways containing ASPA-expressing axons in the hindbrain also expressed AceCS1 (Fig 13). This was particularly evident in several cranial nerve pathways, including the spinal tract of the trigeminal nerve, and the 7th and 8th cranial nerves. In the spinal tract of the trigeminal nerve both AceCS1 (Fig. 13A) and ASPA (Fig. 13B) immunoreactivity in axons cut on end had a characteristic toroidal appearance with stained central axoplasm and a surrounding unstained ring of myelin. In the facial (7th) nerve, many axons were immunoreactive for both AceCS1 (Fig. 13C) and ASPA (Fig. 13D). AceCS1 was expressed predominantly in the nuclei of oligodendrocytes in the nerve, whereas ASPA was expressed strongly in their nuclei, cytoplasm and some processes. In the 8th nerve, both AceCS1 (Fig. 13E) and ASPA (Fig. 13F) were observed in many axons. Dual labeling immunofluorescence studies will be necessary to determine to what extent both proteins are expressed in the same axons.

Spinal cord

ASPA expression in the spinal cord was strong in oligodendrocytes, and moderate in some neurons and many axons (Fig. 14A). Strong expression was observed in the dorsal columns, with the pyramidal tracts and cuneate fasciculi having the highest immunoreactivity (Figs. 14A and 14B). More neuronal cell bodies expressed ASPA in the spinal cord grey matter than any other region of the CNS (Figs. 14C and 14D). The most immunoreactive neurons were motor neurons of layer 9 (Fig 14D). Axonal fibers of the pyramidal tracts (Figs. 14 A and B) and the axons of the cuneate and gracile funiculi were immunoreactive seen cut on end, whereas myelin sheaths appeared unstained (Figs 14E and F). Many axonal fibers in the lateral and ventral funiculi were also immunoreactive.

Discussion

Confirming and expanding upon our earlier findings with a different anti-ASPA antibody (Madhavarao et al., 2004) we found that most cell types in the brain can express ASPA indicating a broader functional repertoire for NAA, and NAA-derived acetate and aspartate, than previously recognized. All ASPA-immunoreactive elements observed in wild type rat brain sections were absent in brain sections from tremor rats, a mutant rat strain which lacks the entire ASPA gene sequence. The previous ASPA antibody used in our laboratory was produced against purified full-length recombinant ASPA protein. That antibody and the one used in the current study both stained the same cellular elements. Astrocytes were the only major cell type in the brain that did not express ASPA in their cytoplasm at detectable levels with either antibody.

ASPA expression in oligodendrocytes

ASPA expression was observed extensively in oligodendrocytes throughout the CNS, with virtually all oligodendrocytes showing some level of ASPA expression. ASPA immunoreactivity in oligodendrocytes was often uniformly strong in both the cytoplasm and nucleus, but in some the staining was stronger in one subcellular compartment or the other. This finding supports previous results showing that ASPA is transported between cytoplasmic and nuclear compartments (Hershfield et al., 2006). ASPA was also expressed in some oligodendrocyte processes, but was absent in many processes and in compact myelin. However, ASPA was expressed immediately adjacent to myelin sheaths in terminal oligodendrocyte processes and possibly in paranodal oligodendrocyte cytoplasm.

Oligodendrocytes are the macroglial cells responsible for producing myelin sheaths in the CNS, and they employ bidirectional trophic and metabolite exchanges with neuronal axons to accomplish this highly specialized cellular behavior (Ledeen, 1984; Nave, 2010; Popko,

2003). NAA is one of these trophic molecules supplied by neurons, and NAA-derived acetate contributes to myelin lipid synthesis in oligodendrocytes during postnatal development (reviewed in Moffett et al., 2007). The contribution of NAA derived acetate to myelination is most critical during the period of postnatal myelination, at which time it constitutes approximately 30% of the carbon incorporated into certain myelin associated lipids including cerebroside (Madhavarao et al., 2005; Traka et al., 2008; Wang et al., 2009). The bulk of acetyl CoA utilized to synthesize myelin lipids comes from citrate synthesized in oligodendrocyte mitochondria. Citrate is produced in mitochondria from pyruvate in the TCA cycle and is then exported to the cytoplasm. The enzyme ATP citrate lyase converts cytosolic citrate into acetyl CoA, which then is used to synthesize the fatty acids and sterols needed for myelination. The NAA-derived acetate produced in oligodendrocytes through the action of ASPA can not be used for lipid synthesis until it is converted to acetyl CoA. This is accomplished by acetyl coenzyme A synthase-1 (AceCS1), which is present in many oligodendrocytes during postnatal myelination (Ariyannur et al., 2010a). AceCS1 catalyses the reaction between free acetate and coenzyme A to produce acetyl CoA, which then can participate in cellular metabolism. Therefore, there are two parallel pathways of acetyl CoA generation in oligodendrocytes for lipid synthesis during myelination, the primary citrate/ATP citrate lyase pathway and the secondary NAA/ASPA-AceCS1 pathway (Moffett et al., 2007; Arun et al., 2010; Ariyannur et al., 2010a).

ASPA has been reported to be present in the brain primarily as a soluble enzyme in the cytosol fraction (D'Adamo, Jr. et al., 1973; D'Adamo, Jr. et al., 1977; Klugmann et al., 2003; Madhavarao et al., 2004). However, other studies have provided evidence that some proportion of brain ASPA is associated with myelin (Chakraborty et al., 2001; Wang et al., 2007), and that detergent is required to dissociate this fraction of enzyme activity from membranes or myelin (Goldstein, 1976; Wang et al., 2007). In the current study, ASPA expression in white matter was observed in some oligodendrocyte processes associated with neuronal axons that could represent localization within myelin sheaths, or the cytoplasmic layers associated with myelin. However, the staining patterns for ASPA and two myelin associated proteins, MBP and OSP, were not similar at low or high magnification (Figs. 3, 4 and 9). At higher magnification, ASPA immunoreactivity in major caudal fiber pathways was clearly present in the axoplasm of many axons cut on end, but was not observed in the ensheathing myelin (Figs. 8, 11 and 14). In contrast, MBP and OSP were associated with myelin sheaths and oligodendrocyte processes respectively.

ASPA expression in microglia and meninges

Microglia expressed ASPA throughout the CNS and were more apparent in cortical and grey matter areas than in fiber pathways. Previously we reported that some microglia were lightly to moderately immunoreactive for ASPA in the rat CNS using a different ASPA antibody (Madhavarao et al., 2004). In the current study utilizing purified ASPA peptide antibodies we observed that microglia were immunoreactive throughout the CNS, and ranged from lightly to strongly immunoreactive. All ASPA expressing microglia were highly ramified in morphology indicating that they were in a resting, non-immunogenic state. Microglia are related to macrophages (Streit et al., 1993; Guillemin et al., 2004), and we observed that macrophages associated with the choroid plexus, so-called epiplexus cells (Ling et al., 1998) also expressed ASPA moderately (Fig. 11F). ASPA expressing microglia were also observed in the subfornical organ (Fig. 6A, inset), a circumventricular organ whose cells lie outside the blood brain barrier. These findings suggest that ASPA expression may be common among phagocytic cells, and in fact, we have observed strong ASPA expression in apparent macrophages throughout the rat spleen (unpublished observations). Recently the N-acetyltransferase-8-like gene has been shown to be the gene encoding the NAA synthesizing enzyme aspartate N-acetyltransferase (Wiame et al., 2010; Ariyannur et al., 2010b). This

gene was found to be expressed in only 3 of the tissues examined; brain, and to lesser extents in spleen and thymus (Wiame et al., 2010). This is consistent with some immune cell types being capable of synthesizing NAA.

Leptomeningeal cells of the pia mater expressed ASPA very strongly, and some endothelial cells of the cerebral vasculature expressed low levels of ASPA. Ependymal cells also expressed ASPA at low to moderate levels. It is possible that ASPA expression in the meninges, ependymal cells and vasculature could provide a mechanism for scavenging NAA from the cerebral circulation and cerebrospinal fluid allowing uptake of the resultant aspartate and acetate for recycling to the brain. This possibility is consistent with observation that NAA levels drop as cerebrospinal fluid passes from ventricles (~55 μ M) to the subarachnoid space (~9 μ M) and then to lumbar spinal cord (~1 μ M) (Faull et al., 1999). Retention of the resultant metabolites would be more critical under low nutrient conditions, or at times of high metabolic demand.

ASPA and ACS1 expression in cell nuclei

In every region of the CNS examined the nuclei of oligodendrocytes, microglial cells, endothelial cells, ependymal cells, epiplexus cells and leptomeningeal cells, as well as numerous unidentified cell nuclei, were immunoreactive for ASPA. This finding is highly relevant to the recently described localization of AceCS1, which was found to be localized predominantly to cell nuclei in the adult rat brain (Ariyannur et al., 2010a). As noted above, AceCS1 is present in the cytoplasm and nuclei of oligodendrocytes during postnatal myelination, but the expression pattern shifts to a predominantly nuclear localization in adult rats (Ariyannur et al., 2010a). AceCS1 was also expressed in the nuclei of neurons and astrocytes, as well as in many unidentified cell nuclei in all regions of the CNS. There are only two known forms of acetyl coenzyme A synthase, types 1 and 2, with type 2 being localized to mitochondria where it synthesizes acetyl CoA for energy derivation (Fujino et al., 2001; Sakakibara et al., 2009). As such, AceCS1 is the only known enzyme capable of converting free acetate into acetyl CoA in cell nuclei, where AceCS1 may be acting to recycle acetate derived from deacetylation reactions such as histone deacetylase activity. In addition to AceCS1 acting to recycle acetate derived from deacetylase reactions, mounting evidence supports a role for NAA and nuclear localized ASPA in providing some of the acetate for protein acetylation reactions including histone acetylation. In such a scenario, NAA present in the nucleus is broken down by nuclear-localized ASPA, and the free acetate is then converted to acetyl CoA by the action of AceCS1. The acetyl CoA produced can then be utilized by histone acetyltransferase enzymes for nuclear histone acetylation reactions involved in chromatin remodeling and epigenetic gene regulation. Recently we found that AceCS1 expression in oligodendrocyte and neuronal cell nuclei is increased in the tremor rat model of Canavan disease, and that chronic treatment with a hydrophobic acetate precursor normalizes AceCS1 expression levels in this model, suggesting that the lack of NAA-derived acetate leads to altered acetyl CoA metabolism in the CNS (Arun et al., 2010; Ariyannur et al., 2010b). The important role of epigenetic gene regulation in oligodendrocyte maturation has been reviewed recently (Coprav et al., 2009; Liu et al., 2010).

In single celled eukaryotes such as yeast AceCS1 is a primary source of acetyl CoA required for histone acetylation reactions and gene transcription (Takahashi et al., 2006). In contrast, mammalian cells derive most of their acetyl CoA for histone acetylation reactions via the ATP citrate lyase pathway (Wellen et al., 2009). This is analogous to the roles of these two enzyme systems in mammalian lipid synthesis. Therefore in mammals both lipid synthesis and histone acetylation derive most of their acetyl CoA from glucose (pyruvate) by way of citrate and ATP citrate lyase. The role of AceCS1 in histone acetylation in mammals appears to be a secondary, parallel pathway for acetyl CoA production.

ASPA expression in neurons and their axons

In the current study we confirm our previous finding that ASPA is expressed in a substantial number of axons in the rodent brain. In a previous study using a different anti-ASPA antibody, we observed mild to moderate ASPA-immunoreactivity in some large neurons in the brainstem and spinal cord, as well as in numerous axons in certain fiber tracts (Madhavarao et al., 2004). In that study, we did not have access to tremor rat tissue, and therefore could not definitively confirm that all immunoreactivity was due to ASPA protein expression. In the present study using the purified ASPA peptide antibody, some neurons were found to express ASPA protein in their perikarya (Figs. 12 E, F, 14 C and D), but tremor rat brain sections showed no immunoreactivity, indicating that the neuronal staining in wild type rats was specific for ASPA. Most neurons did not express detectable ASPA levels in their cytoplasm, but many did express ASPA in their axoplasm, and this was especially apparent in long axonal projections comprised of medium to large diameter fibers, such that they were associated with Type III/IV oligodendrocytes. The lack of ASPA expression in most neuronal cell bodies with concomitant expression in their axons is suggestive of rapid transport of newly synthesized ASPA protein from perikarya to axons.

Previously, it has been assumed that the synthetic and degradative compartments for NAA were invariably located in distinct cell types wherein neurons synthesized NAA, and subsequently transfer it to oligodendrocytes for catabolism (Baslow et al., 1999; Baslow, 2010; Baslow, 2003). Based on the current findings that ASPA is expressed in axons, this assumption must be reexamined. Many of the fiber pathways found to express ASPA in the current study have also been reported to contain both NAA, and the related neuronal dipeptide N-acetylaspartylglutamate (NAAG) (Moffett et al., 1993; Moffett et al., 2006). The observation that a substantial number of axons express low to moderate levels of ASPA indicates that NAA can be both synthesized and degraded in a single cell type within the CNS. A number of caudal fiber tracts such as the spinal tract of the trigeminal nerve, the spinocerebellar tract, several cranial nerves, the pyramidal tracts and the dorsal columns were moderately immunoreactive for ASPA in their axoplasm. The expression of ASPA in the axons of projection pathways and nerves exhibited a generally increasing gradient of expression from forebrain to hindbrain, but this may have been due in part to the diameter of axons in forebrain and hindbrain. It was difficult to image individual axons in forebrain fiber pathways due to their smaller diameter, but the axoplasm of fibers in the medial forebrain bundle in the lateral hypothalamus were immunoreactive for ASPA (Fig. 8F). In the current study we also observed AceCS1 expression in many of the same axonal pathways (Fig. 13) suggesting that NAA-derived acetate can be converted to acetyl CoA for further metabolism in some axons. However, determining the extent of ASPA and AceCS1 colocalization in axons will require future dual labeling immunofluorescence studies.

Existing evidence indicates that NAA is synthesized primarily in neurons from acetyl CoA and aspartate and that a substantial portion is stored in neurons, whereas some is exported to other cell types including oligodendrocytes. It is likely that NAA is synthesized in neurons when acetyl CoA is produced in excess of local requirements, for example when glucose levels are high. The acetyl CoA is converted to NAA in neurons and is stored for later use. It is possible that the low synthesis rate reported for NAA under resting conditions (Harris et al., 2006) is a reflection of this storage role. As metabolic demands outweigh local glucose and pyruvate supplies, some of the stored NAA is broken down by ASPA in order to regenerate acetyl CoA, either in the axon of same neuron or more generally, in cells in contact with the neuron after intercellular NAA transport. Virtually all cell types in the brain are in direct contact with neurons including oligodendrocytes, astrocytes and microglia (Peters et al., 1991), and are therefore in a position to receive intercellularly transported NAA directly via cell contacts. When neuronal activity is high, NAA is transferred to neighboring cells, especially oligodendrocytes, in order to support increased metabolic and

physiological demands. NAA may therefore be acting as a storage and transport form of acetyl CoA (and aspartate) critical for proper nervous system development and function (Ariyannur et al., 2010b; Arun et al., 2010). Because acetyl CoA participates in so many diverse cellular functions, NAA may also participate in the same functions by delivering acetate (as well as aspartate) to the many cell types in the brain that express ASPA.

One way in which NAA could support brain metabolism when neural activity levels were high would be to substitute for cytoplasmic citrate in cells that express both ASPA and AceCS1 (Fig. 15). Each molecule of acetyl CoA produced from NAA-derived acetate means one less citrate molecule must be exported from local mitochondria. As such, preexisting intramitochondrial acetyl CoA is spared from being converted to citrate, and instead can be used to generate energy via the TCA cycle. NAA delivery to the cytoplasm (and nuclei) of oligodendrocytes, microglia and other cell types means that their mitochondria can use their current acetyl CoA complement for oxidative phosphorylation, rather than to synthesize and export citrate. NAA can substitute by providing both acetyl CoA and oxaloacetate (via aspartate) in the cytoplasm, as would be the case when ATP citrate lyase breaks down citrate into acetyl CoA and oxaloacetate. These emerging views on functional roles of ASPA suggest a much more complicated picture than previously recognized.

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Figure 1.

Figure 2.

Figure 3.

Figure 4.

Figure 5.

Figure 6.

Figure 7.

Figure 8.

Figure 9.

Figure 10.

Figure 11.

Figure 12.

Figure 13.

Figure 14.

Figure 15.