Published in final edited form as:

Biol Psychiatry. 2010 January 1; 67(1): 12–19. doi:10.1016/j.biopsych.2009.08.028.

Meta-analysis of 15 genomewide linkage scans of smoking behavior

Shizhong Han¹, Joel Gelernter^{1,2,3,4}, Xingguang Luo^{1,4}, and Bao-Zhu Yang^{1,4,*}

¹Department of Psychiatry, Division of Human Genetics, Yale University School of Medicine, New Haven, CT 06511, USA

²Department of Genetics, Yale University School of Medicine, New Haven, CT 06511, USA

³Department of Neurobiology, Yale University School of Medicine, New Haven, CT 06511, USA

⁴VA CT Healthcare Center 116A2; 950 Campbell Avenue; West Haven, CT 06516

Abstract

Background—A genetic contribution to smoking behavior is well established. To identify loci that increase the risk for smoking behavior, many genomewide linkage scans have been performed using various smoking behavior assessments. Numerous putative susceptibility loci have been identified, but only a few of these were replicated in independent studies.

Methods—We used genome search meta-analysis (GSMA) to identify risk loci by pooling all available independent genome scan results on smoking behavior. Additionally, to minimize locus heterogeneity, subgroup analyses of the smoking behavior assessed by the Fagerstrom test for nicotine dependence (FTND) and maximum number of cigarettes smoked in a 24-hour period (MaxCigs24) were carried out. Samples of European ancestry were also analyzed separately.

Results—A total number of 15 genome scan results were available for analysis, including 3404 families with 10,253 subjects. Overall, the primary GSMA across all smoking behavior identified a genomewide suggestive linkage in chromosome 17q24.3–q25.3 ($P_{SR}=0.001$). A secondary analysis of FTND in European-ancestry samples (625 families with 1878 subjects) detected a genomewide suggestive linkage in 5q33.1–5q35.2 ($P_{SR}=0.0076$). Subgroup analysis of MaxCigs24 (966 families with 3273 subjects) identified a genomewide significant linkage in 20q13.12–q13.32 ($P_{SR}=0.00041$, $P_{OR}=0.048$), where a strongly supported ND candidate gene, CHRNA4, is located.

Conclusions—The regions identified in the current study deserve close attention and will be helpful for candidate gene identification or target resequencing studies in the future.

Keywords

smoking behavior; nicotine dependence; FTND; MaxCigs24; genetic linkage; meta-analysis

© 2010 Society of Biological Psychiatry. Published by Elsevier Inc. All rights reserved.

*Corresponding Author: Bao-Zhu Yang, PhD; Yale University School of Medicine, Department of Psychiatry, Division of Human Genetics in Psychiatry; VA CT 116A2; 950 Campbell Avenue; West Haven, CT 06516; telephone: 203-932-5711 ext 3590; fax, 203-937-4741; bao-zhu.yang@yale.edu.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Introduction

Cigarette smoking is highly prevalent throughout many populations around the globe. Despite increasing awareness of the risks associated with smoking, the World Health Organization (1999) estimated that 1.1 billion people still smoke, and predicted that by 2025, the number will increase to 1.6 billion worldwide. Thus, understanding various factors that influence smoking behavior is critical to the prevention and cessation of smoking. Although the etiology of smoking behavior is complex, a genetic contribution to smoking behavior, presumably based on addiction to nicotine, is well established from twin and adoption studies (1,2). Genetic linkage analysis can be a useful design to detect genes that segregate in families, including common variants, multiple rare variants within one locus, and copy number variation.

More than 20 genomewide linkage scans on smoking behavior have been performed using a variety of smoking behavior assessments, including DSM-IV defined nicotine dependence, the Fagerstrom Test for Nicotine Dependence (FTND) (3,4), the Fagerstrom Tolerance Questionnaire (FTQ), Fagerstrom-derived smoking quantity (SQ) and Heaviness of Smoking Index (HSI), habitual smoking, persistent smoking, maximum number of cigarettes smoked in a 24-hour period (MaxCigs24), and others (5–24). Numerous putative susceptibility loci have been identified, but only a few of these have been replicated in independent studies, which is not uncommon for linkage analysis of genetically complex traits. Considering the high likelihood of many risk loci of low-to-moderate effect for complex traits and the relatively small sample size in each study, the discrepancy among study results is expected, because a single study may be statistically underpowered to detect a low magnitude but real genetic linkage. Although literature reviews have provided valuable overviews of progress in linkage studies on smoking behavior, they are not intended to provide formal statistical assessment of pooled evidence for linkage across studies. Considering the quantity of accumulated genome scan results on smoking behavior now available, a rigorous statistical method of synthesis of the reported results for linkage could provide a powerful approach to detect previously unappreciated linkage signals.

The genome search meta-analysis (GSMA) method has been proposed as a valid and robust meta-analysis technique to combine the evidence for linkage across multiple linkage scans using a non-parametric ranking method (25,26). Apart from the advantage of greater power in detecting small but consistent evidence for linkage, GSMA can combine linkage results from studies with different family structures, marker sets, and statistical analysis methods. While a unique genetic spectrum might characterize each specific smoking behavior, we hypothesize that some risk loci are shared across different assessments. Consequently, the aim of the current study is to identify potential risk loci which are independent of distinct smoking behavior assessments using the GSMA by pooling all independent genome scan results of smoking behavior. Because increased sample homogeneity can be helpful to reduce locus heterogeneity and therefore increase power to detect regions specific for a particularly defined sample set, subgroup GSMA based on FTND and MaxCigs24 was carried out. Samples incorporating subjects of mostly European ancestry were also analyzed separately.

Materials and Methods

Study samples

To identify existing genomewide linkage studies on smoking behavior, we conducted a computerized literature search of the PubMed database using the following keywords and subject terms: 'linkage', 'smoking', 'nicotine dependence', 'genome-wide', or 'genomewide'. Review articles on genetics of smoking behavior were also screened. The genome scans included in the current GSMA were required to meet the following criteria: 1) whole genome linkage scan on smoking related traits performed in humans; 2) whole genome linkage results

either available from the original investigators, or extractable from published graphs; 3) samples used in genome scans should be independent of each other. Linkage studies which were repeated analyses of the same sample using different statistical methods or phenotype measures were identified, and we included only one independent study for which the whole genome linkage results were available. In addition, when a study reported whole genome linkage results on different samples; we treated each sample as a separate genome scan. When a study reported a two-stage analysis, only the original results were used and any follow up studies in candidate regions were excluded, as the GSMA requires a uniform distribution of markers across the genome (25).

In total, 20 studies (24 genome scans, 5428 families) were identified (5–24), and finally 12 studies (15 complete genome scans, 3404 families) met criteria and were included in the GSMA (5–16), as listed in Table 1. Eight studies (9 genome scans, 2024 families) were not included for the following reasons (17–24). Four studies are repeated analyses of the Framingham Heart Study (6,19–21); from these, the result from Goode (6) was used because the whole-genome linkage result was available from the published graph. Three linkage analyses have been performed on the data of the Collaborative Study on the Genetics of Alcoholism (7,17–18); we used the result from Bierut (7) since the whole-genome linkage results were available from the authors. Two linkage analyses on two different assessments of smoking behavior (9,22) were performed for the same sample from the Netherlands Twin Register; the result for “maximum number of cigarettes per day” was included (9). In the earliest study, the authors carried out a 2-stage genome scan, and only the result from stage 1 was used since stage 2 was a follow up genome scan in candidate regions (5). Finally, results for two studies (three genome scans) were not available (23,24). In addition, two studies (15,16) used the same sample of Finnish twin families, but performed the genome scan on different assessments of smoking behavior (FTND and MaxCigs24). The result from Loukola (16) was included for the primary GSMA on smoking behavior and FTND. The result from Saccone (15) was only used for the secondary GSMA on MaxCigs24. Consequently, the data for this analysis were collected from 12 studies (15 genome scans), including 3404 families with 10,253 individuals.

Data extraction

The relevant characteristics of each study included in the GSMA are summarized in Table 1. For each study, the following information was extracted: first author, journal, year of publication, ethnicity of study population, number of families, subjects, definition of phenotype, number of markers, linkage statistic, and software used for linkage analysis. If the genomewide linkage results were available from the published graphs, the required linkage statistics for GSMA were extracted from the figures by the digital software g3data (<http://www.frantz.fi/software/g3data.php>). Otherwise, the authors were invited to contribute the whole genome linkage results including marker names, genetic position and linkage statistics for each marker. The authors of five studies including seven genome scans (7,13–16) provided the original linkage results. Genetic map positions and marker locations were unified based on the Marshfield genetic map.

Statistical analysis

The GSMA method was used to synthesize the evidence for linkage across multiple genome scans. In the primary GSMA, chromosomes are divided into approximately equal length bins traditionally ~30 cM, generating a total number of 118 bins on the autosomes based on the Marshfield genetic map. The notation “c.n” for the bin numbering is used to refer to the nth bin on chromosome c. For each study, each bin was assigned a within-study rank by its highest LOD, NPL or Z score or minimum *P*-value, so that the bin with the highest linkage score or minimum *P*-value is assigned a rank 118 and other bins are ranked in the descending order of their strength for linkage. The ranks were then summed across studies for each bin to obtain

the summed rank (SR), which forms the basic test statistic for assessing linkage within the bin. Bins with high SR may show significant evidence for linkage.

We used GSMA software (27) to evaluate empirically the significance of the SR. Briefly, for each study, the observed rank values were randomly reassigned to 118 bins, allowing for tied ranks in each study to be incorporated in the null distribution. Bin ranks were summed across studies; this procedure was repeated 10,000 times. The empirical P -value of SR was calculated by counting the proportion of bins in which a summed rank value was equal to or larger than the observed one. In addition, a P_{OR} is calculated as the proportion of the simulated n^{th} highest ordered sum rank (OR), which is equal to or greater than the observed n^{th} highest summed rank through the same permutation procedure. Simulation studies have shown that bins with significant P -values ($P < 0.05$) of both SR and OR are likely to identify true linkage signals (26). By applying a Bonferroni correction for multiple testing (assuming 118 independent bins), values of P_{SR} , $0.05/118=0.00042$ and $1/118=0.0085$ correspond to the genomewide significant and suggestive evidence for linkage, respectively. Simulation studies have shown that these thresholds are appropriate for the GSMA (28), with a P -value exceeding the genomewide significant threshold expected once by chance in 20 meta-analyses and a P -value exceeding the genomewide suggestive threshold expected once by chance per single meta-analysis.

We performed both unweighted and weighted GSMA analysis. In unweighted analysis, each study was assumed to contribute equally to the GSMA. The weighted GSMA takes into account the relative contribution from each study. The most appropriate weighting factor is not obvious; simulation studies have shown that the square root of the number of affected cases within each study performed well (26). Since most of the phenotypes we included in this GSMA were quantitative traits (FTND, MaxCigs24) and not binary outcomes, we used the square root of the number of genotyped subjects in each study as the primary weighting factor and the relevant results were reported in detail. To evaluate the influence of different weighting factors on the results, we also used an alternative weighting factor defined by the square root of number of pedigrees $\times$ number of markers used in each study (although we note that the latter is an imperfect approximation of information content in each study owing to varying information content from different markers, and diminishing information beyond a marker set that achieves genomewide coverage).

A bin width of around 30 cM is used in the GSMA most frequently, and was demonstrated to be optimal by simulation (26). In order to detect weak linkage signals near the boundary of two bins, a shifted 30cM bin GSMA was also applied by moving bin boundaries 15 cM and starting at the midpoint of the bins used in the primary 30 cM analysis (29).

Two major sources of heterogeneity in linkage studies arise from differing definitions of smoking behavior and ethnicities of samples, since a variety of smoking behavior assessments were included in the current GSMA across different populations. Although we postulated that some genes that regulate smoking behavior might be independent of specific smoking behavior assessments and population groups, restricting the combined analyses to studies with similar ascertainment criteria or to subjects from similar ethnic backgrounds could potentially increase power to detect linkage to particular regions where the relevant risk loci are specific either to a trait or to a population. Therefore, subgroup analyses of the families assessed by FTND or MaxCigs24 were performed. Subjects of European ancestry were also analyzed separately. Other populations or phenotype groups had too few studies available for separate analysis.

Results

First, we performed the primary 30 cM bin width GSMA over all independent genome scans on smoking behavior, encompassing 3404 families with 10,253 genotyped subjects. Figure 1

illustrates the weighted and unweighted P_{SR} for all bins across the genome. The full details of genetic regions showing bins with nominal significance in weighted analysis are shown in Table 2, and the unweighted analysis results are also included as a comparison with weighted analysis. The strongest evidence for a smoking behavior risk locus was found on chromosome 17q24.3–q25.3 (bin 17.4) where suggestive evidence for linkage was achieved by either unweighted ($P_{SR} = 0.002$) or weighted analysis ($P_{SR} = 0.001$).

To identify loci that might increase susceptibility to a specific smoking behavior trait, we performed subgroup GSMA over the genome scan results on the smoking behaviors measured by FTND and MaxCigs24. Five genome scan results were included in the analysis of FTND (1347 families with 3995 subjects). The weighted and unweighted P_{SR} for all bins across the genome are illustrated in Figure 2. There were no regions that achieved genomewide significant or suggestive evidence for linkage, except six regions showed nominally significant evidence for linkage in the weighted analysis (Table 3). In the subgroup analysis of MaxCigs24, four genome scan results, all from European ancestry populations, were included (966 families with 3273 subjects). The genomewide results with weighted and unweighted analysis are illustrated in Figure 3. A genomewide significant linkage was identified in 20q13.12–q13.32 by both weighted ($P_{SR}=0.00041$, $P_{OR}=0.048$) and unweighted analysis ($P_{SR}=0.00032$, $P_{OR}=0.037$). Three regions (22q12.3–q13.32, 20p12.1–q13.12 and 17q24.3–q25.3) achieved genomewide suggestive evidence for linkage. Eleven regions on six chromosomes (2,12,16,17,20 and 22) with both P_{SR} and P_{OR} less than 0.05 are somewhat likely to harbor risk loci for MaxCigs24 trait (Table 4).

To increase homogeneity and evaluate population specific linkages, samples of European ancestry (or mostly European ancestry) were analyzed separately. Since the primary GSMA on MaxCigs24 only included European ancestry populations, secondary analysis by European ancestry only needed to be carried out for smoking behavior and FTND. Figures S1 and S2 in the Supplement illustrate the results across all bins for smoking behavior and FTND, respectively. Here, we report the results of the European ancestry GSMA compared with the overall GSMA in weighted analysis. Briefly, 11 genome scan results were included for analysis of smoking behavior (2486 families with 7270 subjects). Bin 17.4 still achieved suggestive linkage ($P_{SR}=0.002$). Three bins (11.1, 3.8 and 5.4) were no longer significant and two bins (22.2 and 5.6) became nominally significant (Table 2). For the FTND phenotype, three genome scan results were included (625 families with 1878 subjects). Two bins (6.6 and 5.4) were no longer significant and 2 bins (5.7 and 5.6) became significant (Table 3).

In order to identify weak linkage signals near the boundaries of two bins, we re-analyzed the data using a shifted 30cM bin GSMA starting at the midpoint of the bins used in the primary analysis. Figures S3, S4 and S5 in the Supplement illustrate the genomewide results using the shifted 30cM GSMA for smoking behavior, FTND and MaxCigs24, respectively. Here, we report the bins which achieved genomewide suggestive evidence for linkage (weighted analysis) in this secondary analysis. In the analysis of smoking behavior, bin 16.1 (16p13.2–16p12.1), achieved genomewide suggestive linkage in both all ($P_{SR} = 0.0074$) and European-ancestry ($P_{SR}=0.0085$) samples. Bin 5.6 (5q33.1–5q35.2) reached genomewide suggestive linkage for FTND in European-ancestry samples ($P_{SR}=0.0076$). In the analysis of the MaxCigs24 trait, bin 22.1 (22q11.22–22q13.2) reached genomewide suggestive linkage ($P_{SR}=0.0016$).

Lastly, to evaluate how the different weighting factors could influence the results, we performed the weighted analysis using the weighting factor defined by the square root of number of pedigree $\times$ number of markers. As a result, we found that the resultant top ranked bins and P -values, generated by the two weighting factors, remain close to each other. Wilcoxon signed-rank test further showed no significant difference ($P > 0.7$) on the ranks of

bins by the two different weighting factors for each trait we investigated. A comparison of the top five bins identified for each of the traits using both weighting approaches is shown in Table S1 in the Supplement.

Discussion

The current GSMA, which included 3404 families with 10,253 subjects, has identified many regions with varying degrees of evidence of linkage for smoking behavior. In the primary 30 cM GSMA of combined smoking behavior, genomewide suggestive linkage was detected at chromosome 17q24.3–q25.3. The fact that we did not identify any bins with genomewide significant evidence for linkage in the primary analysis might imply the possible relatively higher genetic heterogeneity due to a variety of different smoking behaviors and sample ancestry. Although only nominal significance was detected in the primary GSMA for the FTND, genomewide suggestive linkage was observed in 5q33.1–5q35.2 by shifting the bin boundary 15 cM in a secondary analysis of European-ancestry samples. Subgroup analysis of the MaxCigs24 phenotype identified numerous linkage signals; this might be attributable to the improved power from the increased homogeneity of both the phenotype and sample ancestry. For example, a genomewide significant linkage in bin 20.3 (20q13.12–q13.32) was identified, and the adjacent bin 20.2 (20p12.1–q13.12) showed suggestive linkage, providing more support for a true linkage signal in this region. Eleven regions with both P_{SR} and P_{OR} < 0.05 support that some or all of these 11 regions are likely to harbor risk loci for the MaxCigs24 trait.

A necessary follow-up step is to evaluate if notable candidate genes map to regions nominated by the current GSMA. A strong candidate gene for nicotine dependence, *CHRNA4* (20q13.2–q13.3) (30–34), is located in bin 20.3 (20q13.12–q13.32), where genomewide significant linkage was reached in the primary GSMA of MaxCigs24. *CHRNA4*, which encodes the nicotinic acetylcholine receptor α_4 subunit gene, is highly expressed in the central nervous system (CNS) and plays a major role in tolerance, reward, and the modulation of mesolimbic dopamine function, all of which are critical to the development of nicotine dependence (35). Two genes, *PLEKHG1* (36) and *OPRM1* (37), are located in bin 6.5 (6q23.2–q25.3), which ranks highest and its adjacent bin 6.6 ranks second highest in the GSMA of FTND. The *PLEKHG1* gene contains a pleckstrin homology (PH) domain and is expressed in the brain and peripheral nervous system (38). It is possible that variants of these PH-domain-containing proteins have an impact on the cell-signaling pathways that regulate neuronal plasticity, and thus could influence predisposition to ND. The μ -opioid receptor gene *OPRM1* has been found to be associated with FTQ nicotine dependence (37) and plays a role in substance use and dependence across several drug classes (39–41). Two other previously identified candidate genes, *DRD4* (42,43) and *COMT* (44), are located at bin 11.1 and bin 22.1 respectively, each of which showed nominal significance in the primary GSMA of smoking behavior or MaxCigs24. Some other well known candidate genes, such as the *NCAM1-TTC12-ANKK1-DRD2* gene cluster (45,46) and *DDC* (47,48), are not represented in any of the chromosomal regions identified. These findings might reflect the possibility that the effect size of these genes is too small to be detected by the current GSMA.

Recent genomewide association studies (GWAS) have identified many more genes implicated in smoking behavior. The first GWAS on smoking using sample pooling and 2.4 million SNPs suggested several novel genes possibly associated with ND (49). Among the top candidate genes list in this first GWAS, five genes were located in the GSMA nominated bins: *NRXN1* in bin 2.3 (2p22.1–p13.2), *FTO* in bin 16.2 (16p12.3–q12.2), *GPSM3* in bin 20.2 (20p12.1–q13.12), *TRPC7* in bin 5.6 (5q31.2–q34) and *FBXL17* in bin 5.4 (5q14.1–q21.3). Another recent GWAS on smoking behavior (50) was conducted for a sample of 840 European-ancestry subjects using ~380,000 SNPs, and has also identified genes possibly associated with smoking

behavior, among which five genes map within regions discovered by the GSMA: *TBC1D22A* in bin 22.2 (22q12.3–q13.32), *PDE10A* in bin 6.6 (6q25.3–q27), *RDH11* in bin 14.3 (14q23.3–q31.1), *CENTD3* in bin 5.6 (5q31.2–q34) and *LEP* in bin 7.5 (7q31.1–q34).

Several GWAS have consistently identified a region on chromosome 15q24 associated with smoking intensity or lung cancer (51–55). Candidate gene studies have also confirmed the association between variation mapped to the gene cluster (*CHRNA5*, *CHRNA3* and *CHRNA4*) at 15q24 and different smoking behaviors (34,56–59). We are not aware of any individual genomewide linkage scan which has reported genomewide suggestive or significant linkage in the region of 15q24. Our primary 30 cM width GSMA did not detect any signal in 15q24, but we did find $P_{SR}=0.058$ at bin 15.2 (15q21.1–q25.1) which covered the region 15q24 in the 30 cM shifted GSMA of smoking behavior in the European descent populations. The evidence for linkage obtained from the current GSMA thus fails to provide linkage support for the region 15q24 that apparently harbors ND susceptibility genes. The fact that we did not find any stronger evidence for linkage in this region might imply that the genetic effect is small and the current GSMA does not have sufficient power to achieve significant evidence for linkage in this region or that the heterogeneity among different samples obscure the discovery of some linkage loci with minor effects. This is hard to reconcile with the consistency with which this region has been identified in the GWAS.

We discuss briefly additional novel candidate genes mapped to regions discovered by the GSMA. In particular, we focus on the region of chromosome 17q24.3–q25.3 (bin 17.4), since this region ranks highest in the meta-analysis of the combined smoking behavior, and was consistently nominated by the meta-analysis of FTND and MaxCigs24. According to NCBI database information (<http://www.ncbi.nlm.nih.gov/>), there are 253 genes in 17q24.3–q25.3, among which we find two particularly promising candidate genes for smoking. One gene is G protein pathway suppressor 1 (*GPS1*), which suppresses G-protein and mitogen-activated signal transduction. Variants of this gene might influence the regulation of the dopamine signaling pathway and associated with smoking behavior. Another promising candidate gene is suppressor of cytokine signaling 3 (*SOCS3*). A recent GWAS discovered a genomewide significant association of *IL15* with smoking behavior in males (50). As *IL15* is an important cytokine that regulates T and natural killer cell activation and proliferation, the genetic association of *IL15* with smoking may serve as paradigmatic for a novel mechanism for nicotine dependence involving immune modulation through the *IL15* pathway. Hence, it is reasonable to suspect that variants in *SOCS3* gene might influence the regulation of immune system through a cytokine signaling pathway.

One advantage of GSMA is to confirm consistent evidence for linkage across studies. We compared the regions nominated in the current GSMA with regions that were previously identified as consistent across several independent genome scans. Genomic regions on chromosomes 9 (from 91.9 to 136.5 cM based on the Marshfield map), 10 (62–158 cM), 11 (2–76 cM), and 17 (20–82 cM) have been replicated independently more often than other regions (1). However, no strong evidence was achieved for these regions in the current GSMA. GSMA is not used for exclusion mapping and the failure to show strong evidence of linkage for these regions does not necessarily mean that those regions do not harbor risk loci for smoking behavior. The GSMA method is particularly useful to identify regions that show weak but consistent evidence of linkage across multiple studies. It does not take into account directly whether the linkage signals aggregated have reached genomewide, or “suggestive,” evidence for linkage, themselves. Nonetheless, it is notable that some evidence from the current GSMA does support linkage on chromosome 11 and 17. For example, in the primary 30 cM GSMA of smoking behavior, bin 11.1 (0–29.6 cM) showed nominal significance ($P_{SR}=0.017$). Suggestive evidence for linkage has been achieved for bin 17.4 (95–126 cM) which is near the region (20–82 cM) most frequently reported on chromosome 17.

Although GWAS has greater power to detect small effects on phenotype of common variants and copy number variations (CNVs), an adequately powered linkage study design has the advantage of detecting diverse genetic effects that segregate in families, including common variants, multiple rare variants within one locus, and heritable CNVs. With the growing evidence for the role of rare variants and CNVs in psychiatry disorders (60,61), the consensus regions discovered by linkage studies may serve as a useful complement to the emerging GWAS approach in reconstructing the genetic architecture of psychiatry disorders, especially in pinpointing the causal rare variants that cannot be captured by common tag SNPs in the GWAS design. In conclusion, the current meta-analysis including 15 genome scans of smoking behavior has identified many regions showing evidence of linkage with smoking behavior. Known and novel candidate genes map to the highly ranked regions are of particular interest. Therefore, the regions identified in the current study deserve close attention and will be helpful for candidate gene identification or target resequencing studies in the future.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

This work was supported by the U.S. National Institutes of Health (career development award K01 DA024758 to B.Z.Y., and R01 DA12690 and R01 DA12849 to J.G.) and by the US Department of Veterans Affairs (VA CT REAP and New England MIRECC Center). We express our appreciation to the following investigators who contributed their original linkage results to the current meta-analysis: Dr. Laura Jean Bierut; Dr. Kirk C. Wilhelmsen; Dr. Jaakko Kaprio and Dr. Scott F. Saccone. We would like to thank Dr. Cathryn Lewis for helping in understanding the GSMA. We also acknowledge the help of Dr. Paola Forabosco for providing valuable scripts for accomplishing the GSMA. Finally, we thank Ms. Yingdi Xu for digitalizing the linkage results from the published graphs.

References

1. Li MD. Identifying susceptibility loci for nicotine dependence: 2008 update based on recent genome-wide linkage analyses. *Hum Genet* 2008;123:119–131. [PubMed: 18205015]
2. Leshov-Schlaggar CN, Pergadia ML, Khroyan TV, Swan GE. Genetics of nicotine dependence and pharmacotherapy. *Biochem Pharmacol* 2008;75:178–195. [PubMed: 17888884]
3. Fagerstrom KO. Measuring degree of physical dependence to tobacco smoking with reference to individualization of treatment. *Addict Behav* 1978;3:235–241. [PubMed: 735910]
4. Heatherton TF, Kozlowski LT, Frecker RC, Fagerstrom KO. The Fagerstrom Test for Nicotine Dependence: A revision of the Fagerstrom Tolerance Questionnaire. *Br J Addict* 1991;86:1119–1127. [PubMed: 1932883]
5. Straub RE, Sullivan PF, Ma Y, Myakishev MV, Harris-Kerr C, Wormley B, et al. Susceptibility genes for nicotine dependence: a genome scan and follow up in an independent sample suggest that regions on chromosomes 2, 4, 10, 16, 17 and 18 merit further study. *Mol Psychiatry* 1999;4:129–144. [PubMed: 10208445]
6. Goode EL, Badzioch MD, Kim H, Gagnon F, Rozek LS, Edwards KL, et al. Multiple genome-wide analyses of smoking behavior in the Framingham Heart Study. *BMC Genet* 2003;4(Suppl 1):S102. [PubMed: 14975170]
7. Bierut LJ, Rice JP, Goate A, Hinrichs AL, Saccone NL, Foroud T, et al. A genomic scan for habitual smoking in families of alcoholics: common and specific genetic factors in substance dependence. *Am J Med Genet A* 2004;124:19–27. [PubMed: 14679582]
8. Gelernter J, Liu X, Hesselbrock V, Page GP, Goddard A, Zhang H. Results of a genomewide linkage scan: support for chromosomes 9 and 11 loci increasing risk for cigarette smoking. *Am J Med Genet B Neuropsychiatr Genet* 2004;128B:94–101. [PubMed: 15211640]
9. Vink JM, Beem AL, Posthuma D, Neale MC, Willemssen G, Kendler KS, et al. Linkage analysis of smoking initiation and quantity in Dutch sibling pairs. *Pharmacogenomics J* 2004;4:274–282. [PubMed: 15170444]

10. Ehlers CL, Wilhelmsen KC. Genomic screen for loci associated with tobacco usage in Mission Indians. *BMC Med Genet* 2006;10:7–9.
11. Li MD, Payne TJ, Ma JZ, Lou XY, Zhang D, Dupont RT, et al. A genomewide search finds major susceptibility loci for nicotine dependence on chromosome 10 in African Americans. *Am J Hum Genet* 2006;79:745–751. [PubMed: 16960812]
12. Morley KI, Medland SE, Ferreira MA, Lynskey MT, Montgomery GW, Heath AC, et al. A possible smoking susceptibility locus on chromosome 11p12: evidence from sex-limitation linkage analyses in a sample of Australian twin families. *Behav Genet* 2006;36:87–99. [PubMed: 16365831]
13. Swan GE, Hops H, Wilhelmsen KC, Lessov-Schlaggar CN, Cheng LS, Hudmon KS, et al. A genome-wide screen for nicotine dependence susceptibility loci. *Am J Med Genet B Neuropsychiatr Genet* 2006;141B:354–360. [PubMed: 16671072]
14. Gelernter J, Panhuysen C, Weiss R, Brady K, Poling J, Krauthammer M, et al. Genomewide linkage scan for nicotine dependence: identification of a chromosome 5 risk locus. *Biol Psychiatry* 2007;61:119–126. [PubMed: 17081504]
15. Saccone SF, Pergadia ML, Loukola A, Broms U, Montgomery GW, Wang JC, et al. Genetic linkage to chromosome 22q12 for a heavy-smoking quantitative trait in two independent samples. *Am J Hum Genet* 2007;80:856–866. [PubMed: 17436240]
16. Loukola A, Broms U, Maunu H, Widén E, Heikkilä K, Siivola M, et al. Linkage of nicotine dependence and smoking behavior on 10q, 7q and 11p in twins with homogeneous genetic background. *Pharmacogenomics J* 2008;8:209–219. [PubMed: 17549066]
17. Bergen AW, Korczak JF, Weissbecker KA, Goldstein AM. A genome-wide search for loci contributing to smoking and alcoholism. *Genet Epidemiol* 1999;17(Suppl 1):S55–S60. [PubMed: 10597412]
18. Duggirala R, Almasy L, Blangero J. Smoking behavior is under the influence of a major quantitative trait locus on human chromosome 5q. *Genet Epidemiol* 1999;17(Suppl 1):S139–S144. [PubMed: 10597426]
19. Li MD, Ma JZ, Cheng R, Dupont RT, Williams NJ, Crews KM, et al. A genome-wide scan to identify loci for smoking rate in the Framingham Heart Study population. *BMC Genet* 2003;4(Suppl 1):pS103.
20. Saccone NL, Neuman RJ, Saccone SF, Rice JP. Genetic analysis of maximum cigarette-use phenotypes. *BMC Genet* 2003;4(Suppl 1):S105. [PubMed: 14975173]
21. Wang D, Ma JZ, Li MD. Mapping and verification of susceptibility loci for smoking quantity using permutation linkage analysis. *Pharmacogenomics J* 2005;5:166–172. [PubMed: 15724146]
22. Vink JM, Posthuma D, Neale MC, Eline Slagboom P, Boomsma DI. Genome-wide linkage scan to identify Loci for age at first cigarette in Dutch sibling pairs. *Behav Genet* 2006;36:100–111. [PubMed: 16374522]
23. Li MD, Ma JZ, Payne TJ, Lou XY, Zhang D, Dupont RT, et al. Genome-wide linkage scan for nicotine dependence in European Americans and its converging results with African Americans in the Mid-South Tobacco Family sample. *Mol Psychiatry* 2008;13:407–416. [PubMed: 17579606]
24. Pomerleau OF, Pomerleau CS, Chu J, Kardia SL. Genome-wide linkage analysis for smoking-related regions, with replication in two ethnically diverse populations. *Nicotine Tob Res* 2007;9:955–958. [PubMed: 17763112]
25. Wise LH, Lanchbury JS, Lewis CM. Meta-analysis of genome searches. *Ann Hum Genet* 1999;63 (Pt 3):263–272. [PubMed: 10738538]
26. Levinson DF, Levinson MD, Segurado R, Lewis CM. Genome scan meta-analysis of schizophrenia and bipolar disorder, part I: Methods and power analysis. *Am J Hum Genet* 2003;73:17–33. [PubMed: 12802787]
27. Pardi F, Levinson DF, Lewis CM. GSMA: software implementation of the genome search meta-analysis method. *Bioinformatics* 2005;21:4430–4431. [PubMed: 16249265]
28. Forabosco P, Gorman JD, Cleveland C, Kelly JA, Fisher SA, Ortmann WA, et al. Meta-analysis of genome-wide linkage studies of systemic lupus erythematosus. *Genes Immun* 2006;7:609–614. [PubMed: 16971955]

29. Hermanowski J, Bouzigon E, Forabosco P, Ng MY, Fisher SA, Lewis CM. Meta-analysis of genome-wide linkage studies for multiple sclerosis, using an extended GSMA method. *Eur J Hum Genet* 2007;15:703–710. [PubMed: 17377519]
30. Feng Y, Niu T, Xing H, Xu X, Chen C, Peng S, et al. A common haplotype of the nicotine acetylcholine receptor $\alpha 4$ subunit gene is associated with vulnerability to nicotine addiction in men. *Am J Hum Genet* 2004;75:112–121. [PubMed: 15154117]
31. Li MD, Beuten J, Ma JZ, Payne TJ, Lou XY, Garcia V, et al. Ethnic- and gender-specific association of the nicotinic acetylcholine receptor $\alpha 4$ subunit gene (CHRNA4) with nicotine dependence. *Hum Mol Genet* 2005;14:1211–1219. [PubMed: 15790597]
32. Hutchison KE, Allen DL, Filbey FM, Jepson C, Lerman C, Benowitz NL, et al. CHRNA4 and tobacco dependence: from gene regulation to treatment outcome. *Arch Gen Psychiatry* 2007;64:1078–1086. [PubMed: 17768273]
33. Breitling LP, Dahmen N, Mittelstraß K, Rujescu D, Gallinat J, Fehr C, et al. Association of nicotinic acetylcholine receptor subunit $\alpha 4$ polymorphisms with nicotine dependence in 5500 Germans. *Pharmacogenomics J*. 2009 [Epub ahead of print].
34. Saccone NL, Saccone SF, Hinrichs AL, Stitzel JA, Duan W, Pergadia ML, et al. Multiple distinct risk loci for nicotine dependence identified by dense coverage of the complete family of nicotinic receptor subunit (CHRN) genes. *Am J Med Genet B Neuropsychiatr Genet*. 2009 [Epub ahead of print].
35. Tapper AR, McKinney SL, Nashmi R, Schwarz J, Deshpande P, Labarca C, et al. Nicotine activation of $\alpha 4$ receptors: sufficient for reward, tolerance, and sensitization. *Science* 2004;306:1029–1032. [PubMed: 15528443]
36. Yu Y, Kranzler HR, Panhuysen C, Weiss RD, Poling J, Farrer LA, et al. Substance dependence low-density whole genome association study in two distinct American populations. *Hum Genet* 2008;123:495–506. [PubMed: 18438686]
37. Zhang L, Kendler KS, Chen X. The mu-opioid receptor gene and smoking initiation and nicotine dependence. *Behav Brain Funct* 2006;2:28. [PubMed: 16887046]
38. Harlan JE, Hajduk PJ, Yoon HS, Fesik SW. Pleckstrin homology domains bind to phosphatidylinositol-4,5-bisphosphate. *Nature* 1994;371:168–170. [PubMed: 8072546]
39. Schinka JA, Town T, Abdullah L, Crawford FC, Ordorica PI, Francis E, et al. A functional polymorphism within the mu-opioid receptor gene and risk for abuse of alcohol and other substances. *Mol Psychiatry* 2002;7:224–228. [PubMed: 11840318]
40. Luo X, Kranzler HR, Zhao H, Gelernter J. Haplotypes at the OPRM1 locus are associated with susceptibility to substance dependence in European Americans. *Am J Med Genet (Neuropsych Genet)* 2003;120B:97–108.
41. Zhang H, Luo X, Kranzler HR, Lappalainen J, Yang BZ, Krupitsky E, et al. Association between two μ opioid receptor gene (OPRM1) haplotype blocks and drug or alcohol dependence. *Human Molecular Genetics* 2006;15:807–819. [PubMed: 16476706]
42. Shields PG, Lerman C, Audrain J, Bowman ED, Main D, Boyd NR, et al. Dopamine D4 receptors and the risk of cigarette smoking in African-Americans and Caucasians. *Cancer Epidemiol Biomarkers Prev* 1998;7:453–458. [PubMed: 9641486]
43. Vandenbergh DJ, O'Connor RJ, Grant MD, Jefferson AL, Vogler GP, Strasser AA, et al. Dopamine receptor genes (DRD2, DRD3 and DRD4) and gene-gene interactions associated with smoking-related behaviors. *Addict Biol* 2007;12:106–116. [PubMed: 17407504]
44. Beuten J, Payne TJ, Ma JZ, Li MD. Significant association of catechol-O-methyltransferase (COMT) haplotypes with nicotine dependence in male and female smokers of two ethnic populations. *Neuropsychopharmacology* 2006;31:675–684. [PubMed: 16395295]
45. Gelernter J, Yu Y, Weiss R, Brady K, Panhuysen C, Yang BZ, et al. Haplotype spanning TTC12 and ANKK1, flanked by the DRD2 and NCAM1 loci, is strongly associated to nicotine dependence in two distinct American populations. *Hum Mol Genet* 2006;15:3498–3507. [PubMed: 17085484]
46. Yang BZ, Kranzler HR, Zhao H, Gruen JR, Luo X, Gelernter J. Association of Haplotypic Variants in DRD2, ANKK1, TTC12 and NCAM1 to Alcohol Dependence in Independent Case-control and Family Samples. *Hum Mol Genet* 2007;16:2844–2853. [PubMed: 17761687]

47. Ma JZ, Beuten J, Payne TJ, Dupont RT, Elston RC, Li MD. Haplotype analysis indicates an association between the DOPA decarboxylase (DDC) gene and nicotine dependence. *Hum Mol Genet* 2005;14:1691–1698. [PubMed: 15879433]
48. Yu Y, Panhuysen C, Kranzler HR, Hesselbrock V, Rounsaville B, Weiss R, et al. Intronic variants in the dopa decarboxylase (DDC) gene are associated with smoking behavior in European-Americans and African-Americans. *Hum Mol Genet* 2006;15:2192–2199. [PubMed: 16740595]
49. Bierut LJ, Madden PA, Breslau N, Johnson EO, Hatsukami D, Pomerleau OF, et al. Novel genes identified in a high-density genome wide association study for nicotine dependence. *Hum Mol Genet* 2007;16:24–35. [PubMed: 17158188]
50. Liu YZ, Pei YF, Guo YF, Wang L, Liu XG, Yan H, et al. Genome-wide association analyses suggested a novel mechanism for smoking behavior regulated by IL15. *Mol Psychiatry* 2009;14:668–680. [PubMed: 19188921]
51. Thorgeirsson TE, Geller F, Sulem P, Rafnar T, Wiste A, Magnusson KP, et al. A variant associated with nicotine dependence, lung cancer and peripheral arterial disease. *Nature* 2008;452:638–642. [PubMed: 18385739]
52. Berrettini W, Yuan X, Tozzi F, Song K, Francks C, Chilcoat H, et al. Alpha-5/alpha-3 nicotinic receptor subunit alleles increase risk for heavy smoking. *Mol Psychiatry* 2008;13:368–373. [PubMed: 18227835]
53. Amos CI, Wu X, Broderick P, Gorlov IP, Gu J, Eisen T, et al. Genome-wide association scan of tag SNPs identifies a susceptibility locus for lung cancer at 15q25.1. *Nat Genet* 2008;40:616–622. [PubMed: 18385676]
54. Hung RJ, McKay JD, Gaborieau V, Boffetta P, Hashibe M, Zaridze D, et al. A susceptibility locus for lung cancer maps to nicotinic acetylcholine receptor subunit genes on 15q25. *Nature* 2008;452:633–637. [PubMed: 18385738]
55. Caporaso N, Gu F, Chatterjee N, Sheng-Chih J, Yu K, Yeager M, et al. Genome-wide and candidate gene association study of cigarette smoking behaviors. *PLoS ONE* 2009;4(2):e4653. [PubMed: 19247474]
56. Saccone SF, Hinrichs AL, Saccone NL, Chase GA, Konvicka K, Madden PA, et al. Cholinergic nicotinic receptor genes implicated in a nicotine dependence association study targeting 348 candidate genes with 3713 SNPs. *Hum Mol Genet* 2007;16:36–49. [PubMed: 17135278]
57. Weiss RB, Baker TB, Cannon DS, von Niederhausern A, Dunn DM, Matsunami N, et al. A candidate gene approach identifies the CHRNA5-A3-B4 region as a risk factor for age-dependent nicotine addiction. *PLoS Genet* 2008;4(7) e1000125.
58. Stevens VL, Bierut LJ, Talbot JT, Wang JC, Sun J, Hinrichs AL, et al. Nicotinic receptor gene variants influence susceptibility to heavy smoking. *Cancer Epidemiol Biomarkers Prev* 2008;17:3517–3525. [PubMed: 19029397]
59. Freathy RM, Ring SM, Shields B, Galobardes B, Knight B, Weedon MN, et al. A common genetic variant in the 15q24 nicotinic acetylcholine receptor gene cluster (CHRNA5-CHRNA3-CHRNA4) is associated with a reduced ability of women to quit smoking in pregnancy. *Hum Mol Genet*. 2009 May 9; [Epub ahead of print].
60. Stefansson H, Rujescu D, Cichon S, Pietiläinen OP, Ingason A, Steinberg S, et al. Large recurrent microdeletions associated with schizophrenia. *Nature* 2008;455:232–236. [PubMed: 18668039]
61. Walsh T, McClellan JM, McCarthy SE, Addington AM, Pierce SB, Cooper GM, et al. Rare structural variants disrupt multiple genes in neurodevelopmental pathways in schizophrenia. *Science* 2008;320:539–543. [PubMed: 18369103]

Figure 1.

The primary 30 cM width GSMA results for all independent genome scans on smoking behavior (3404 families with 10,253 genotyped subjects). Significance levels corresponding to nominal ($P_{SR} < 0.05$), suggestive ($P_{SR} < 0.0085$) and genome-wide significance ($P_{SR} < 0.00042$) are shown by horizontal lines.

Figure 2.

The primary 30 cM width GSMA results for all independent genome scans on FTND (1347 families with 3995 genotyped subjects). Significance levels corresponding to nominal ($P_{SR} < 0.05$), suggestive ($P_{SR} < 0.0085$) and genome-wide significance ($P_{SR} < 0.00042$) are shown by horizontal lines.

Figure 3.

The primary 30 cM width GSMA results for all independent genome scans on MaxCigs24 (966 families with 3273 genotyped subjects). Significance levels corresponding to nominal ($P_{SR} < 0.05$), suggestive ($P_{SR} < 0.0085$) and genome-wide significance ($P_{SR} < 0.00042$) are shown by horizontal lines.

Table 1

Characteristics of individual genome scans included in the meta-analysis

First author(reference)	Year	Project ¹	Population	No. of			Phenotype ²	Program	Statistics	Included in GSMA
				Families	Individuals	Markers				
Straub (5)	1999	New Zealand	New Zealand (90% Caucasian)	130	343	451	FTQ	Genehunter	Zall	graph
Swan(13)	2006	SMOFAM	American (84% Caucasian)	158	607	739	FTND	Merlin	LOD	data
Li(11)	2006	MSTF	African American	402	1261	385	FTND	Genehunter	LOD	graph
Gelernter(14)	2007	GCOD	African American	320	856	415	FTND	Merlin	LOD	data
Gelernter(14)	2007	GCOD	European American	314	763	415	FTND	Merlin	LOD	data
Loukola(16)	2008	FTF	Finland	153	508	380	FTND	Autogscan	LOD	data
Goode(6)	2003	FHS	American (90% Caucasian)	330	1055	401	MaxCigs24	SOLAR	LOD	graph
Bierut(7)	2004	COGA wave1	American (77% Caucasian)	78	308	336	HS	ASPEX	LOD	data
Bierut(7)	2004	COGA wave2	American (81% Caucasian)	97	346	336	HS	ASPEX	LOD	data
Vink(9)	2004	NTR	Netherlands	192	642	379	MaxCigs24	Mx	LOD	graph
Gelernter(8)	2004	FSPD	European American	12	142	416	HS	Allegro	Zlr	graph
Ehlers(10)	2006	Mission Indians	Native American	41	243	791	PS	SOLAR	LOD	graph
Morley(12)	2006	ATR	Australian	751	1603	1376	CCI	Mx	P-values	graph
Saccone(15)	2007	NAG	Australian (90% Caucasian)	289	953	381	MaxCigs24	Merlin	LOD	data
Saccone(15)	2007	NAG	Finland	155	623	385	MaxCigs24	Merlin	LOD	data

Note:

¹ SMOFAM, Smoking in Families Study; MSTF, Mid-South Tobacco Family; GCOD, Genetics of Cocaine or Opioid Dependence study; FTF, Finnish Twin Families; FHS, Framingham Heart Study; COGA, Collaborative Study on the Genetics of Alcoholism; NTR, Netherlands Twin Register; FSPD, Family Study of Panic Disorder; ATR, Australian Twin Registry; NAG, Nicotine Addiction Genetics project;

² FTQ, Fagerstrom Tolerance Questionnaire; FTND, Fagerstrom test for nicotine dependence; MaxCigs24, Maximum number of cigarettes smoked in a 24-hour period; HS, habitual smoking; PS, persistent smoking; CCI, cigarette consumption including nonsmokers.

Table 2
Bins achieving nominally significant evidence for linkage ($P_{SR} < 0.05$) of smoking behavior

Pos	Bin	Marshfield location (cM)	Physical location (Mb)	Cytogenetic location	Weighted Analysis		Unweighted Analysis		
					SR	P _{SR}	SR	P _{OR}	
All families									
1	17.4	95–126	68–78	17q24.3–q25.3	1228.09	0.0010	1183.5	0.0024	0.25
2	11.1	0–30	0–25	11p15.5–p14.3	1113.62	0.017	1126	0.0098	0.32
3	7.5	121–152	108–142	7q31.1–q34	1105.44	0.019	1105.5	0.015	0.26
4	20.3	67–101	46–57	20q13.12–q13.32	1093.84	0.024	1052	0.042	0.013
5	16.1	0–34	0–18	16p13.3–p12.3	1083.85	0.029	1067.5	0.032	0.063
6	2.3	60–90	39–73	2p22.1–p13.2	1063.68	0.041	1059.5	0.037	0.016
7	3.8	200–228	186–199	3q25.31–q29	1061.97	0.042	1070	0.030	0.13
8	5.4	85–113	78–105	5q14.1–q21.3	1058.34	0.045	1044	0.048	0.012
9	6.5	129–161	132–159	6q23.2–q25.3	1054.68	0.048	1018	0.073	0.053
10	12.2	29–57	12–24	12p13.2–p12.1	1051.01	0.050	975	0.13	0.51
Mostly Caucasian									
1	17.4	95–126	68–78	17q24.3–q25.3	978.68	0.0022	943.5	0.0044	0.41
2	22.2	31–62	31–48	22q12.3–q13.32	922.67	0.010	922.5	0.0078	0.24
3	7.5	121–152	108–142	7q31.1–q34	910.24	0.014	882.5	0.021	0.22
4	2.3	60–90	39–73	2p22.1–p13.2	899.77	0.018	890	0.017	0.34
5	16.1	0–34	0–18	16p13.3–p12.3	865.91	0.036	860.5	0.033	0.18
6	12.2	29–57	12–24	12p13.2–p12.1	855.07	0.044	792	0.11	0.19
7	20.3	67–101	46–57	20q13.12–q13.32	852.85	0.046	818	0.073	0.12
8	6.5	129–161	132–159	6q23.2–q25.3	850.75	0.048	841.5	0.048	0.34
9	5.6	141–170	136–166	5q31.2–q34	847.45	0.051	832.5	0.057	0.20

Note: Bins with nominally significant evidence for linkage ($P_{SR} < 0.05$) are shown for 30 cM bin width weighted GSMA of all (3404 families with 10,253 genotyped subjects) and mostly European ancestry samples (2486 families with 7270 genotyped subjects). The unweighted analysis results were also included as a comparison with weighted analysis. The weighted analysis results were ordered by their position (Pos, 1=best). SR, summed rank; P_{SR} summed rank P -value; P_{OR} ordered rank P -value. $P_{SR} < 0.0085$ and $P_{OR} < 0.00042$ correspond to genome-wide suggestive and significant thresholds for linkage, respectively.

Table 3

Bins achieving nominally significant evidence for linkage ($P_{SR} < 0.05$) of FTND

Pos	Bin	Marshfield location (cM)	Physical location (Mb)	Cytogenetic location	Weighted Analysis		Unweighted Analysis	
					SR	P _{SR}	SR	P _{OR}
All families								
1	6.5	129–161	132–159	6q23.2–q25.3	467.41	0.012	472	0.0093
2	6.6	161–193	159–171	6q25.3–q27	458.62	0.016	451.5	0.020
3	17.4	95–126	68–78	17q24.3–q25.3	452.91	0.020	451	0.021
4	5.4	85–113	78–105	5q14.1–q21.3	444.46	0.027	432.5	0.038
5	16.1	0–34	0–18	16p13.3–p12.3	440.41	0.031	449.5	0.022
6	4.2	30–61	16–47	4p15.32–p12	430.95	0.041	440	0.030
Mostly Caucasian								
1	6.5	129–161	132–159	6q23.2–q25.3	306.91	0.012	303	0.015
2	5.7	170–198	165–180	5q34–q35.3	301.51	0.016	305	0.013
3	4.2	30–61	16–47	4p15.32–p12	299.46	0.018	299	0.019
4	16.1	0–34	0–18	16p13.3–p12.3	290.78	0.028	295.5	0.022
5	17.4	95–126	68–78	17q24.3–q25.3	280.39	0.043	279	0.046
6	5.6	141–170	136–165	5q31.2–q34	280.27	0.043	285	0.036

Note: Bins with nominally significant evidence for linkage ($P_{SR} < 0.05$) are shown for 30 cM bin width weighted GSMA of all (1347 families with 3995 genotyped subjects) and mostly European ancestry samples (625 families with 1878 genotyped subjects). The unweighted analysis results were also included as a comparison with weighted analysis. $P_{SR} < 0.0085$ and $P_{SR} < 0.00042$ correspond to genome-wide suggestive and significant thresholds for linkage, respectively. Other legend reference Table 2

Table 4

Bins achieving nominally significant evidence for linkage ($P_{SR} < 0.05$) of MaxCigs24

Pos	Bin	Marshfield location (cM)	Physical location (Mb)	Cytogenetic location	Weighted Analysis		Unweighted Analysis	
					SR	P_{SR}	SR	P_{SR}
1	20.3	67–101	46–57	20q13.12–q13.32	436.32	0.00041	438.5	0.00032
2	22.2	31–62	31–48	22q12.3–q13.32	423.12	0.0015	423	0.0015
3	20.2	34–67	13–46	20p12.1–q13.12	403.69	0.0054	399.5	0.0066
4	17.4	95–126	68–78	17q24.3–q25.3	398.83	0.0070	396	0.0080
5	2.8	209–239	213–233	2q34–q37.1	384.89	0.014	382	0.015
6	12.2	29–57	12–24	12p13.2–p12.1	380.59	0.017	374	0.021
7	22.1	0–31	0–31	22pter–q12.3	374.19	0.022	384	0.014
8	2.4	90–120	72–108	2p13.3–q12.3	357.92	0.039	357	0.039
9	2.3	60–90	39–73	2p22.1–p13.2	357.32	0.041	360	0.036
10	16.2	34–67	18–51	16p12.3–q12.2	352.63	0.047	351	0.049
11	2.5	120–150	108–146	2q12.3–q22.3	351.43	0.049	355.5	0.042

Note: Bins with nominally significant evidence for linkage ($P_{SR} < 0.05$) are shown for 30 cM bin width weighted GSMA (966 families with 3273 genotyped subjects). The unweighted analysis results were also included as a comparison with weighted analysis. $P_{SR} < 0.0085$ and $P_{SR} < 0.00042$ correspond to genome-wide suggestive and significant thresholds for linkage, respectively. Other legend reference Table 2