Published in final edited form as:

J Cardiovasc Magn Reson. 2007 ; 9(1): 71–76. doi:10.1080/10976640600843330.

Reproducibility and Reliability of Atherosclerotic Plaque Volume Measurements in Peripheral Arterial Disease with Cardiovascular Magnetic Resonance

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Abstract

A high resolution, noninvasive approach to quantify atherosclerotic plaque in the peripheral vasculature could have significant clinical and research utility. Seventeen patients with peripheral arterial disease (PAD) were studied in a 1.5T CMR scanner. Atherosclerotic plaque volume in the superficial femoral artery was measured and interobserver, intraobserver, and test-retest variability determined. Nineteen vessels were studied with mean acquisition time of 13.1 minutes per vessel. Mean plaque volume was $7.27 \pm 3.73 \text{ cm}^3$. Intra-observer intraclass correlation was $R = 0.997$, inter-observer was $R = 0.987$, and test-retest reproducibility was $R = 0.996$. Thus, high resolution measurement of plaque volume in PAD is reliable and reproducible.

Keywords

Atherosclerosis; Cardiovascular Magnetic Resonance; Peripheral Arterial Disease; Peripheral Vascular Disease; Superficial Femoral Artery

INTRODUCTION

Peripheral arterial disease (PAD) is an increasingly common condition characterized by atherosclerotic obstruction of the arteries supplying the lower limbs. Although PAD is now recognized as an important manifestation of systemic atherosclerosis, it remains underdiagnosed and undertreated (1). While co-existing coronary and cerebrovascular disease is responsible for nearly 75% of deaths among PAD patients (2–4), morbidity directly attributable to peripheral vascular obstruction is substantial (5). Of the few therapies

proven beneficial in the treatment of PAD, little is understood about their mechanisms of benefit (6–8).

The ankle-brachial index is an excellent test for identifying peripheral arterial obstruction (1) but limited in its utility for assessing disease severity and predicting clinical events (9). Direct measurement of plaque and peripheral arterial remodeling may be better suited for defining disease burden and predicting clinical progression. Because of the inherent limitations of lumenographic techniques in evaluating and assessing the extent of atheroma (10,11), novel approaches for directly evaluating the vessel wall are needed. Intravascular ultrasound (IVUS) is currently the diagnostic standard for plaque quantification, but the technique is expensive, invasive, and poorly suited for the peripheral circulation (12).

With its safety record, flexibility, and excellent spatial resolution, cardiovascular magnetic resonance (CMR) could be an ideal approach for the noninvasive assessment of atherosclerotic plaque burden in the peripheral circulation. While CMR has been employed for measuring and characterizing plaque in the coronary (13,14), carotid (15,16), and aortic circulations (15–17), few studies have rigorously investigated its role in PAD (12,18). The goal of this study was to develop and refine a practical technique for quantifying plaque in the superficial femoral artery (SFA) of patients with mild to moderate PAD that would allow for determination of plaque burden and noninvasive, serial monitoring of plaque progression in patients treated with both established and novel therapies.

METHODS

Study population

Patients between the ages of 30–85 years with symptoms of intermittent claudication without critical limb ischemia and an ankle-brachial index (ABI) between 0.4 and 0.9 were eligible for this study. The study was carried out in accordance with a protocol that was approved by the Human Investigation Committee at the University of Virginia Health System and all participants signed informed consent.

Protocol

All subjects were placed supine in a 1.5T MR scanner (Siemens Medical Solutions, Erlangen, Germany) with the calf at the isocenter of the magnet. Monitoring of the electrocardiogram and blood pressure was performed with an InVivo 3155MVS (Intermetrics Companies, Orlando, FL) throughout the study. A custom-built flexible, linear four-element (10 cm × 10 cm square element) surface coil array (Nova Medical, Wilmington, MA) was placed over the thigh and SFA. A multi-slice turbo-spin-echo pulse sequence with fat presaturation was used. Flowing blood was suppressed using spatial presaturation, with a combination of periodic excitation of upstream slices and spatial presaturation. Additional flow suppression for downstream slices was provided by the imaging excitations themselves, so that blood was suppressed throughout the multislice data set. Other imaging parameters included: repetition time 11 ms, echo time 7.6 ms, echo spacing 7.5 ms, turbo factor (9), voxel size 0.5 × 0.5 × 3 mm, 4 signal averages, with interleaved image sets used to cover the length of the SFA. Imaging began above the femoral bifurcation and continued through the adductor canal. Representative contiguous images of the femoral artery from a patient with PAD are shown (Fig. 1).

Image analysis

Before quantification of plaque volume, image quality was assessed and deemed adequate or inadequate for analysis. A total of 23 patients were imaged, and 17 had image quality suitable for analysis. In (4) of the (6) excluded data sets, complete occlusion of the SFA

precluded analysis. Occlusion of flow was verified with velocity encoded imaging. In 1 subject, obesity (BMI = 33) and a metal artifact precluded analysis, and another subject's obesity (BMI > 42) led to poor image quality.

In those subjects with a patent SFA and adequate image quality, atherosclerotic plaque volume (APV) of the SFA was contoured by two independent operators using VesseIMASS software (University of Leiden, Leiden, The Netherlands). For each individual slice, both the luminal and adventitial borders were manually delineated and cross-sectional area (CSA) of the vessel wall measured. Total vessel wall volume was calculated as:

$$\text{Volume} = \sum_{i=1}^n \text{CSA}_i \times H,$$

where H is the slice thickness and n the number of slices in the 3-D data set. Volume measurements were initiated at the bifurcation of the common femoral artery, and total vessel distance covered for each analysis was predefined. Nine patients returned (at a mean of 24 days) for evaluation of test-retest reliability using anatomic landmarks and matched number of acquired slices. Imaging time was measured from acquisition of the first scouts to completion of the imaging protocol but does not include time for patient positioning, coil placement, and insertion and removal from the scanner.

Statistical analysis

Subject characteristics are summarized as mean and standard deviation. The intraclass correlation coefficients of reliability were calculated for the interobserver, intraobserver, and reproducibility data using the output of analysis of variance from SAS statistical software (version 9.0, Cary, NC). Reproducibility and both inter and intraobserver variability were also analyzed using the method of Bland and Altman (19). MedCalc software version 8.1.0.0, Mariakerke, Belgium) was used to generate Bland-Altman plots.

Using the test-retest reliability data, we estimated the sample sizes needed to detect the true difference between the means of APV with both 80% and 90% power. Assuming a two-tailed test, a significance level of 0.05, and using the estimated standard deviation, the sample sizes needed to detect the true mean differences ranging from 1% to 5% of the mean APV were established.

RESULTS

Seventeen patients (age 63 ± 10 yrs) with mild to moderate PAD (ankle brachial index 0.72 ± 0.11) were studied. Characteristics of patients are shown in Table 1. Nineteen SFA vessels in the 17 patients were evaluated (mean length 17.1 ± 5.2 cm) with total image acquisition time of 13.1 ± 2.7 minutes per vessel. Mean APV measured was 7.27 ± 3.73 cm³.

For the inter-observer data, intraclass correlation coefficient was $R = 0.987$ (95% CI 0.967–0.995). Bland-Altman plot for inter-observer variability is shown in Fig. 2. The percent difference between operator measurements was $5.2 \pm 3.3\%$ of the total plaque volume measured per patient. A single operator performed repeat analysis of the 19 vessel segments at a mean of 24 days later. For intra-observer data, intraclass correlation coefficient was $R = 0.997$ (95% CI 0.993 to 0.998). Bland-Altman plot for intra-observer data is shown in Fig. 3. For reproducibility data, intraclass correlation coefficient was $R = 0.996$ (95% CI 0.991 to 0.999). Bland-Altman analysis of test-retest reliability ($n = 9$) is shown in Fig. 4.

Sample size estimates for a study evaluating APV progression/regression were made using a paired t-test and standard deviation = 0.267 obtained from the test-retest reliability data (Table 2). These estimates assume all patients enrolled will have adequate image quality for analysis.

DISCUSSION

The present study demonstrates that high resolution, high volume MR measurement of atherosclerotic plaque in the superficial femoral artery in peripheral arterial disease is feasible and highly reproducible and reliable. Potential applications include diagnosing preclinical vascular disease, assessing disease severity in those with established PAD, and monitoring atherosclerotic plaque progression in response to both established and novel therapies. Furthermore, this noninvasive approach could reduce required sample sizes for clinical studies using atherosclerotic plaque progression rate as a surrogate efficacy endpoint.

Contrast x-ray angiography remains the most commonly used clinical tool for diagnosing atherosclerosis and assessing its severity. The technique has also been used extensively in studies evaluating plaque progression rate (20–22) a dynamic parameter now established as an important surrogate marker of clinical risk (23). However, angiography is associated with the risk of procedural complications and is hampered by its inability to visualize the vessel wall, a pitfall that can lead to systematic underestimation of plaque burden (11). It is now recognized that atherosclerotic plaque in coronary arteries may not encroach the lumen until the lesion occupies up to 40% of the combined arterial wall and lumen volume, a phenomenon commonly referred to as positive remodeling (10). Positive remodeling also appears to play an important role in disease progression in the peripheral vasculature (24), suggesting that a technique capable of visualizing the vessel wall will be most accurate for the assessment of atherosclerotic disease burden in the periphery.

In recent years, IVUS has emerged as the diagnostic gold standard for imaging vessel wall pathology. IVUS is a catheter based imaging technique that provides cross-sectional images of both the vessel lumen and wall with excellent spatial resolution, but it is invasive and can be associated with vessel spasm, dissection, and guide wire entrapment (25,26). Although increasingly employed to evaluate plaque burden and vascular remodeling as a surrogate efficacy endpoint in randomized clinical studies (23,27,28), IVUS is not an ideal strategy for making serial measurements in humans, prompting investigators to seek out novel noninvasive approaches for quantifying atheromatous plaque.

There are a number of advantages to using CMR in the study of human peripheral atherosclerosis. In addition to its excellent safety profile, recent technological advances in sequence design, imaging speed, and detection coils have made possible rapid, high-resolution imaging of atherosclerosis in-vivo (29). Furthermore, its sensitivity to different chemical components of tissue can be exploited to noninvasively characterize plaque composition (30–32). Enthusiasm for the MR approach has already led to several studies evaluating atherosclerotic plaque progression in humans (15–17), but few studies have evaluated the utility of CMR in measuring plaque in the peripheral circulation among PAD. In the most rigorously performed study to date, 7 patients with peripheral arterial occlusive disease were examined by both high-resolution MR imaging and IVUS (12). The MR approach consisted of a three-dimensional time-of-flight sequence used to measure vessel wall parameters with an in-plane resolution of $0.78 \times 0.49 \text{ mm}^2$ and slice thickness of 2 mm. High-resolution MR using this bright blood technique consistently overestimated plaque area in the peripheral vessels compared to IVUS measurements. Moreover, posterior echo

shadowing precluded reliable IVUS analysis in nearly half of the segments assessed, also highlighting the limitations of IVUS in PAD.

We sought to develop an approach to high resolution assessment of plaque volume in the peripheral circulation of patients with mild to moderate PAD. To facilitate this, a flexible, linear four-element surface array coil was designed and built specifically for this protocol. In order to clearly delineate both the luminal and adventitial border of the vessel wall, a black blood technique with fat saturation was chosen. Rather than using double inversion recovery sequences to generate black blood, flow saturation bands were applied in order to reduce acquisition times. With this protocol, nearly 17 cm of the SFA was imaged in just over 13 minutes with excellent in-plane spatial resolution.

There are several potential advantages of imaging plaque volume in the SFA. The lack of SFA movement can significantly limit artifacts compared to imaging in other vascular territories such as the carotids, coronaries, and aorta which can be limited by swallowing or respiratory motion. Also, the relative proximity of the arteries in the leg to the body surface allows the utilization of high-resolution surface coils like the one custom-built for this protocol. Perhaps the biggest advantage of imaging plaque in the SFA is the sheer volume of atheroma that can be measured, $7.27 \pm 3.73 \text{ cm}^3$ in this study, several orders of magnitude greater than volumes reported in either the carotid or coronary circulations. The large volume of plaque that can be quantified and excellent reproducibility of the measurements would reduce sample sizes needed to adequately power clinical studies using atherosclerotic plaque progression as a surrogate endpoint (Table 2).

When estimating the potential impact of this technique on sample size estimates for clinical studies, one must consider the other noninvasive strategies currently being employed. Using carotid intimal medial thickness measurements as a surrogate efficacy end-point, Bots et al. reported that 468 patients would be required to detect a treatment effect of 30% (power 0.80) (33). Three-dimensional ultrasound of plaque in the carotids would require fewer patients to demonstrate efficacy, yet a recent study reported that 203 patients would still be needed to detect a 10% change in plaque volume (power = 0.90) measured over a three month period (34). Measuring the progression of coronary calcium with cardiac computed tomography has been proposed as yet another noninvasive alternative to evaluating disease progression, yet sample sizes required for these studies remain large, and there is the strong suggestion that calcification represents a fundamentally distinct biological process which may correlate less with clinical events than plaque volume progression (35–37). Sample size estimates for our noninvasive approach are similar to that reported using coronary IVUS (27).

Limitations

Using the techniques described, the ability to accurately define the luminal border of the vessel was predicated on the generation of black blood. Whether using inversion recovery or flow saturation bands, as done in this study, black blood contrast requires adequate inflow from outside of the selected slice. Even in this population of patients with mild to moderate disease, 17% had near-complete or complete occlusion of the SFA making images impossible to analyze. The reliability and reproducibility of this imaging strategy may suffer further when performed in patients with more advanced vascular disease and significantly reduced flow with potentially higher rates of SFA occlusion. While techniques that do not rely on the generation of black blood (steady-state free precession) to define the lumen may be adequate in patients with advanced disease and very slow blood flow, these techniques will still be insufficient in the setting of complete occlusions.

Because we utilized a small-element surface coil, vessel image quality suffered in more obese patients. However, only one patient in this study could not be analyzed as a consequence of obesity, and two individuals with BMI > 50 were included in the analysis. A larger coil array and longer scan times with more signal averages would improve image quality in larger patients. We also found that image quality diminished in the distal 1/4 of the SFA in some patients, typically as the vessel coursed deeper into the tissue and posteriorly into the popliteal fossa. This could be problematic as the adductor canal region of the SFA has a known predilection for atherosclerosis and is a common site of disease progression (38,39). In the coronary vasculature, even modest changes in plaque progression may be associated with large differences in vascular outcomes (20,21,23). Whether this relationship between plaque progression and vascular events holds true in the periphery remains undetermined.

Sample size estimates are based on plaque volumes measured in patients with established PAD. In clinical studies evaluating therapeutic efficacy among patients with subclinical atherosclerosis, larger total sample sizes may be required using our approach because of smaller total plaque burden in this population. While required sample sizes may be reduced with measurement of plaque volume progression in the SFA, total imaging costs may be similar to that of other techniques because of the increased expense of CMR. However, these imaging costs would be small compared to the potential savings realized by having a substantially smaller patient cohort to enroll, image, analyze, and follow.

Future directions

We have demonstrated that accurate noninvasive quantification of peripheral plaque burden is feasible in PAD. This approach could be used to better understand the natural history of PAD and characterize plaque volume and distribution in different PAD patient subsets, including those who are asymptomatic, have atypical symptoms, or are diabetic. We also hope to identify plaque characteristics that can be used to recognize PAD at highest risk for adverse clinical outcomes in the lower extremities. Furthermore, serial noninvasive measurement of plaque volume with CMR could allow for the study and validation of both established and novel therapies for PAD, including behavioral, pharmacologic, genetic, stem cell, and percutaneous interventions. Correlation of plaque progression in the peripheral circulation with vascular events in the coronary, cerebral, and peripheral circulation is also warranted.

Acknowledgments

Supported by NIH RO1 HL075792 (CMK) and AHA 0425674U (DCI).

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Figure 1. Representative sequential images (upper left to lower right) from the femoral artery of a subject with mild to moderate peripheral arterial disease with both the luminal and adventitial border clearly delineated. Note the slice to slice variation in plaque morphology.

Figure 2. Bland-Altman plot of inter-observer variability. Mean difference between different operator measurements is 0.19cm³.

Figure 3.
Bland-Altman plot of intra-observer variability. Mean difference between same operator measurements on single data-set is 0.01 cm³.

Figure 4. Bland-Altman plot of test-retest reproducibility. Mean difference between test-retest is 0.11 cm³.

Table 1

Patient characteristics

ID	ABI	CAD	Stroke	DM	Tobacco	SBP (mmHg)	BMI
1	0.66	Yes	No	Yes	Yes	125	22.7
2	0.8	Yes	No	No	Yes	153	23.5
3	0.77	Yes	No	Yes	Yes	130	24.2
4	0.48	No	No	No	Yes	195	N/A
5	0.81	Yes	No	Yes	Yes	103	27.1
6	0.85	No	No	No	Yes	180	21.1
7	0.6	Yes	No	No	Yes	165	21
8	0.73	Yes	Yes	Yes	Yes	150	39.8
9	0.6	Yes	No	Yes	Yes	140	53.2
10	0.8	Yes	No	Yes	Yes	140	28.3
11	0.69	Yes	Yes	No	Yes	130	25.5
12	0.58	Yes	No	Yes	Yes	136	50.2
13	0.83	No	No	No	Yes	132	24
14	0.65	No	No	Yes	Yes	136	21.7
15	0.85	No	No	No	Yes	132	24.9
16	0.55	No	No	No	Yes	155	26.4
17	0.80	No	No	No	Yes	133	26.3
Mean	0.72					143	28.7
SD	0.11					22	10.0

N/A indicates data not available

Table 2

Sample size estimates

Mean Δ APV	N (power = 0.80)	N (power = 0.90)
1%	107	142
2%	27	36
3%	12	16
5%	5	6