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Design of HIV-1 Protease Inhibitors with Pyrrolidinones and Oxazolidinones as Novel P1'-Ligands to Enhance Backbone-binding interactions with Protease: Synthesis, Biological Evaluation and Protein-ligand X-ray Studies

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Abstract

Structure-based design, synthesis and biological evaluation of a series of novel HIV-1 protease inhibitors are described. In an effort to enhance interactions with protease backbone atoms, we have incorporated stereochemically defined methyl-2-pyrrolidinone and methyl oxazolidinone as the P1'-ligands. These ligands are designed to interact with Gly-27' carbonyl and Arg-8 side chain in the S1'-subsite of the HIV protease. We have investigated the potential of these ligands in combination with our previously developed bis-tetrahydrofuran (bis-THF) and cyclopentanyltetrahydrofuran (Cp-THF) as the P2-ligands. Inhibitor **19b** with an (*S*)-aminomethyl-2-pyrrolidinone and a Cp-THF was shown to be the most potent compound. Inhibitor **19b** maintained near full potency against multi-PI-resistant clinical HIV-1 variants. A high resolution protein-ligand X-ray crystal structure of **19b**-bound HIV-1 protease revealed that the P1'-pyrrolidinone heterocycle and the P2-Cp-ligand are involved in several critical interactions with the backbone atoms in the S1' and S2-subsites of HIV-1 protease.

Introduction

Advances in the treatment of HIV/AIDS with HIV-1 protease inhibitors in combination with reverse transcriptase inhibitors have been widely documented. The combination therapy, also known as highly active antiretroviral therapy (HAART), blocks critical viral replication at two different stages of the replication cycle.² The HAART regimens have resulted in dramatic reduction of blood plasma viral load levels, increased CD₄⁺ lymphocyte counts, improved life

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The PDB accession code for **19b**-bound HIV-1 protease X-ray structure is 3H5B.

Supporting Information Available. HPLC and HRMS data of inhibitors. This material is available free of charge via the Internet at <http://pubs.acs.org>.

expectancy and significantly reduced HIV/AIDS-related mortality in the developed world.³ Despite these important advances, effective long-term suppression of HIV infection with HAART regimens is a complex issue in medicine for a number of reasons. These include drug side effects, poor penetration into protected HIV reservoir sites, poor oral bioavailability and interactions between drugs.⁴ Perhaps one of the most daunting problems in future management of HIV is the emergence of drug-resistant HIV-1 variants and the transmission of these viral strains.^{5,6} Thus, development of antiretroviral therapy with broad-spectrum activity and minimal drug side effects is critical for an effective management of current and future HIV/AIDS treatment. We recently reported the design and development of a number of exceedingly potent nonpeptidic HIV-1 protease inhibitors (PIs) ^{1–3,7–9} One of those PIs is darunavir (¹, TMC-114), which was approved by the FDA in 2006 for treatment of HIV/AIDS patients who are harboring drug-resistant HIV and do not respond to other therapies.¹⁰ More recently, darunavir has received full approval for all HIV/AIDS patients.¹¹

To combat drug resistance, our structure-based design strategies are to maximize the protease active-site interactions with the inhibitor and particularly to promote extensive hydrogen bonding with the protein backbone atoms.¹² It is evident that active site backbone conformation of mutant proteases is only minimally distorted compared to that of the wild-type HIV-1 protease.^{13,14} Therefore, the ‘backbone binding’ strategy may be important to combat drug resistance.¹² Using high resolution protein-ligand X-ray structures of ¹ and ³-bound HIV-1 protease, we have shown that these PIs were engaged in extensive hydrogen bonding interactions with the backbone atoms throughout the active site cavity from the S2 to S2’ regions.^{9,15} To further enhance ‘backbone binding’ interactions, we became interested in designing an appropriately functionalized P1’-ligand that could interact with the backbone atoms, particularly with the Gly-27’ and Arg-8 in the S1’-subsite. This enhancement of ‘backbone binding’ interaction may lead to inhibitors with improved drug-resistance profiles. Herein, we report the design, synthesis and biological evaluation of a series of potent HIV-1 protease inhibitors that incorporated structure-based designed stereochemically defined lactam and oxazolidinone derivatives as the P1’-ligands in combination with the bis-THF or Cp-THF as the P2-ligands. Inhibitor ⁴ incorporating a (*S*)-5-aminomethyl-2-pyrrolidinone as the P1’-ligand and Cp-THF as the P2-ligand is the most potent PI in the series. Interestingly, this PI has retained full potency against a range of multi-drug-resistant HIV-1 variants. The protein-ligand X-ray structure of ⁴-bound HIV-1 protease revealed important molecular insight into the ligand-binding site interactions.

Chemistry

The optically active synthesis of the requisite 5-aminomethyl-2-pyrrolidinone for P1-ligands and their conversion to respective sulfonamide isostere is shown in Scheme 1. Commercially available 5-(*S*)-hydroxymethyl-2-pyrrolidinone ⁵ was reacted with tosyl chloride and triethylamine to provide the corresponding tosylate. Displacement of the resulting tosylate with sodium azide in DMF at 55 °C for 9 h provided the azide derivative in 92% yield over two steps. Catalytic hydrogenation of the azide over 10% Pd-C in ethyl acetate afforded optically active amine ⁶ in quantitative yield. 5-(*R*)-hydroxymethyl-2-pyrrolidinone (ent-⁵) was similarly converted to optically active amine ent-⁶ in comparable yield. Amine ⁶ was reacted with commercially available epoxide ⁷ in the presence of *i*Pr₂NEt (DIPEA) in 2-propanol at 70 °C for 36 h to provide epoxide opened product ⁸ in 85% yield.¹⁶ Amine ⁸ was converted to *p*-methoxybenzenesulfonamide derivative ⁹ by reaction with *p*-methoxybenzenesulfonyl chloride in the presence of aqueous NaHCO₃ in quantitative yield. Treatment of amine ⁸ with *p*-nitrobenzenesulfonyl chloride afforded the corresponding nitrosulfonamide. Catalytic hydrogenation over 10% Pd-C gave the corresponding aniline derivative, which was reacted with benzyl chloroformate in the presence of pyridine to furnish Cbz-derivative ¹⁰ in 63% yield

for 3 steps. Enantiomeric amine (*ent*-⁶) was converted to the respective methoxy and Cbz-derived ¹¹ and ¹² by analogous procedures.

The synthesis of various PIs incorporating methyl-pyrrolidinones as the P1'-ligand is shown in Scheme 2. Exposure of Boc-derivatives ⁹ and ¹⁰ to 30% CF₃CO₂H in CH₂Cl₂ at 23 °C for 40 min resulted in the respective amines ¹³ and ¹⁴. Alkoxycarbonylation of amine ¹³ with activated mixed carbonate ¹⁵¹⁶ and ¹⁶⁹ in the presence of Et₃N in CH₂Cl₂ furnished inhibitors **17a** and **17b** in 98% and 87% yields, respectively.¹⁷ Alkoxycarbonylation of amine ¹⁴ with activated carbonates ¹⁵ and ¹⁶ afforded the corresponding Cbz-protected urethanes of **18a** and **18b**. Removal of the Cbz-group by catalytic hydrogenation over 10% Pd-C in ethyl acetate provided inhibitor **18a** and **18b** in 58% and 62% yields, respectively. Sulfonamide derivative ¹¹ and ¹² containing enantiomeric P1'-ligands were converted to inhibitors **19a,b** and **20a,b** by following analogous procedures.

The synthesis of sulfonamide isosteres incorporating methyl oxazolidinone as the P1'-ligand is shown in Scheme 3. Optically active dimethyloxazolidines ²¹ and *ent*-²¹ were prepared by following the procedure described by Dondini and co-workers.¹⁸ Reduction of these azides by catalytic hydrogenation in methanol afforded the respective amine. Reaction of ²¹-derived amine with epoxide ⁷ in the presence of *i*Pr₂NEt in 2-propanol afforded amine ²³ in 41% yield. Reaction of amine ²² with *p*-methoxybenzenesulfonyl chloride or *p*-nitrobenzenesulfonyl chloride as described previously afforded sulfonamide derivatives ²³ and ²⁴ in 80% and 92% yields, respectively. The isopropylidene functionality in ²³ and ²⁴ was converted to the corresponding oxazolidinone derivative in a three steps sequence involving: (1) treatment of ²³ by a catalytic amount of *p*-toluenesulfonic acid (PTSA) in methanol resulted in the removal of the isopropylidene group; (2) reaction of the resulting Boc-amino alcohol with mesyl chloride in the presence of triethylamine to provide the corresponding mesylate and (3) treatment of the resulting mesylate with *i*Pr₂NEt in chloroform at reflux. This has provided oxazolidinone ²⁵ in 45% yield over 3 steps. The nitrosulfonamide derivative ²⁴ was similarly converted to the corresponding oxazolidinone. Catalytic hydrogenation of the resulting nitro derivative with 10% Pd-C in methanol provided aniline derivative ²⁶ in 37% overall yield over 4 steps. Enantiomeric azide *ent*-²¹ was converted to oxazolidinone derivatives ²⁷ and ²⁸ by following analogous procedures.

The synthesis of inhibitors containing oxazolidinone as P1'-ligand and bis-THF as the P2-ligand is shown in Scheme 4. Treatment of oxazolidinones ^{25–28} with 30% CF₃CO₂H in CH₂Cl₂ at 23 °C afforded the corresponding amines. Reaction of the resulting amines with activated mixed carbonate ¹⁵ in the presence of Et₃N in CH₂Cl₂ afforded the target inhibitor ^{29–32} in excellent yields (80–90%). The structures of these inhibitors are shown in Table 1.

Results and Discussion

Our examination of the X-ray structure of ¹-bound HIV-1 protease and its respective modeling initially suggested that a methyl-2-pyrrolidinone may interact well with residues in the S1'-site.¹⁵ As shown in Table 1, our first set of inhibitors contain a (*R*)-hydroxyethylamine sulfonamide isostere with either the bis-THF or Cp-THF as the P2-ligand and *p*-methoxysulfonamide or *p*-aminosulfonamide as the P2'-ligand. The enzyme inhibitory potency of these PIs was evaluated according to the procedure reported by Toth and Marshall.¹⁹ Inhibitor **17a** with (*S*)-methyl-2-pyrrolidinone displayed an enzyme *K_i* of 1 nM. Inhibitor **17b** with a Cp-THF showed a 3-fold improvement of potency. Antiviral activity of these inhibitors was determined in MT-2 human T-lymphoid cells exposed to HIV-1_{Iai}. Interestingly, both inhibitors have shown dramatic reduction in antiviral activity. Inhibitors **17a** and **17b** have shown IC₅₀ values of 0.48 μM, and 0.23 μM, respectively. However, these inhibitors are significantly less potent compared to inhibitors with an isobutyl group as the P1'-ligand.

Incorporation of *p*-aminosulfonamide (PIs **18a** and **18b**) as the P2'-ligand led to a drop in enzyme inhibitory as well as antiviral potency. Inhibitor **19a** containing (*R*)-methyl-2-pyrrolidinone as the P1'-ligand has shown 10-fold enhancement of enzyme K_i over the (*S*)-isomer **17a**. It showed a slight improvement in antiviral activity compared to inhibitor **17a**. Inhibitor **19b** with (*R*)-methyl-2-pyrrolidinone as the P1'-ligand and Cp-THF as the P2'-ligand resulted in the most potent inhibitor in the series. It has shown an enzymatic K_i of 99 pM and a 10-fold improvement ($IC_{50} = 0.026 \mu M$) in antiviral activity relative to epimeric (*S*)-pyrrolidinone derivative **17b**, suggesting an important role for the P1'-ring stereochemistry. Indeed, an X-ray structure of **19b**-bound HIV-1 protease revealed that the pyrrolidinone carbonyl as well as the NH functionalities were positioned to hydrogen bond with residues in the S1'-site. Interestingly, the combination of P1'-methylpyrrolidinone and polar P2'-*p*-aminosulfonamide led to PIs, with subnanomolar enzyme activity. However, antiviral activity was reduced drastically. In PIs ^{29–32}, we have incorporated both (*S*)- and (*R*)-oxazolidinone derivatives as substitutes for the respective pyrrolidinone isomers. As can be seen, oxazolidinone derivatives ^{29–32} have shown subnanomolar enzyme inhibitory potency. Inhibitors with *p*-methoxysulfonamide as the P2'-ligand displayed comparable antiviral activity relative to pyrrolidinone derivatives. Consistent with stereochemical preference, the (*R*)-oxazolidinone with *p*-methoxysulfonamide has shown better enzyme K_i values. However, the antiviral activity of these compounds is very similar. In general, both pyrrolidinone and oxazolidinone functionalities appear to be nicely accommodated in the S1'-site.

While inhibitor ³¹ is very potent in enzyme inhibitory assay, the significant reduction of antiviral potency is possibly due to poor cellular permeability of this polar functionality. Inhibitor **19b** appeared to be most potent among the series of inhibitors examined. It exhibited comparable antiviral activity with the FDA approved PIs amprenavir, saquinavir, and indinavir in the same assay.

Inhibitor **19b** was subsequently examined for its activity against a clinical wild-type X₄-HIV-1 isolate (HIV-1_{ERS104pre}) along with various multidrug-resistant clinical X₄- and R₅- HIV-1 isolates using PBMCs as target cells.^{8b} As can be seen in Table 2, the potency of **19b** against HIV-1_{ERio4pre} ($IC_{50} = 28 \text{ nM}$) was comparable to FDA approved PIs, indinavir, amprenavir and lopinavir with IC_{50} values 28, 25, and 30 nM, respectively. Darunavir on the other hand, is nearly 10-fold more potent ($IC_{50} = 3.6 \text{ nM}$) than **19b** and the above mentioned PIs. Interestingly, of all the PIs tested, indinavir was least able to suppress the replication of the multi-drug-resistant clinical isolate examined (HIV-1_{mdr/mm}, HIV-1_{mdr/tm}, HIV-1_{mdr/c} and HIV-1_{mdr/g}) with IC_{50} values greater than 1 μM . Both amprenavir and lopinavir displayed 10-fold or greater reduction in potency except against HIV-1MDR/G, where lopinavir showed a 5-fold reduction in potency. A more detailed virologic study using inhibitor **19b** will be published elsewhere.²⁰ Darunavir has maintained impressive activity against all the multi-drug-resistant variant. Inhibitor **19b**, while less potent than darunavir, maintained near full potency against multi-drug-resistant clinical isolates examined. This impressive drug-resistance property of **19b** is possibly due to its extensive interactions, particularly its ability to make extensive hydrogen-bonds throughout the active site of the protease's backbone. Furthermore, inhibitor **19b** blocked the infection and replication of each of the HIV-1_{nl4-3} variants exposed to and selected by up to 5 μM of saquinavir, amprenavir, indinavir, nelfinavir, or ritonavir and a 1 μM of lopinavir or atazanavir with EC_{50} values ranging 0.036 – 0.14 μM .²⁰

X-Ray Crystallography

The binding mode of inhibitor **19b** was determined from the X-ray crystal structure of its complex with wild-type HIV-1 protease. The crystal structure was solved and refined at 1.29 Å resolution with an R factor of 14.1 %. In this high resolution structure, the inhibitor was

bound to the HIV-1 protease active site in two orientations with the relative occupancy of 0.8/0.2. The protease dimer comprises residues 1–99 and 1'–99' of the two subunits, and the inhibitor binding site is formed by both subunits. The P1'-pyrrolidine ring also showed two alternate conformations with equal occupancy and related by about 18° rotation around the C12–C13 bond. A stereoview of the major conformation is shown in Figure 2 [only one conformation is shown for P1']. As shown, extensive interactions from P2 to P2' were observed between the inhibitor and protease active site, most notably favorable polar interactions including hydrogen bonds, weaker C-H...O and C-H... π interactions. The isostere hydroxyl group forms asymmetric hydrogen bonds to the carboxylate oxygen atoms of the catalytic Asp25 and Asp25' with distances of 2.4 to 3.3 Å. Also, four direct hydrogen bonds are formed between oxygens or nitrogens of the inhibitor atoms and the protease backbone atoms. These include: cyclic ether oxygen of the P2-Cp-THF and the Asp-29 NH; the urethane NH with the carbonyl oxygen of Gly-27; P2'-methoxy oxygen and Asp-30'. One conformation of the P1'-pyrrolidinone formed a hydrogen bond between the NH and the carbonyl oxygen of Gly-27' and a water-mediated hydrogen bond between the P1'-pyrrolidinone carbonyl and the side chain of Arg-8. The other conformation of the P1' group formed hydrophobic and C-H...O interactions with Pro-81' and Val-82', as shown in Figure 3. Also, there exists a tetracoordinated water-mediated interaction where the amides of Ile50 and 50' donate hydrogen bonds and the inhibitor's urethane carbonyl and one of the sulfonamide oxygen accept hydrogen bonds from the water molecule. These interactions are conserved in a majority of other HIV-1 protease complexes with inhibitors,²¹ or substrate analogs.²²

The weaker polar interactions such as C-H...O and water- π interactions can be analyzed accurately in this high resolution structure.^{23,24} These interactions are important for inhibitor-protease binding and must be considered in the design of inhibitors. The C-H...O interactions of the inhibitor with the carbonyl oxygens of Gly-48, Gly-48' and Gly-27' mimic the conserved hydrogen bonds observed peptide analog structures^{21,22}. A conserved water- π interaction is observed between the P2' aromatic ring of the inhibitor and the amide of Asp29', which is similar to the interaction with darunavir and other structure-based designed PIs from our laboratories.^{25,26} The bigger polar P1' group of the 2-pyrrolidinone ring in inhibitor **19b** instead of the isobutyl group in PI's **1** and **2** introduces a new direct hydrogen bond with the backbone of HIV-1 protease and one new water-mediated hydrogen bond between the inhibitor and the side chain residue of the protease. The two alternate conformations of the P1' group with occupancy of 0.5/0.5 provide more flexible binding within the S1' subsite, which is likely to enhance the inhibition of resistant proteases.

As mentioned earlier, inhibitor **19b** maintained near full potency against multi-drug-resistant clinical isolates examined. On the other hand, **19b** is less potent than darunavir possibly due to the bigger and less optimum size of the P1'-ligand. The design strategy of incorporating new polar interactions with conserved backbone regions of the protease warrants further investigation in light of the current molecular insight into these ligand-binding site interactions.

Conclusion

We have designed a number of HIV-1 protease inhibitors with methyl-2-pyrrolidinone and methyl oxazolidinone as the P1'-ligand to enhance hydrogen bonding with the protein backbone atoms in the S1'-subsite. The ligands were synthesized in enantiomerically pure forms and a series of inhibitors were prepared and evaluated in combination with P2-bis-THF and P2-Cp-THF ligands. In general, these inhibitors exhibited enzyme inhibitory activity lower than the corresponding inhibitors with a P1'-isobutyl group. Our SAR studies indicated the importance of ligand stereochemistry and also preference for the P2-Cp-THF ligand. Interestingly, the polar P1'-ligand influenced their cellular properties. The inhibitors exhibited reduction in antiviral activity possibly due to changes in the molecule's overall lipophilicity. Our

investigation resulted in the identification of inhibitor **19b** which has displayed similar antiviral potency as the other FDA approved inhibitors such as indinavir, lopinavir and amprenavir. Inhibitor **19b**, however, is nearly 10-fold less potent than darunavir. Of particular importance, **19b** has maintained full potency against the examined panel of multidrug-resistant HIV-1 variants. A high resolution X-ray structure of **19b**-bound HIV-1 protease revealed a new hydrogen bonding of the P1'-pyrrolidinone NH with the backbone carboxyl of Gly27'. Also, there is a water mediated hydrogen bond with the pyrrolidinone carboxyl and Arg8' side chain. Furthermore, the P1'-pyrrolidinone showed two alternate conformations that filled the S1' subsite. These new interactions and the conformational flexibility most likely contributed to its impressive properties against multidrug-resistant clinical variants. Further investigations including optimization of ligand-binding properties are in progress.

Experimental Section

General

All moisture sensitive reactions were carried out under nitrogen or argon atmosphere. Anhydrous solvents were obtained as follows: THF, diethyl ether and benzene, distilled from sodium and benzophenone; dichloromethane, pyridine, triethylamine, and diisopropylethylamine, distilled from CaH₂. All other solvents were HPLC grade. Silica gel column chromatography was performed with Whatman 240–400 mesh silica gel under low pressure. TLC was carried out with E. Merck silica gel 60-F-254 plates. ¹H and ¹³C NMR spectra were recorded on Varian Mercury 300 and Bruker Avance 400 and 500 spectrometers. Optical rotations were measured using a Perkin-Elmer 341 polarimeter.

(S)-5-(Aminomethyl)-2-pyrrolidinone **6**

To a stirred solution of (S)-5-(hydroxymethyl)-2-pyrrolidinone **5** (300 mg, 2.61 mmol) and *p*-toluenesulfonyl chloride (646 mg, 3.34 mmol) in CH₂Cl₂ (6 mL) at 0 °C was added DMAP (64 mg, 0.52 mmol) and Et₃N (472 μL, 3.34 mmol). The resulting mixture was allowed to warm to 23 °C and stirred for 12 h. The reaction was then quenched with 7 mL of water and the aqueous layer was extracted with CH₂Cl₂. The combined organic extracts were washed with 1N HCl and dried over anhydrous Na₂SO₄. Removal of solvent under reduced pressure, followed by flash chromatography purification (100% EtOAc as the eluent) yielded (S)-toluenesulfonate (0.7 g, 93%) as a yellowish solid. *R*_f = 0.50 (5% MeOH in CHCl₃); ¹H-NMR (400 MHz, CDCl₃) δ 1.75–1.80 (m, 1H), 2.19–2.35 (m, 3H), 2.44 (s, 3H), 3.85–3.92 (m, 2H), 4.00–4.03 (m, 1H), 6.49 (s, 1H), 7.35 (d, 2H, *J* = 8.08 Hz), 7.77 (d, 2H, *J* = 8.17 Hz); ¹³C-NMR (100 MHz, CDCl₃) δ 21.68, 22.77, 29.27, 52.59, 71.97, 121.92, 130.08, 132.32, 145.37, 178.20.

To a stirred solution of the above tosylate (638 mg, 2.37 mmol) in DMF (20 mL) was added NaN₃ (462 mg, 2.37 mmol). The resulting solution was stirred at 55 °C for 9 h. Removal of solvent under reduced pressure, followed by flash chromatography purification (6% MeOH in CHCl₃ as the eluent) provided the (S)-azidopyrrolidinone (330 mg, 99%) as a yellow oil. *R*_f = 0.50 (10% MeOH in CHCl₃); ¹H-NMR (400 MHz, CDCl₃) δ 1.76–1.84 (m, 1H), 2.18–2.44 (m, 3H), 3.28 (dd, 1H, *J* = 6.5, 12.25 Hz), 3.43 (dd, 1H, *J* = 4.6, 12.3 Hz), 3.77–3.83 (m, 1H), 7.38 (s, 1H); ¹³C-NMR (100 MHz, CDCl₃) δ 23.97, 29.79, 53.66, 55.89, 178.73.

To a solution of the above azide (125 mg, 0.89 mmol) in EtOAc (10 mL) was added Pd/C (15 mg). The mixture was stirred at 23 °C under a hydrogen filled balloon for 4 h, then filtered over Celite and the filter cake was washed with EtOAc and MeOH. Removal of solvent under reduced pressure, followed by flash chromatography purification (5% MeOH in CHCl₃ as the eluent) afforded the corresponding (S)-amine **6** (105 mg, quantitative) as a yellow oil. *R*_f = 0.05 (20% MeOH in CHCl₃); ¹H-NMR (400 MHz, CDCl₃) δ 1.41 (brs 2H), 1.56–1.65 (m, 1H),

2.01–2.12 (m, 1H), 2.19–2.24 (m, 2H), 2.52 (dd, 1H, $J = 7.5, 12.8$ Hz), 2.69 (dd, 1H, $J = 4.3, 12.9$ Hz), 3.50–3.57 (m, 1H), 7.90 (brs, 1H); ^{13}C -NMR (100 MHz, CDCl_3) δ 24.07, 30.29, 47.34, 57.10, 179.03.

***tert*-Butyl(2*R*,3*R*)-3-hydroxy-4-(((*S*)-5-oxopyrrolidin-2-yl)methylamino)-1-phenylbutan-2-yl-carbamate 8**

To a solution of amine **6** (107 mg, 0.94 mmol) in $i\text{PrOH}$ (5 mL) were added *tert*-butyl-[(*S*-(*R,R*)]-(-)-(1-oxiran-2-yl)-2-phenylethyl]carbamate **7** (62 mg, 0.23 mmol) and $i\text{Pr}_2\text{EtN}$ (204 μL , 1.2 mmol). The resultant mixture was stirred at 65 °C for 18 h and then concentrated under reduced pressure. Flash chromatography purification (15% MeOH in CHCl_3 as the eluent) yielded title compound **8** (76 mg, 85%). $R_f = 0.47$ (25% MeOH in CHCl_3); ^1H -NMR (400 MHz, CDCl_3) δ 1.30 (s, 9H), 1.62–1.71 (m, 1H), 2.13–2.18 (m, 1H), 2.30–2.32 (m, 2H), 2.52 (d, 1H, $J = 8.86$ Hz), 2.64–2.73 (m, 4H), 2.96 (d, 1H, $J = 9.8$ Hz), 3.54 (s, 1H), 3.72–3.75 (m, 4H), 4.99 (brs, 1H), 7.15–7.26 (m, 5H), 8.02 (s, 1H); ^{13}C -NMR (100 MHz, CDCl_3) δ 24.65, 28.32, 30.29, 36.37, 51.74, 54.09, 54.41, 55.30, 71.53, 79.26, 126.20, 128.29, 129.47, 138.17, 155.96, 178.98; LRMS-ESI (m/z) [$M + \text{Na}$] $^+$ 400.

***tert*-Butyl(2*R*,3*R*)-3-hydroxy-4-(4-methoxy-*N*-(((*S*)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-ylcarbamate 9**

To a stirred solution of amine **8** (22 mg, 0.06 mmol) in CH_2Cl_2 (3 mL) and aqueous sat. NaHCO_3 (3 mL) was added 4-methoxybenzenesulfonyl chloride (35.6 mg, 0.17 mmol). This reaction mixture was stirred for 12 h followed by extraction of the aqueous layer with CH_2Cl_2 ; the combined organic extracts were dried over anhydrous Na_2SO_4 . Removal of solvent under reduced pressure, followed by flash chromatography purification (6% MeOH in CHCl_3 as the eluent) provided compound **9** (31 mg, quantitative). $R_f = 0.40$ (10% MeOH in CHCl_3); ^1H -NMR (500 MHz, CDCl_3) δ 1.28 (s, 9H), 1.54–1.62 (m, 1H), 2.14–2.21 (m, 1H), 2.32–2.35 (m, 2H), 2.68–2.75 (m, 2H), 2.72 (s, 3H), 2.81–2.88 (m, 2H), 2.97–3.03 (m, 3H), 3.64–3.72 (m, 1H), 4.01–4.05 (m, 1H), 5.06 (d, 1H, $J = 8.9$ Hz), 6.93 (d, 2H, $J = 8.6$ Hz), 7.16–7.19 (m, 3H), 7.27–7.28 (m, 3H), 7.61 (d, 2H, $J = 8.5$ Hz); ^{13}C -NMR (125 MHz, CDCl_3) δ 23.94, 28.16, 29.62, 29.93, 36.10, 53.40, 53.99, 54.57, 55.56, 56.01, 71.96, 79.51, 114.38, 126.31, 126.37, 129.37, 129.52, 137.79, 155.93, 163.15, 178.42; LRMS-ESI (m/z) [$M + \text{Na}$] $^+$ 570.

***tert*-Butyl(2*R*,3*R*)-4-(4-Cbz-amino-*N*-(((*S*)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-ylcarbamate 10**

To a stirred solution of amine **8** (93.6 mg, 0.25 mmol) in CH_2Cl_2 (10 mL) and aqueous sat. NaHCO_3 (10 mL) was added 4-nitrobenzenesulfonyl chloride (60 mg, 0.27 mmol). This reaction mixture was stirred for 7 h followed by extraction of the aqueous layer with CH_2Cl_2 ; the combined organic extracts were dried over anhydrous Na_2SO_4 . Removal of solvent under reduced pressure, followed by flash chromatography purification (dry transfer, 8% MeOH in CHCl_3 as the eluent) provided (*S*)-nitrosulfone (112 mg, 80%) as a yellowish solid. $R_f = 0.56$ (10% MeOH in CHCl_3).

The above nitrosulfone (103 mg, 0.18 mmol) was dissolved in EtOAc (20 mL) and Pd/C (11 mg) was added. The mixture was stirred under a hydrogen filled balloon for 8 h at 23 °C. It was then filtered over Celite and the filter cake was washed with EtOAc and MeOH. Removal of solvent under reduced pressure, followed by flash chromatography purification (2.5% MeOH in CHCl_3 as the eluent) afforded the corresponding (*S*)-amine (77 mg, 79%) as a white solid. $R_f = 0.26$ (5% MeOH in CHCl_3); ^1H -NMR (500 MHz, CDCl_3) 1.35 (s, 9H), 1.59–1.57 (m, 1H), 2.15–2.23 (m, 1H), 2.29–2.42 (m, 2H), 2.82–2.87 (m, 3H), 3.02 (dd, 1H, $J = 4.75, 14.0$), 3.13 (dd, 1H, $J = 10, 13.2$ Hz), 3.30 (dd, 2H, $J = 1.8, 14.4$ Hz), 3.73–3.82 (m, 1H), 3.90–3.95 (m, 1H), 3.99 (d, 1H, $J = 6.2$ Hz), 4.72 (d, 1H, $J = 8.2$ Hz), 6.72 (d, 2H, $J = 7.9$ Hz), 7.20–

7.32 (m, 5H), 7.37 (s, 1H), 7.57 (d, 2H, $J = 8.2$ Hz); ^{13}C -NMR (125 MHz, CDCl_3) δ 24.35, 28.69, 30.09, 36.58, 53.67, 54.42, 54.92, 56.66, 72.63, 79.85, 114.48, 125.07, 126.71, 128.78, 129.92, 130.09, 138.38, 151.58.

To a stirred solution of the above amine (33.1 mg, 0.0621 mmol) in CH_2Cl_2 (3 mL) was added pyridine (30 μL , 0.372 mmol) and benzyl chloroformate (20 μL , 0.137 mmol). This reaction mixture was stirred for 3 h, then quenched with 5 drops of benzyl amine, followed by removal of solvent under reduced pressure. Column chromatography over silica gel (2.5% MeOH in CHCl_3 as the eluent) provided (*S*)-Cbz-amine ¹⁰ (41 mg, 99%) as a white solid. $R_f = 0.37$ (5% MeOH in CHCl_3); ^1H -NMR (500 MHz, CDCl_3) δ 1.56 (brs, 1H), 2.12–2.18 (m, 1H), 2.26–2.41 (m, 2H), 2.71–2.78 (m, 2H), 2.79–89 (m, 1H), 3.02 (dd, 1H, $J = 8.2$, 18.0 Hz), 3.19–3.26 (m, 1H), 3.38 (d, 1H, $J = 14.4$ Hz), 3.77 (brs, 1H), 3.94–3.99 (m, 2H), 4.73 (d, 1H, $J = 8.5$ Hz), 5.21 (s, 2H), 7.19–7.22 (m, 3H), 7.25–7.29 (m, 3H), 7.31–7.39 (m, 4H), 7.57 (d, 2H, $J = 7.6$ Hz), 7.69 (d, 2H, $J = 7.7$ Hz); ^{13}C -NMR (125 MHz, CDCl_3) δ 24.16, 28.74, 30.12, 36.65, 53.38, 54.34, 54.89, 56.89, 67.79, 72.64, 80.01, 118.63, 126.83, 128.77, 128.85, 128.93, 129.08, 129.43, 129.99, 131.66, 136.03, 138.13, 143.07, 153.37, 156.33, 178.99; LRMS-ESI (m/z) $[\text{M} + \text{Na}]^+ 689$.

***tert*-Butyl(2*R*,3*R*)-3-hydroxy-4-(4-methoxy-*N*-(((*R*)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-ylcarbamate 11**

To a stirred solution of (*R*)-5-(hydroxymethyl)-2-pyrrolidinone ent-⁵ (500 mg, 4.34 mmol) and *p*-toluenesulfonyl chloride (1.08 g, 5.6 mmol) CH_2Cl_2 (10 mL) at 0 °C was added DMAP (106 mg, 0.87 mmol) and Et_3N (780 μL , 5.6 mmol). The resulting mixture was allowed to warm to 23 °C and stirred for 12 h. The reaction was then quenched with 10 mL of water and the aqueous layer was extracted with CH_2Cl_2 . The combined organic extracts were washed with 1N HCl and dried over anhydrous Na_2SO_4 . Removal of solvent under reduced pressure, followed by flash chromatography (2.5% MeOH in CHCl_3 as the eluent) yielded the (*R*)-toluenesulfonate (1.8 g, 93%) as a yellowish solid. $R_f = 0.50$ (5% MeOH in CHCl_3); ^1H -NMR (400 MHz, CDCl_3) δ 1.70–1.79 (m, 1H), 2.13–2.35 (m, 3H), 2.42 (s, 3H), 3.84–3.89 (m, 2H), 3.96–3.04 (m, 1H), 6.76 (s, 1H), 7.33 (d, 2H, $J = 8.04$ Hz), 7.76 (d, 2H, $J = 8.2$ Hz); ^{13}C -NMR (100 MHz, CDCl_3) δ 21.67, 22.76, 29.33, 52.61, 71.95, 127.91, 130.06, 132.30, 145.33, 178.11.

To a stirred solution of the above toluenesulfonate (1.08 g, 4.03 mmol) in DMF (30 mL) was added NaN_3 (1.31 g, 20.2 mmol). The resulting solution was stirred at 55 °C for 12 h. Solvent was then removed under reduced pressure, followed by flash chromatography purification (6% MeOH in CHCl_3 as the eluent) provided the (*R*)-azidopyrrolidinone (558 mg, 99%) as a yellow oil. $R_f = 0.44$ (5% MeOH in CHCl_3); ^1H -NMR (500 MHz, CDCl_3) δ 1.72–1.79 (m, 1H), 2.13–2.37 (m, 3H), 3.22 (dd, 1H, $J = 6.3$, 12.3 Hz), 3.37 (dd, 1H, $J = 4.7$, 12.3 Hz), 3.72–3.78 (m, 1H), 7.69 (s, 1H); ^{13}C -NMR (125 MHz, CDCl_3) δ 24.32, 30.26, 54.13, 56.19, 179.32.

To a solution of the above azide (528 mg, 3.77 mmol) in EtOAc (35 mL) was added Pd/C (40 mg). The mixture was stirred at 23 °C under a hydrogenfilled balloon for 4 h, then filtered over Celite and the filter cake was washed with EtOAc and MeOH. Removal of solvent under reduced pressure, followed by flash chromatography purification (5% MeOH in CHCl_3 as the eluent) afforded the (*R*)-amine ent-⁶ (257 mg, 95%) as a yellow oil. $R_f = 0.05$ (20% MeOH in CHCl_3); ^1H -NMR (400 MHz, CDCl_3) δ 1.59–1.69 (m, 1H), 2.06–2.15 (m, 1H), 2.23–2.30 (m, 4H), 2.57 (dd, 1H, $J = 7.6$, 12.8 Hz), 2.74 (dd, 1H, $J = 4.1$, 12.9 Hz), 3.56–3.65 (m, 1H), 7.80 (brs, 1H); ^{13}C -NMR (100 MHz, CDCl_3) δ 24.12, 30.23, 47.11, 56.84, 179.04.

To a solution of amine ent-⁶ (430 mg, 3.76 mmol) in $i\text{PrOH}$ (20 mL) were added *tert*-butyl-[*S*-(*R,R*)]-(1-oxiranyl-2-phenylethyl)carbamate ⁷ (250 mg, 0.94 mmol) and $i\text{Pr}_2\text{EtN}$ (1.5 mL, 8.6 mmol). The resultant mixture was stirred at 65 °C for 36 h and then concentrated under reduced pressure. Flash chromatography purification (10% MeOH in CHCl_3 as the eluent)

yielded the (*R*)-hydroxyamine (**8R**) (300 mg, 84%). R_f 0.33 (20% MeOH in CHCl_3); ^1H -NMR (400 MHz, CDCl_3) δ 1.33 (s, 9H), 1.64–1.73 (m, 1H), 2.13–2.22 (m, 1H), 2.28–2.34 (m, 2H), 2.54 (dd, 1H, J = 9.0, 11.8 Hz), 2.65 (dd, 1H, J = 7.2, 13.2 Hz), 2.73–2.85 (m, 4H), 2.94 (dd, 1H, J = 4.4, 14.0 Hz), 3.43 (s, 1H), 3.51–3.60 (m, 1H), 3.70–3.76 (m, 1H), 3.79–3.84 (m, 1H), 5.02 (d, 1H, J = 8.9 Hz), 7.16–7.27 (m, 5H), 7.94 (s, 1H); ^{13}C -NMR (100 MHz, CDCl_3) δ 24.64, 28.32, 30.28, 36.37, 52.14, 54.29, 54.69, 55.46, 71.62, 79.26, 126.21, 128.30, 129.47, 138.13, 155.93, 178.99; LRMS-ESI (m/z) [$\text{M} + \text{Na}$] $^+$ 400.

To a stirred solution of above (*R*)-hydroxyamine (**8R**) (40 mg, 0.105 mmol) in CH_2Cl_2 (4 mL) and aqueous sat. NaHCO_3 (4 mL) was added 4-methoxybenzenesulfonyl chloride (66 mg, 0.318 mmol). This reaction mixture was stirred for 12 h followed by extraction of the aqueous layer with CH_2Cl_2 ; the combined organic extracts were dried over anhydrous Na_2SO_4 . Removal of solvent under reduced pressure, followed by flash chromatography purification (4% MeOH in CHCl_3 as the eluent) provided compound **11** (54 mg, 93%). R_f 0.40 (10% MeOH in CHCl_3); ^1H -NMR (400 MHz, CDCl_3) δ 1.32 (s, 9H), 1.64–1.68 (m, 1H), 2.17–2.21 (m, 2H), 2.34–2.40 (m, 2H), 2.76–2.84 (m, 1H), 2.91–3.06 (m, 3H), 3.16–3.29 (m, 2H), 3.75–3.80 (m, 1H), 3.84 (s, 3H), 3.96–4.02 (m, 2H), 4.99 (d, 1H, J = 8.7 Hz), 6.95 (d, 2H, J = 8.8 Hz), 7.16–7.28 (m, 5H), 7.68 (d, 2H, J = 8.8 Hz), 7.93 (s, 1H); ^{13}C -NMR (100 MHz, CDCl_3) δ 24.41, 28.29, 29.99, 35.78, 35.46, 54.71, 55.21, 55.63, 55.88, 57.89, 73.94, 79.50, 114.43, 126.29, 128.37, 129.31, 129.51, 138.05, 155.91, 163.12, 178.64; LRMS-ESI (m/z) [$\text{M} + \text{Na}$] $^+$ 670.

tert*-Butyl(2*R*,3*R*)-4-(4-Cbz-amino-*N*-(((*R*)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-ylcarbamate **12*

To a stirred solution of (*R*)-hydroxyamine (**8R**) (116 mg, 0.3 mmol) in CH_2Cl_2 (10 mL) and aqueous sat. NaHCO_3 (10 mL) was added 4-nitrobenzenesulfonyl chloride (74 mg, 0.33 mmol). This reaction mixture was stirred for 12 h followed by extraction of the aqueous layer with CH_2Cl_2 ; the combined organic extracts were dried over anhydrous Na_2SO_4 . Removal of solvent under reduced pressure, followed by flash chromatography purification (dry transfer, 5% MeOH in CHCl_3 as the eluent) provided the (*R*)-nitrosulfone (164 mg, 96%) as a yellowish solid. R_f 0.56 (10% MeOH in CHCl_3).

The above nitrosulfone (154 mg, 0.27 mmol) was redissolved in EtOAc (25 mL) and treated with Pd/C (16 mg) under argon. Argon was then replaced with a hydrogenfilled balloon and the reaction was allowed to stir for 12 h at 23 °C. It was then filtered over Celite and the filter cake was washed with EtOAc and MeOH. Removal of solvent under reduced pressure, followed by flash chromatography purification (6% MeOH in CHCl_3 as the eluent) afforded the corresponding (*R*)-aniline (123 mg, 83%) as an amorphous solid. R_f 0.45 (10% MeOH in CHCl_3); ^1H -NMR (500 MHz, CDCl_3) δ 1.31 (s, 9H), 1.58–1.64 (m, 1H), 2.15–2.21 (m, 1H), 2.29 (t, 2H, J = 8.2 Hz), 2.73–2.86 (m, 3H), 2.99 (dd, 1H, J = 4.4, 13.9 Hz), 3.23 (d, 1H, J = 13.8 Hz), 3.30 (d, 1H, J = 14.8 Hz), 3.74 (brs, 1H), 3.92 (brs, 1H), 3.99 (d, 1H, J = 5.7), 4.31 (s, 1H), 5.01 (d, 1H, J = 9.1 Hz), 6.63 (d, 2H, J = 8.5 Hz), 7.16–7.21 (m, 3H), 7.24–7.27 (m, 2H), 7.48 (d, 2H, J = 8.5 Hz); ^{13}C -NMR (125 MHz, CDCl_3) δ 24.35, 28.16, 29.83, 35.59, 54.66, 55.09, 55.50, 57.71, 73.83, 79.51, 113.98, 125.09, 126.23, 128.28, 129.34, 129.41, 137.83, 151.02, 155.96, 178.25; LRMS-ESI (m/z) [$\text{M} + \text{Na}$] $^+$ 555.

To a stirred solution of the above (*R*)-aniline (101 mg, 0.19 mmol) in CH_2Cl_2 (15 mL) was added pyridine (34 μL , 0.41 mmol) and benzyl chloroformate (60 μL , 0.41 mmol). This reaction mixture was stirred for 1.5 h, then quenched with 3 drops of benzyl amine, followed by removal of solvent under reduced pressure. Column chromatography over silica gel (6% MeOH in CHCl_3 as the eluent) provided the (*R*)-Cbz-amine **12** (120 mg, 95%) as a white solid. R_f 0.35 (10% MeOH in CHCl_3); ^1H -NMR (500 MHz, CDCl_3) δ 1.29 (s, 9H), 1.55–1.65 (m, 1H), 2.15–2.23 (m, 1H), 2.26–2.31 (m, 2H), 2.60–2.75 (m, 3H), 2.97 (dd, 1H, J = 8.2, 18.1

Hz), 3.29 (d, 1H, $J = 17.7$ Hz), 3.36 (dd, 1H, $J = 2.4, 14.9$ Hz), 3.64 (s, 1H), 3.88–3.92 (m, 1H), 3.98–4.02 (m, 1H), 5.12 (d, 1H, $J = 9.0$ Hz), 5.17 (s, 2H), 7.14–7.19 (m, 3H), 7.21–7.28 (m, 3H), 7.30–7.37 (m, 4H), 7.54 (d, 2H, $J = 8.6$ Hz), 7.63 (d, 2H, $J = 8.8$); ^{13}C -NMR (125 MHz, CDCl_3) δ 24.82, 28.61, 30.28, 36.06, 55.14, 55.85, 55.98, 58.25, 67.65, 74.40, 80.08, 118.66, 126.76, 128.63, 128.77, 128.86, 129.03, 129.06, 129.77, 130.32, 131.28, 136.11, 138.25, 143.43, 153.68, 178.97; LRMS-ESI (m/z) $[\text{M} + \text{Na}]^+$ 689.

(3R,3aS,6aR)-Hexahydrofuro[2,3-b]furan-3-yl(2S,3R)-3-hydroxy-4-(4-methoxy-N-(((S)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-yl-carbamate 17a

A solution of compound ⁹ (10 mg, 0.02 mmol) in 30% trifluoroacetic acid (in CH_2Cl_2 , 3 mL) was stirred at 23 °C for 40 min, then concentrated under reduced pressure to give the crude amine ¹³(S). This residue was redissolved in CH_2Cl_2 (3 mL), treated with Et₃N (20 μL , 0.13 mmol), followed by carbonate ¹⁵ (5.5 mg, 0.02 mmol) and stirred at 23 °C for 12 h. The reaction mixture was then concentrated under reduced pressure and the residue was purified by flash chromatography (2% MeOH in CHCl_3 as the eluent) to give inhibitor **17a** (11.3 mg, 98%) as a white solid, $R_f = 0.48$ (10% MeOH in CHCl_3); ^1H -NMR (500 MHz, CDCl_3) δ 1.48 (dd, 1H, $J = 5.5, 13.2$ Hz), 1.58–1.68 (m, 2H), 2.17–2.26 (m, 1H), 2.34–2.49 (m, 2H), 2.76 (dd, 1H, $J = 9.8, 14.0$ Hz), 2.85–2.95 (m, 3H), 3.10–3.16 (m, 2H), 3.22 (dd, 1H, (dd, 1H, $J = 9.9, 13.7$ Hz), 3.67–3.74 (m, 2H), 3.82–3.85 (dt, 1H, $J = 1.8, 8.4$ Hz), 3.87 (s, 3H), 3.94 (dd, 1H, $J = 6.2, 9.6$ Hz), 3.96–4.01 (m, 1H), 4.04–4.08 (m, 1H), 5.0 (q, 1H, $J = 6.1, 7.9$ Hz), 5.64 (d, 1H, $J = 5.2$ Hz), 6.98 (d, 2H, $J = 8.9$ Hz), 7.19–7.29 (m, 3H), 7.26–7.29 (m, 2H), 7.58 (brs, 1H), 7.69 (d, 2H, $J = 8.8$ Hz); ^{13}C -NMR (125 MHz, CDCl_3) δ 23.97, 25.75, 29.59, 35.94, 45.29, 53.44, 53.94, 55.16, 55.60, 56.47, 69.53, 70.88, 72.39, 73.34, 109.24, 114.42, 126.45, 128.34, 128.63, 129.26, 129.49, 137.60, 155.47, 163.24, 178.51. LRMS-ESI (m/z) $[\text{M} + \text{H}]^+$ 604.2; HRMS-ESI (m/z) $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{29}\text{H}_{38}\text{N}_3\text{O}_9\text{S}$ 604.2329 found 604.23

(3aS,5R,6aR)-Hexahydro-2H-cyclopenta[b]furan-5-yl(2S,3R)-3-hydroxy-4-(4-methoxy-N-(((S)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-ylcarbamate 17b

A solution of compound ⁹ (11.8 mg, 0.02 mmol) in 30% trifluoroacetic acid (in CH_2Cl_2 , 1.5 mL) was stirred at 23 °C for 40 min, then concentrated under reduced pressure to give the crude amine ¹³S. This crude residue was redissolved in CH_2Cl_2 (1.5 mL), treated with Et₃N (63 μL , 0.45 mmol), followed by carbonate ¹⁶ (6.4 mg, 0.02 mmol), and stirred at 23 °C for 6 h. The reaction mixture was then concentrated under reduced pressure and the residue was purified by flash chromatography (1% MeOH in CHCl_3 as the eluent) to give inhibitor **17b** (11.5 mg, 87%) as a white solid, $R_f = 0.35$ (5% MeOH in CHCl_3); ^1H -NMR (500 MHz, CDCl_3) δ 1.45 (d, 1H, $J = 14.3$ Hz), 1.55–1.59 (m, 1H), 1.88 (d, 1H, $J = 15.1$ Hz), 1.95–2.06 (m, 3H), 2.17–2.24 (m, 1H), 2.33–2.48 (m, 2H), 2.60–2.67 (m, 1H), 2.78 (dd, 1H, $J = 9.1, 14.1$ Hz), 2.88–2.97 (m, 2H), 3.09 (dd, 1H, $J = 4.3, 14.1$ Hz), 3.12–3.18 (m, 2H), 3.64–3.68 (m, 1H), 3.82–3.85 (m, 2H), 3.86 (s, 3H), 3.89–3.95 (m, 1H), 3.99–4.05 (m, 1H), 4.39–4.42 (m, 1H), 4.69 (d, 1H, $J = 4.1$ Hz), 4.87–4.90 (m, 1H), 4.91 (d, 1H, $J = 8.9$ Hz), 6.98 (d, 2H, $J = 8.9$ Hz), 7.20–7.23 (m, 3H), 7.27–7.30 (m, 2H), 7.42 (s, 1H), 7.70 (d, 2H, $J = 8.9$ Hz); ^{13}C -NMR (125 MHz, CDCl_3) δ 24.00, 29.89, 33.85, 35.99, 38.31, 39.43, 41.59, 53.26, 53.33, 53.91, 54.89, 55.56, 56.36, 67.64, 72.23, 83.71, 114.39, 126.42, 128.43, 128.78, 129.33, 129.53, 137.54, 156.18, 163.11, 178.32. LRMS-ESI (m/z) $[\text{M} + \text{Na}]^+$ 624.0; HRMS-ESI (m/z) $[\text{M} + \text{Na}]^+$ calcd for $\text{C}_{30}\text{H}_{39}\text{N}_3\text{NaO}_8\text{S}$ 624.2356 found 624.2352.

(3R,3aS,6aR)-Hexahydrofuro[2,3-b]furan-3-yl(2S,3R)-4-(4-amino-N-(((S)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-ylcarbamate 18a

The Cbz-protected amine ¹⁰ (31 mg, 0.04 mmol) was treated with 30% trifluoroacetic acid in CH_2Cl_2 (6 mL) and stirred at 23 °C for 40 min, then concentrated under reduced pressure to give the crude amine ¹⁴-(S). This residue was redissolved in CH_2Cl_2 (6 mL), charged with

Et₃N (64 μ L, 0.46 mmol), followed by carbonate ¹⁵ (14 mg, 0.05 mmol), and stirred at 23 °C for 12 h. Reaction was quenched with 3 drops of benzyl amine and concentrated under reduced pressure. Flash chromatography purification (4% MeOH in CHCl₃ as the eluent) provided the Cbz-protected inhibitor (23 mg, 86%) as a white solid, *R_f* = 0.46 (10% MeOH in CHCl₃).

To above Cbz-protected inhibitor (13.3 mg, 0.018 mmol), in EtOAc (6 mL) under argon, was added Pd/C (3 mg). The mixture was stirred at 23 °C under a hydrogenfilled balloon for 3 h, then filtered over Celite and the filter cake was washed with EtOAc and MeOH. Removal of solvent under reduced pressure, followed by flash chromatography purification (3% MeOH in CHCl₃ as the eluent) provided the title inhibitor **18a** (7.4 mg, 68%) as a white solid. *R_f* = 0.19 (5% MeOH in CHCl₃); ¹H-NMR (500 MHz, CDCl₃) δ 1.44 (d, 1H, *J* = 5.4 Hz), 1.59–1.69 (m, 2H), 2.16–2.25 (m, 1H), 2.36 (t, 2H, *J* = 7.9 Hz), 2.68 (dd, 1H, *J* = 9.8, 13.9 Hz), 2.85–2.95 (m, 3H), 3.03–3.09 (m, 2H), 3.13 (dd, 1H, *J* = 4.4, 14.1 Hz), 3.67–3.72 (m, 2H), 3.79–3.89 (m, 3H), 3.93 (dd, 1H, *J* = 6.0, 9.7 Hz), 4.01–4.06 (m, 1H), 4.96 (q, 1H, *J* = 5.9, 7.9 Hz), 5.63 (d, 1H, *J* = 5.1 Hz), 6.72 (d, 2H, *J* = 8.2 Hz), 7.17–7.21 (m, 3H), 7.24–7.28 (m, 2H), 7.50 (d, 2H, *J* = 8.4 Hz); ¹³C-NMR (125 MHz, CDCl₃) δ 24.38, 26.22, 30.07, 36.69, 45.83, 53.67, 54.28, 55.62, 56.69, 70.06, 71.51, 72.72, 73.64, 109.77, 114.63, 124.92, 126.82, 128.83, 129.72, 129.97, 130.34, 151.40, 155.98, 178.30. LRMS-ESI (*m/z*) [M+H]⁺ 589.2; HRMS-ESI (*m/z*) [M+H]⁺ calcd for C₂₈H₃₇N₄O₈S 589.2332 found 589.2336.

(3a*S*,5*R*,6a*R*)-Hexahydro-2H-cyclopenta[*b*]furan-5-yl-(2*S*,3*R*)-4-(4-amino-N-(((*S*)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-ylcarbamate 18b

The cbz-protected amine ¹⁰ (29.6 mg, 0.04 mmol) was treated with 30% trifluoroacetic acid (in CH₂Cl₂, 6 mL) and stirred at 23 °C for 40 min, then concentrated under reduced pressure to give the crude amine ¹⁴*S*. The residue was redissolved in CH₂Cl₂ (6 mL), charged with Et₃N (31 μ L, 0.22 mmol), followed by carbonate ¹⁶ (13.1 mg, 0.05 mmol), and stirred at 23 °C for 4 h. Reaction was quenched with 2 drops of benzyl amine and concentrated under reduced pressure. Flash chromatography purification (4% MeOH in CHCl₃ as the eluent) provided the cbz-protected inhibitor (26.1 mg, 82%) as a white solid, *R_f* = 0.49 (10% MeOH in CHCl₃). To a solution of the above protected inhibitor (17 mg, 0.02 mmol), in EtOAc (5 mL) under argon, was added Pd/C (3 mg). The mixture was stirred at 23 °C under a H₂ filled balloon for 5 h, then filtered over Celite and the filter cake was washed with EtOAc and MeOH. Removal of solvent under reduced pressure, followed by flash chromatography purification (5% MeOH in CHCl₃ as the eluent) provided inhibitor **18b** (14 mg, 75%) as a white solid. *R_f* = 0.27 (10% MeOH in CHCl₃); ¹H-NMR (500 MHz, CDCl₃) δ 1.41–1.45 (m, 1H), 1.56–1.67 (m, 3H), 1.85 (d, 1H, *J* = 13.4 Hz), 1.96–2.04 (m, 3H), 2.14–2.21 (m, 1H), 2.31–2.42 (m, 2H), 2.61 (brs, 1H), 2.74–2.85 (m, 3H), 3.09 (dd, 1H, *J* = 4.4, 14.4 Hz), 3.15–3.20 (m, 1H), 3.25 (d, 1H, *J* = 14.3 Hz), 3.66–3.70 (q, 1H, *J* = 7.1, 7.4 Hz), 3.83–3.88 (m, 2H), 3.91–3.96 (m, 1H), 3.96–4.25 (m, 1H), 4.40 (t, 1H, *J* = 5.9 Hz), 4.87 (brs, 1H), 4.93 (d, 1H, *J* = 8.9 Hz), 6.69 (d, 2H), 7.18–7.22 (m, 3H), 7.28–7.30 (m, 2H), 7.55 (d, 2H, *J* = 8.4 Hz), 7.63 (s, 1H); ¹³C-NMR (125 MHz, CDCl₃) δ 24.39, 30.12, 34.19, 36.71, 38.75, 39.85, 41.99, 53.56, 54.34, 55.31, 56.89, 68.09, 72.89, 84.17, 114.69, 125.34, 126.84, 128.87, 129.92, 130.19, 138.19, 151.43, 156.63, 178.99. LRMS-ESI (*m/z*) [M+H]⁺ 586.9; HRMS-ESI (*m/z*) [M+H]⁺ calcd for C₂₉H₃₉N₄O₇S 587.2539 found 587.2539.

(3*R*,3a*S*,6a*R*)-Hexahydrofuro[2,3-*b*]furan-3-yl-(2*S*,3*R*)-3-hydroxy-4-(4-methoxy-N-(((*R*)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-ylcarbamate 19a

A solution of compound ¹¹ (20 mg, 0.03 mmol) in 30% trifluoroacetic acid (in CH₂Cl₂, 3 mL) was stirred 23 °C for 40 min, then concentrated under reduced pressure. The residue was redissolved in CH₂Cl₂ (3 mL), treated with Et₃N (51 μ L, 0.36 mmol), followed by carbonate ¹⁵ (11 mg, 0.04 mmol), and stirred at 23 °C for 12 h. The reaction mixture was then concentrated under reduced pressure and the residue was purified by flash chromatography (2% MeOH in

CHCl₃ as the eluent) to give inhibitor **19a** (21 mg, 92%) as a white solid, R_f = 0.41 (10% MeOH in CHCl₃); ¹H-NMR (500 MHz, CDCl₃) δ 1.44 (dd, 1H, J = 5.6, 13.2 Hz), 1.57–1.67 (m, 2H), 1.81–1.91 (m, 1H), 2.19–2.27 (m, 1H), 2.33–2.41 (m, 2H), 2.73 (dd, 1H, J = 10.35, 13.9 Hz), 2.83–2.91 (m, 2H), 2.95 (dd, 1H, J = 8.9, 14.9 Hz), 3.11 (dd, 1H, J = 4.2, 14.0 Hz), 3.26–3.30 (ddd, 2H, J = 2.6, 7.0, 14.3 Hz), 3.64–3.70 (m, 1H), 3.74 (dd, 1H, J = 5.5, 9.8 Hz), 3.76–3.81 (dt, 1H, J = 1.62, 8.0 Hz), 3.87 (s, 3H), 3.89–3.93 (q, 1H, J = 4.0, 5.6 Hz), 4.03–4.06 (m, 2H), 4.98–5.02 (q, 1H, J = 5.65, 7.9 Hz), 5.62 (d, 1H, J = 5.4 Hz), 6.98 (d, 2H, J = 8.9 Hz), 7.17–7.27 (m, 5H), 7.70 (d, 2H, J = 8.8 Hz), 7.89 (s, 1H); ¹³C-NMR (125 MHz, CDCl₃) δ 24.25, 25.78, 29.89, 35.43, 45.43, 55.06, 55.37, 55.58, 55.77, 58.11, 69.56, 71.06, 73.34, 74.02, 109.29, 114.46, 126.34, 128.33, 128.88, 129.29, 129.41, 137.84, 155.52, 163.24, 178.51. LRMS-ESI (m/z) [M+Na]⁺ 626.3; HRMS-ESI (m/z) [M+Na]⁺ calcd for C₂₉H₃₇N₃ Na O₉S 626.2148 found 626.2156.

((3a*S*,5*R*,6a*R*)-Hexahydro-2H-cyclopenta[*b*]furan-5-yl-(2*S*,3*R*)-3-hydroxy-4-(4-methoxy-*N*-(((*R*)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-yl)carbamate **19b**

A solution of compound 11 (21 mg, 0.04 mmol) in 30% trifluoroacetic acid (in CH₂Cl₂, 3 mL) was stirred 23 °C for 40 min, then concentrated under reduced pressure. The residue was redissolved in CH₂Cl₂ (3 mL), treated with Et₃N (27 μL, 0.19 mmol), followed by carbonate **16** (12 mg, 0.04 mmol) and stirred at 23 °C for 12 h. The reaction mixture was then concentrated under reduced pressure and the residue was purified by flash chromatography (1% MeOH in CHCl₃ as the eluent) to give inhibitor **19b** (21 mg, 93%) as a white solid, R_f = 0.31 (5% MeOH in CHCl₃); ¹H-NMR (500 MHz, CDCl₃) δ 1.48 (d, 1H, J = 14.1 Hz), 1.58–1.62 (m, 1H), 1.83–2.04 (m, 5H), 2.19–2.25 (m, 1H), 2.31–2.41 (m, 2H), 2.59–2.67 (m, 1H), 2.73 (dd, 1H, J = 9.0, 13.9 Hz), 3.03 (dd, 1H, J = 7.0, 15.0 Hz), 3.08–3.16 (m, 2H), 3.19 (d, 1H, J = 14.9 Hz), 3.57–3.63 (m, 1H), 3.83–3.86 (m, 3H), 3.86 (s, 3H), 3.94–3.99 (m, 1H), 4.78 (d, 1H, J = 13.7 Hz), 4.90 (s, 1H), 5.36 (d, 1H, J = 8.1 Hz), 6.97 (d, 2H, J = 8.6 Hz), 7.18–7.28 (m, 5H), 7.46 (d, 1H, J = 18.4 Hz), 7.70 (d, 2H, J = 8.8 Hz); ¹³C-NMR (125 MHz, CDCl₃) δ 22.55, 30.00, 31.49, 33.97, 35.88, 38.25, 39.59, 41.76, 54.60, 54.91, 55.56, 56.08, 57.76, 67.72, 74.00, 83.97, 114.44, 126.21, 128.27, 128.90, 129.35, 129.47, 137.91, 156.16, 163.14, 178.52. LRMS-ESI (m/z) [M+H]⁺ 601.7; HRMS-ESI (m/z) [M+H]⁺ calcd for C₃₀H₄₀N₃O₈S 602.2536 found 602.2536.

(3*R*,3a*S*,6a*R*)-Hexahydrofuro[2,3-*b*]furan-3-yl-(2*S*,3*R*)-4-(4-amino-*N*-(((*R*)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-yl)carbamate **20a**

A solution of the free amine **12** (15 mg, 0.03 mmol) in 30% trifluoroacetic acid (in CH₂Cl₂, 3 mL) was stirred at 23 °C for 40 min, then concentrated under reduced pressure. The residue was redissolved in CH₂Cl₂ (3 mL), treated with Et₃N (40 μL, 0.28 mmol), followed by carbonate **15** (9.2 mg, 0.03 mmol), and stirred at 23 °C for 6 h. The reaction mixture was then concentrated under reduced pressure and the residue was purified by flash chromatography (4% MeOH in CHCl₃ as the eluent) to give inhibitor **20a** (12.5 mg, 94%) as a white solid, R_f = 0.26 (10% MeOH in CHCl₃); ¹H-NMR (500 MHz, CDCl₃) δ 1.35 (dd, 1H, J = 5.7 Hz), 1.50–1.63 (m, 2H), 2.15–2.23 (m, 1H), 2.27–2.31 (m, 2H), 2.56–2.72 (m, 3H), 2.82–2.87 (m, 2H), 3.07 (dd, 1H, J = 4.0, 14.0 Hz), 3.36 (q, 1H, J = 1.4, 3.5 Hz), 3.36 (q, 1H, J = 2.0, 3.4 Hz), 3.60–3.65 (m, 1H), 3.70 (dd, 1H, J = 5.4, 9.8 Hz), 3.75–3.79 (dt, 1H, J = 1.96, 8.3 Hz), 3.80–3.86 (m, 1H), 3.88 (dd, 1H, J = 5.9, 9.8 Hz), 3.90–3.94 (m, 1H), 3.99–4.02 (m, 1H), 4.92–4.97 (q, 1H, J = 5.7, 8.1 Hz), 5.58 (d, 1H, J = 5.8 Hz), 5.92 (d, 1H, J = 9.5 Hz), 6.63 (d, 2H, J = 8.7 Hz), 7.13–7.22 (m, 5H), 7.46 (d, 2H, J = 8.7 Hz); ¹³C-NMR (125 MHz, CDCl₃) δ 24.16, 25.70, 29.75, 35.39, 45.40, 55.01, 55.51, 55.61, 58.11, 69.55, 71.05, 73.19, 73.94, 109.27, 113.85, 124.27, 126.23, 128.22, 129.19, 129.35, 137.89, 151.44, 155.67, 178.63. LRMS-ESI (m/z) [M+Na]⁺ 611.4; HRMS-ESI (m/z) [M+Na]⁺ calcd for C₂₈H₃₆N₄NaO₈S 611.2152 found 611.2149.

(3a*S*,5*R*,6a*R*)-Hexahydro-2*H*-cyclopenta[*b*]furan-5-yl-(2*S*,3*R*)-4-(4-amino-*N*-(((*R*)-5-oxopyrrolidin-2-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-ylcarbamate 20b

The Cbz-amine ¹² (40 mg, 0.06 mmol) was treated with 30% trifluoroacetic acid (in CH₂Cl₂, 6 mL) and stirred at 23 °C for 40 min, then concentrated under reduced pressure. This residue was redissolved in CH₂Cl₂ (6 mL), charged with Et₃N (42 μL, 0.3 mmol), followed by carbonate ¹⁶ (19 mg, 0.07 mmol), and stirred at 23 °C for 12 h. Reaction was quenched with 3 drops of benzyl amine and concentrated under reduced pressure. Flash chromatography purification (5% MeOH in CHCl₃ as the eluent) provided the Cbz-protected inhibitor (32.1 mg, 75%) as a white solid, *R*_f = 0.41 (10% MeOH in CHCl₃).

To above Cbz-protected inhibitor (25.8 mg, 0.03 mmol), in EtOAc (5 mL) under argon, was added Pd/C (5 mg). The mixture was stirred at 23 °C under a hydrogen filled balloon for 3 h, then filtered over Celite and the filter cake was washed with EtOAc and MeOH. Removal of solvent under reduced pressure, followed by flash chromatography purification (7.5% MeOH in CHCl₃ as the eluent) provided the title inhibitor **20b** (16.2 mg, 77%) as a white solid. *R*_f = 0.51 (15% MeOH in CHCl₃); ¹H-NMR (500 MHz, CDCl₃) δ 1.46 (m, 1H), 1.58–1.64 (m, 1H), 1.77–1.84 (m, 1H), 1.86–2.04 (m, 5H), 2.15–2.23 (m, 1H), 2.32–2.36 (m, 2H), 2.59–2.65 (m, 1H), 2.68 (dd, 1H, *J* = 9.2, 14.0 Hz), 2.95–3.04 (m, 2H), 3.11 (d, 2H, *J* = 13.7 Hz), 3.18 (d, 1H, *J* = 14.9 Hz), 3.57–3.62 (q, 1H, *J* = 6.9, 7.7 Hz), 3.82–3.87 (q, 2H, *J* = 6.5, 7.9 Hz), 3.92–3.97 (m, 1H), 4.30 (s, 2H), 4.37 (t, 1H, *J* = 5.7 Hz), 4.75 (s, 1H), 4.89 (s, 1H), 5.40 (d, 1H, *J* = 8.4 Hz), 6.66 (d, 2H, *J* = 8.6 Hz), 7.16–7.22 (m, 3H), 7.24–7.27 (m, 2H), 7.40 (s, 1H), 7.51 (d, 2H, *J* = 8.6 Hz); ¹³C-NMR (125 MHz, CDCl₃) δ 23.72, 30.01, 33.94, 35.94, 38.18, 39.59, 41.78, 54.62, 54.85, 55.99, 57.64, 67.74, 74.04, 76.68, 84.01, 114.10, 124.98, 126.21, 128.27, 129.35, 129.48, 137.91, 151.08, 156.20, 178.40. LRMS-ESI (*m/z*) [M+Na]⁺ 609.0; HRMS-ESI (*m/z*) [M+Na]⁺ calcd for C₂₉H₃₈N₄NaO₇S 609.2359 found 609.2362.

(*R*)-*tert*-butyl-4-(((2*R*,3*S*)-3-(*tert*-butoxycarbonylamino)-2-hydroxy-4-phenylbutylamino)methyl)-2,2-dimethyloxazolidine-3-carboxylate 22

To a solution of the (*R*)-*tert*-butyl 4-(azidomethyl)-2,2-dimethyloxazolidine-3-carboxylate ²¹ (411 mg, 1.60 mmol) in MeOH (10 mL) was added Pd/C (40 mg). This mixture was stirred at 23 °C under a hydrogen filled balloon for 2 h, then filtered over Celite, and the filter cake was washed with MeOH. Solvent was removed under reduced pressure and the resultant residue was redissolved in ⁱPrOH, followed by addition of *tert*-butyl-[*S*-(*R*,*R*)]-(*-*)-(1-oxiranyl-2-phenylethyl) carbamate ⁷ (121 mg, 0.46 mmol). The mixture was allowed to stir at 65°C for 24 h, solvent was then removed under reduced pressure. Flash chromatography purification (2% MeOH in CHCl₃ as the eluent) yielded the (*R*)-amine ²² (93 mg, 41%) as a yellow oil. *R*_f = 0.32 (10% MeOH in CHCl₃); ¹H-NMR (500 MHz, CDCl₃) δ 1.35 (s, 18H), 1.48 (s, 24H), 1.54 (s, 3H), 1.58 (s, 3H), 2.67–2.92 (m, 10), 2.95 (d, 1H, *J* = 4.1 Hz), 2.98 (d, 1H, *J* = 4.1 Hz), 3.47 (brs, 2H), 3.78 (brs, 3H), 3.89–4.02 (m, 5H), 4.61 (brs, 1H), 4.68 (brs, 1H), 7.18–7.22 (m, 6H), 7.27–7.30 (m, 4H); ¹³C-NMR (125 MHz, CDCl₃) δ 22.94, 24.27, 26.68, 27.46, 28.17, 28.33, 36.43, 51.45, 56.95, 65.97, 70.74, 70.90, 79.32, 79.45, 79.66, 80.35, 93.46, 93.77, 126.23, 128.30, 129.41, 137.55, 137.77, 152.62, 155.86; LRMS-ESI (*m/z*) [M + Na]⁺ 516.

(*R*)-*tert*-butyl-4-((*N*-((2*R*,3*S*)-3-(*tert*-butoxycarbonylamino)-2-hydroxy-4-phenylbutyl)-4-methoxyphenylsulfonamido)methyl)-2,2-dimethyloxazolidine-3-carboxylate 23

To a stirred solution of amine ²² (37 mg, 0.07 mmol) in CH₂Cl₂ (4 mL) and aqueous sat. NaHCO₃ (4 mL) was added 4-methoxybenzenesulfonyl chloride (34 mg, 0.16 mmol). This reaction mixture was stirred for 12 h followed by extraction of the aqueous layer with CH₂Cl₂; the combined organic extracts were dried over anhydrous Na₂SO₄. Removal of solvent under reduced pressure, followed by flash chromatography purification (30% EtOAc in Hexane as the eluent) provided compound ²³ (40 mg, 80%) as a white solid. *R*_f = 0.37 (40% EtOAc in Hex); ¹H-NMR (500 MHz, CDCl₃) δ 1.34 (s, 9H), 1.48 (s, 9H), 1.60 (s, 6H), 2.84–

2.89 (m, 1H), 2.95–3.06 (m, 2H), 3.15 (dd, 1H, $J = 6.5, 13.0$ Hz), 3.29 (d, 1H, $J = 9.7$ Hz), 3.41 (d, 1H, $J = 15.1$ Hz), 3.77 (brs, 2H), 3.85 (s, 3H), 3.90–3.94 (m, 1H), 3.99 (d, 1H, $J = 9.0$ Hz), 4.2 (brs, 1H), 4.73 (brs, 0.5H), 4.98 (brs, 0.5H), 6.95 (d, 2H, $J = 8.0$ Hz), 7.19–7.24 (m, 3H), 7.25–7.28 (m, 2H), 7.69 (d, 2H, $J = 8.9$ Hz); ^{13}C -NMR (125 MHz, CDCl_3) δ 24.07, 27.42, 28.21, 28.31, 36.03, 53.30, 54.01, 55.48, 55.89, 56.38, 65.69, 72.47, 79.06, 80.97, 93.95, 114.29, 126.09, 128.15, 129.31, 129.61, 129.92, 137.80, 152.82, 155.43, 162.91; LRMS-ESI (m/z) $[\text{M} + \text{Na}]^+$ 686.

(*R*)-*tert*-butyl-4-((*N*-((2*R*,3*S*)-3-(*tert*-butoxycarbonylamino)-2-hydroxy-4-phenylbutyl)-4-nitrophenylsulfonamido)methyl)-2,2-dimethyloxazolidine-3-carboxylate 24

To a stirred solution of amine ²² (41 mg, 0.08 mmol) in CH_2Cl_2 (4 mL) and aqueous sat. NaHCO_3 (4 mL) was added 4-nitrobenzenesulfonyl chloride (37 mg, 0.16 mmol). This reaction mixture was stirred for 12 h followed by extraction of the aqueous layer with CH_2Cl_2 ; the combined organic extracts were dried over anhydrous Na_2SO_4 . Removal of solvent under reduced pressure, followed by flash chromatography purification (30% EtOAc in Hexane as the eluent) provided (*R*)-nitrosulfone ²⁴ (51 mg, 92%) as a white solid. $R_f = 0.52$ (40% EtOAc in Hex); ^1H -NMR (500 MHz, CDCl_3) δ 1.35 (s, 18H), 1.49 (s, 24H), 1.62 (s, 6H), 3.43 (bd, 2H, $J = 9.6$ Hz), 3.55 (d, 2H, $J = 15.1$ Hz), 3.75 (brs, 4H), 3.95 (s, 4H), 4.28 (brs, 2H), 4.63 (d, 2H, $J = 7.8$ Hz), 5.16 (brs, 2H), 7.19–7.23 (m, 6H), 7.25–7.31 (m, 4H), 7.92 (d, 4H, $J = 8.8$ Hz), 8.30 (d, 4H, $J = 8.05$ Hz); ^{13}C -NMR (125 MHz, CDCl_3) δ 24.01, 27.39, 28.19, 28.29, 35.88, 53.75, 54.06, 55.55, 55.98, 65.61, 72.27, 79.47, 81.33, 94.16, 124.34, 126.31, 128.29, 128.40, 129.50, 137.46, 144.32, 149.95, 153.05, 155.53; LRMS-ESI (m/z) $[\text{M} + \text{Na}]^+$ 701.

***tert*-butyl-(2*S*,3*R*)-3-hydroxy-4-(4-methoxy-*N*-(((*R*)-2-oxooxazolidin-4-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-ylcarbamate 25**

To (*R*)-toluenesulfonate ²³ (32 mg, 0.05 mmol) in methanol (3 mL) was added *p*-toluenesulfonic acid monohydrate (2 mg), and the reaction mixture was allowed to stir at 23 °C. After 12 h reaction reached maximum conversion and did not move any further. It was quenched with sat. aqueous NaHCO_3 , extracted with EtOAc, washed with water and brine, and dried over Na_2SO_4 . Flash chromatography purification (30–50% EtOAc in Hexane) yielded 7 mg (21%) of starting material and 18 mg (58%) of the expected primary alcohol.

The above alcohol (17 mg, 0.0278 mmol), in CH_2Cl_2 (2 mL) and Et_3N (16 μL , 0.11 mmol), was cooled to -10 °C and treated with methanesulfonyl chloride (2.5 μL , 0.03 mmol). The reaction mixture was stirred at 0 °C for 2 h, followed by removal of solvent via rotavap and pump. The resultant residue was redissolved in chloroform (2 mL), treated with DIPEA (19 μL , 0.11 mmol), refluxed for 8 h, and concentrated under reduced pressure. Flash chromatography purification (1% MeOH in CHCl_3 as the eluent) afforded (*R*)-oxazolidinone ²⁵ (12 mg, 78%) as a white solid. $R_f = 0.33$ (5% MeOH in CHCl_3); ^1H -NMR (500 MHz, CDCl_3) δ 1.32 (s, 9H), 2.77 (dd, 1H, $J = 8.4, 13.9$ Hz), 2.93–3.03 (m, 3H), 3.07 (d, 1H, $J = 14.5$ Hz), 3.13 (dd, 1H, $J = 9.3, 14.1$ Hz), 3.69–3.79 (m, 2H), 3.85 (s, 3H), 3.97 (dd, 1H, $J = 5.2, 8.3$ Hz), 4.23–4.29 (m, 1H), 4.43 (t, 1H, $J = 8.7$ Hz), 6.94 (d, 2H, $J = 8.8$ Hz), 7.18–7.24 (m, 3H), 7.26–7.30 (m, 2H), 7.64 (d, 2H, $J = 8.6$ Hz); ^{13}C -NMR (125 MHz, CDCl_3) δ 28.12, 35.97, 52.02, 54.16, 54.60, 54.73, 55.55, 67.47, 71.97, 79.70, 114.42, 126.37, 128.42, 128.48, 129.30, 129.44, 137.52, 155.99, 159.44, 163.22; LRMS-ESI (m/z) $[\text{M} + \text{Na}]^+$ 572.

***tert*-butyl-(2*S*,3*R*)-4-(4-amino-*N*-(((*R*)-2-oxooxazolidin-4-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-ylcarbamate 26**

To (*R*)-nitrosulfone ²⁴ (42 mg, 0.06 mmol) in methanol (4 mL) was added *p*-toluenesulfonic acid monohydrate (2 mg), and the reaction mixture was allowed to stir at 23 °C. After 36 h reaction appeared to have moved only 75% to completion, and formation of free amines was evident on TLC. At this point it was quenched with sat. aqueous NaHCO_3 , extracted with

EtOAc, washed with water and brine, and dried over Na_2SO_4 . Flash chromatography purification (30–50% EtOAc in Hexane) yielded 7 mg (17%) of starting material and 24 mg (55%) of the expected primary alcohol.

The above alcohol (23 mg, 0.03 mmol), in CH_2Cl_2 (3 mL) and Et_3N (20 μL , 0.14 mmol), was cooled to -10°C and treated with methanesulfonyl chloride (53 μL , 0.04 mmol). The reaction mixture was stirred at 0°C for 2 h, followed by removal of solvent via rotavap and pump. The resultant residue was redissolved in chloroform (3 mL), treated with DIPEA (25 μL , 0.14 mmol), and refluxed for 12 h; product crashed out of the chloroform and was found to be only slightly soluble in methanol. Solvent was removed under reduced pressure and the crude material was treated with methanol (3 mL) and Pd/C (3 mg). The mixture was stirred at 23°C under a H_2 filled balloon for 2 h, then filtered over Celite and the filter cake was washed with MeOH. Removal of solvent, followed by flash chromatography purification (3% MeOH in CHCl_3 as the eluent) provided (*R*)-oxazolidinone ²⁶ (13 mg, 68%) as a white solid. $R_f = 0.19$ (5% MeOH in CHCl_3); $^1\text{H-NMR}$ (500 MHz, CDCl_3) δ 1.31 (s, 9H), 2.73 (dd, 1H, $J = 8.7, 13.6$ Hz), 2.88 (dd, 1H, $J = 9.6, 14.5$ Hz), 2.91–2.94 (m, 1H), 2.98 (dd, 1H, $J = 4.2, 13.9$ Hz), 3.10 (dd, 2H, $J = 9.2, 14.0$ Hz), 3.70–3.81 (m, 2H), 3.96 (dd, 1H, $J = 5.2, 8.4$ Hz), 4.21–4.24 (m, 1H), 4.40 (t, 1H, $J = 8.7$ Hz), 4.92 (d, 1H, $J = 8.5$ Hz), 6.63 (d, 2H, $J = 8.7$ Hz), 7.17–7.21 (m, 3H), 7.25–7.28 (m, 2H), 7.46 (d, 2H, $J = 8.4$ Hz); $^{13}\text{C-NMR}$ (125 MHz, CD_3OD) δ 28.13, 29.57, 35.99, 51.91, 52.00, 54.09, 54.73, 67.53, 72.12, 79.63, 113.98, 124.40, 126.31, 128.36, 129.32, 129.45, 137.62, 155.12, 156.01, 159.52; LRMS-ESI (m/z) [$\text{M} + \text{Na}$]⁺ 557.

(*tert*-butyl(2*S*,3*R*)-3-hydroxy-4-(4-methoxy-*N*-(((*S*)-2-oxooxazolidin-4-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-yl)carbamate 27

To a solution of the (*S*)-*tert*-butyl 4-(azidomethyl)-2,2-dimethyloxazolidine-3-carboxylate ent-²¹ (840 mg, 3.32 mmol) in MeOH (20 mL) was added Pd/C (80 mg). This mixture was stirred at 23°C under a hydrogen-filled balloon for 2 h, then filtered over Celite, and the filter cake was washed with MeOH. Solvent was removed under reduced pressure and the resultant residue was redissolved in i PrOH, followed by addition of *tert*-butyl-[*S*-(*R,R*)]-($-$)-(1-oxiranyl-2-phenylethyl) carbamate ⁷ (218 mg, 0.83 mmol). The mixture was allowed to stir at 65°C for 24 h, solvent was then removed under reduced pressure. Flash chromatography purification (2% MeOH in CHCl_3 as the eluent) yielded the corresponding (*S*)-amine (²²*S*) (187 mg, 46%) as an amorphous solid. $R_f = 0.28$ (10% MeOH in CHCl_3); $^1\text{H-NMR}$ (500 MHz, CDCl_3) δ 1.35 (s, 18H), 1.48 (s, 24H), 1.54 (s, 3H), 1.58 (s, 3H), 2.64–2.91 (m, 10), 2.95–3.03 (m, 2H), 3.44 (brs, 1H), 3.50 (brs, 1H), 3.77 (brs, 3H), 3.87–4.00 (m, 5H), 4.61 (brs, 1H), 4.64 (brs, 1H), 7.18–7.22 (m, 6H), 7.27–7.30 (m, 4H); $^{13}\text{C-NMR}$ (125 MHz, CDCl_3) δ 22.96, 24.27, 26.69, 27.47, 28.16, 28.32, 36.48, 51.47, 56.73, 56.87, 65.95, 70.68, 79.35, 79.65, 80.34, 93.45, 93.76, 126.21, 126.30, 128.30, 129.39, 137.61, 137.79, 152.58, 155.88; LRMS-ESI (m/z) [$\text{M} + \text{Na}$]⁺ 576.

To a stirred solution of above *S*-amine (²²*S*) (85 mg, 0.17 mmol) in CH_2Cl_2 (6 mL) and aqueous sat. NaHCO_3 (6 mL) was added 4-methoxybenzenesulfonyl chloride (71 mg, 0.34 mmol). This reaction mixture was stirred for 12 h followed by extraction of the aqueous layer with CH_2Cl_2 ; the combined organic extracts were dried over anhydrous Na_2SO_4 . Removal of solvent under reduced pressure, followed by flash chromatography purification (30% EtOAc in Hexane as the eluent) provided *S*-toluenesulfonate (²³*S*) (107 mg, 93%) as a white solid. $R_f = 0.40$ (40% EtOAc in Hexane); $^1\text{H-NMR}$ (500 MHz, CDCl_3) δ 1.34 (s, 18H), 1.46 (s, 24H), 1.58 (s, 6H), 2.82–2.87 (m, 2H), 2.96–3.13 (m, 4H), 3.18–3.24 (m, 3H), 3.32 (brs, 1H), 3.75 (brs, 1H), 3.80 (brs, 1H), 3.84 (s, 6H), 3.89–3.93 (4H), 4.07 (d, 2H, $J = 9.2$ Hz), 4.16–4.19 (m, 2H), 4.65 (brs, 1H), 4.87 (d, 1H, $J = 7.31$ Hz), 6.93–6.97 (m, 4H), 7.18–7.23 (6H), 7.26–7.28 (m, 4H), 7.64–7.71 (4H); $^{13}\text{C-NMR}$ (125 MHz, CDCl_3) δ 23.98, 27.38, 28.18, 28.28, 35.68,

52.47, 54.35, 55.24, 55.47, 56.88, 65.66, 71.86, 79.19, 80.93, 93.84, 114.22, 126.13, 128.24, 129.39, 129.47, 129.64, 137.97, 152.69, 155.64, 162.85; LRMS-ESI (m/z) $[M + Na]^+$ 686.

To above *S*-toluenesulfonate (23*S*) (96 mg, 0.14 mmol) in methanol (5 mL) was added *p*-toluenesulfonic acid monohydrate (4.5 mg, 0.02 mmol), and the reaction mixture was allowed to stir at 23 °C. After 8 h reaction reached maximum conversion and did not move any further. It was quenched with sat. aqueous $NaHCO_3$, extracted with EtOAc, washed with water and brine, and dried over Na_2SO_4 . Flash chromatography purification (30–50% EtOAc in Hexane) yielded 13 mg (13%) of starting material and 59 mg (65%) of the expected primary alcohol.

The above alcohol (58 mg, 0.09 mmol), in CH_2Cl_2 (4 mL) and Et_3N (33 μ L, 0.23 mmol), was cooled to –10 °C and treated with methanesulfonyl chloride (7.9 μ L, 0.1 mmol). The reaction mixture was stirred at 0 °C for 2 h, followed by removal of solvent via rotavap and pump. The resultant residue was redissolved in chloroform (4 mL), treated with DIPEA (65 μ L, 0.37 mmol), refluxed for 12 h, and concentrated under reduced pressure. Flash chromatography purification (2% MeOH in $CHCl_3$ as the eluent) afforded *S*-oxazolidinone²⁷ (44 mg, 85%) as a white solid. R_f = 0.27 (5% MeOH in $CHCl_3$); ^1H -NMR (500 MHz, CD_3OD) δ 1.27 (s, 9H), 2.52 (dd, 1H, J = 10.95, 13.7 Hz), 2.67 (dd, 1H, J = 8.9, 14.9 Hz), 2.83 (dd, 1H, J = 6.1, 14.0 Hz), 3.10 (dd, 1H, J = 3.5, 13.8 Hz), 3.47–3.52 (m, 2H), 3.58–3.63 (m, 1H), 3.80–3.83 (m, 1H), 3.85 (s, 3H), 4.23–4.29 (m, 1H), 4.31 (dd, 1H, J = 4.8, 8.9 Hz), 4.47 (t, 1H, J = 8.6 Hz), 7.06 (d, 2H, J = 8.93 Hz), 7.12–7.16 (m, 1H), 7.20–7.24 (m, 4H), 7.75 (d, 2H, J = 8.9 Hz); ^{13}C -NMR (125 MHz, CD_3OD) δ 27.19, 35.63, 52.00, 54.38, 54.613, 54.70, 55.17, 68.04, 73.35, 78.52, 114.05, 125.57, 127.64, 128.92, 129.08, 129.31, 138.67, 156.55, 160.46, 163.36; LRMS-ESI (m/z) $[M + Na]^+$ 572.

***tert*-butyl-(2*S*,3*R*)-4-(4-amino-*N*-(((*S*)-2-oxooxazolidin-4-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-ylcarbamate 28**

To a stirred solution of *S*-amine (22*S*) (87 mg, 0.17 mmol) in CH_2Cl_2 (6 mL) and aqueous sat. $NaHCO_3$ (6 mL) was added 4-nitrobenzenesulfonyl chloride (78 mg, 0.35 mmol). This reaction mixture was stirred for 12 h followed by extraction of the aqueous layer with CH_2Cl_2 ; the combined organic extracts were dried over anhydrous Na_2SO_4 . Removal of solvent under reduced pressure, followed by flash chromatography purification (30% EtOAc in Hexane as the eluent) provided *S*-nitrosulfone (24*S*) (102 mg, 85%) as a yellow solid. R_f = 0.54 (40% EtOAc in Hex); ^1H -NMR (500 MHz, $CDCl_3$) δ 1.34 (s, 18H), 1.47 (s, 24H), 1.59 (s, 6H), 2.81 (dd, 2H, J = 8.0, 12.9 Hz), 3.03 (bd, 2H, J = 9.75 Hz), 3.16–3.36 (m, 8H), 3.82 (brs, 2H), 3.91 (dd, 4H, J = 5.15, 8.7 Hz), 4.00 (d, 2H, J = 6.2 Hz), 4.20 (q, 2H, J = 5.6, 5.7 Hz), 4.80 (brs, 1H), 4.86 (d, 1H, J = 7.2 Hz), 7.20–7.24 (m, 6H), 7.27–7.30 (m, 4H), 7.88 (d, 4H, J = 8.4 Hz), 8.29 (d, 4H, J = 8.4 Hz); ^{13}C -NMR (125 MHz, $CDCl_3$) δ 23.93, 27.38, 28.15, 28.27, 35.82, 52.35, 54.57, 54.97, 56.65, 65.50, 71.67, 79.57, 81.18, 94.02, 124.21, 126.31, 128.40, 128.40, 128.51, 129.34, 137.79, 144.05, 149.93, 152.77, 155.82, 171.05; LRMS-ESI (m/z) $[M + Na]^+$ 701.

To above *S*-nitrosulfone (24*S*) (92 mg, 0.13 mmol) in methanol (5 mL) was added *p*-toluenesulfonic acid monohydrate (4 mg, 0.02 mmol), and the reaction mixture was allowed to stir at 23 °C. After 18 h reaction appeared to have moved only 50% to completion, and formation of free amines was evident on TLC. At this point it was quenched with sat. aqueous $NaHCO_3$, extracted with EtOAc, washed with water and brine, and dried over Na_2SO_4 . Flash chromatography purification (30–50% EtOAc in Hexane) yielded 32 mg (35%) of starting material and 44 mg (51%) of the expected primary alcohol.

The above alcohol (43 mg, 0.06 mmol), in CH_2Cl_2 (3 mL) and Et_3N (24 μ L, 0.17 mmol), was cooled to –10 °C and treated with methanesulfonyl chloride (5.5 μ L, 0.07 mmol). The reaction mixture was stirred at 0 °C for 2 h, followed by removal of solvent via rotavap and pump. The

resultant residue was redissolved in chloroform (2 mL), treated with DIPEA (29 μ L, 0.17 mmol), and refluxed for 12 h; product crashed out of the chloroform and was found to be only slightly soluble in methanol. Solvent was removed under reduced pressure and the crude material was treated with methanol (4 mL) and Pd/C (5 mg). The mixture was stirred at 23 °C under a hydrogen filled balloon for 3 h, then filtered over Celite and the filter cake was washed with MeOH. Removal of solvent, followed by flash chromatography purification (3% MeOH in CHCl_3 as the eluent) provided *S*-oxazolidinone ²⁸ (21 mg, 58%) as a white solid. R_f = 0.21 (5% MeOH in CHCl_3); $^1\text{H-NMR}$ (500 MHz, 1:1 $\text{CD}_3\text{OD}/\text{CDCl}_3$) δ 1.24 (s, 9H), 2.54 (d, 1H), 2.56 (dd, 1H, J = 8.7, 14.9 Hz), 2.70 (dd, 1H, J = 7.6, 14.3 Hz), 2.97 (dd, 1H, J = 4.0, 13.9 Hz), 3.32 (dd, 1H, J = 5.0, 14.3 Hz), 3.38 (dd, 1H, J = 3.1, 14.8 Hz), 3.64–3.70 (m, 1H), 3.78–3.83 (m, 1H) 4.05 (dd, 1H, J = 5.3, 8.8 Hz), 4.16–4.20 (m, 1H), 4.39 (t, 1H, J = 8.7 Hz), 6.59 (d, 2H, J = 8.7 Hz), 7.09–7.15 (m, 3H), 7.18–7.21 (m, 2H), 7.40 (d, 2H, J = 8.7 Hz); $^{13}\text{C-NMR}$ (125 MHz, 1:1 $\text{CD}_3\text{OD}/\text{CDCl}_3$) δ 31.88, 33.40, 39.42, 53.38, 56.90, 58.52, 58.84, 59.77, 71.89, 77.48, 83.34, 117.62, 127.44, 130.02, 132.05, 133.11, 133.20, 141.93, 156.09, 160.19, 164.05; LRMS-ESI (m/z) [$\text{M} + \text{Na}$]⁺ 557.

(3*R*,3*aS*,6*aR*)-hexahydrofuro[2,3-*b*]furan-3-yl-(2*S*,3*R*)-3-hydroxy-4-(4-methoxy-*N*-(*(R)*-2-oxooxazolidin-4-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-ylcarbamate 29

A solution of *S*-aminosulfone ²⁵ (10.9 mg, 0.02 mmol) in 30% trifluoroacetic acid (in CH_2Cl_2 , 2 mL) was stirred at 23 °C for 40 min, then concentrated under reduced pressure. The residue was redissolved in CH_2Cl_2 (2 mL), treated with Et_3N (30 μ L, 0.21 mmol), followed by carbonate ¹⁵ (6.3 mg, 0.02 mmol) and stirred at 23 °C for 12 h. The reaction mixture was then concentrated under reduced pressure and the residue was purified by flash chromatography (3% MeOH in CHCl_3 as the eluent) to give inhibitor ²⁹ (10 mg, 85%) as a white solid, R_f = 0.54 (10% MeOH in CHCl_3); $^1\text{H-NMR}$ (500 MHz, CDCl_3) δ 1.39 (dd, 1H, J = 5.5, 13.1 Hz), 1.56–1.63 (m, 1H), 2.67 (dd, 1H, J = 10.0, 14.0 Hz), 2.85–2.90 (m, 1H), 2.95–3.01 (m, 2H), 3.10 (dd, 1H, J = 4.1, 14.1 Hz), 3.20 (d, 1H, J = 12.8 Hz), 3.28 (dd, 1H, J = 9.2, 14.1 Hz), 3.63–3.72 (m, 1H), 3.73 (dd, 1H, J = 5.2, 9.8 Hz), 3.79 (dt, 1H, J = 1.6, 8.2 Hz), 3.86 (s, 3H), 3.88–3.95 (m, 3H), 4.04 (dd, 1H, J = 5.0, 8.8 Hz), 4.25 (brs, 1H), 4.28–4.34 (m, 1H), 4.47 (t, 1H, J = 8.7 Hz), 4.99 (q, 1H, J = 5.6, 13.4 Hz), 5.38 (d, 1H, J = 9.8 Hz), 5.63 (d, 1H, J = 5.2 Hz), 6.87 (s, 1H), 6.97 (d, 2H, J = 8.9 Hz), 7.16–7.22 (m, 3H), 7.24–7.28 (m, 2H), 7.69 (d, 2H, J = 8.8 Hz); $^{13}\text{C-NMR}$ (125 MHz, CDCl_3) δ 25.81, 35.92, 45.47, 51.96, 54.18, 55.09, 55.30, 55.63, 67.61, 69.62, 71.13, 72.58, 73.42, 109.29, 114.54, 126.44, 128.43, 128.47, 129.23, 129.44, 137.58, 155.64, 159.60, 163.35; LRMS-ESI (m/z) [$\text{M} + \text{Na}$]⁺ 628.2; HRMS-ESI (m/z) [$\text{M} + \text{Na}$]⁺ calcd for $\text{C}_{28}\text{H}_{35}\text{N}_3\text{O}_{10}\text{S}$ 628.1941 found 628.1937.

(3*R*,3*aS*,6*aR*)-hexahydrofuro[2,3-*b*]furan-3-yl-(2*S*,3*R*)-4-(4-amino-*N*-(*(R)*-2-oxooxazolidin-4-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-ylcarbamate 30

A solution of the free amine ²⁶ (12 mg, 0.02 mmol) in 30% trifluoroacetic acid in CH_2Cl_2 (2 mL) was stirred at 23 °C for 40 min, then concentrated under reduced pressure. The residue was redissolved in CH_2Cl_2 (2 mL), treated with carbonate ¹⁵ (7.1 mg, 0.02 mmol) and Et_3N (40 μ L, 0.28 mmol), and stirred at 23 °C for 12 h at 40 °C. The reaction mixture was then concentrated under reduced pressure and the residue was purified by flash chromatography (5% MeOH in CHCl_3 as the eluent) to give inhibitor ³⁰ (11 mg, 85%) as a white solid, R_f = 0.26 (10% MeOH in CHCl_3); $^1\text{H-NMR}$ (500 MHz, CDCl_3) δ 1.38 (dd, 1H, J = 5.6, 13.2 Hz), 1.54–1.63 (m, 1H), 2.61 (dd, 1H, J = 9.5, 13.9 Hz), 2.83–2.91 (m, 2H), 3.10 (d, 2H, J = 13.3 Hz), 3.18 (dd, 1H, J = 9.6, 13.9 Hz), 3.62–3.70 (m, 2H), 3.78–3.84 (m, 3H), 3.88 (dd, 1H, J = 6.0, 9.8 Hz), 3.97 (dd, 1H, J = 5.0, 8.9 Hz), 4.21–4.27 (m, 1H), 4.41 (t, 1H, J = 8.7 Hz), 4.93 (q, 1H, J = 5.7, 13.6 Hz), 5.60 (d, 1H, J = 5.25 Hz), 5.69 (d, 1H, J = 8.8 Hz), 6.64 (d, 2H, J = 8.7 Hz), 7.16–7.19 (m, 3H), 7.21–7.26 (m, 2H), 7.45 (d, 2H, J = 8.7 Hz); $^{13}\text{C-NMR}$ (125 MHz, CDCl_3) δ 25.73, 29.56, 36.12, 45.37, 51.74, 54.14, 54.93, 55.18, 67.50, 69.57, 70.87, 71.05, 73.21, 109.26, 113.92, 123.90, 126.33, 128.33, 129.21, 129.45, 137.73, 151.44, 155.60, 159.65;

LRMS-ESI (m/z) $[M+Na]^+$ 613.2; HRMS-ESI (m/z) $[M+Na]^+$ calcd for $C_{27}H_{34}N_4O_9S$ 613.1944 found 613.1939.

(3*R*,3*aS*,6*aR*)-hexahydrofuro[2,3-*b*]furan-3-yl(2*S*,3*R*)-3-hydroxy-4-(4-methoxy-*N*((*S*)-2-oxooxazolidin-4-yl)methyl)phenylsulfonamido)-1-phenylbutan-2-ylcarbamate 31

A solution of *S*-aminosulfone ²⁷ (16.5 mg, 0.03 mmol) in 30% trifluoroacetic acid (in CH_2Cl_2 , 3 mL) was stirred at 23 °C for 40 min, then concentrated under reduced pressure. The residue was redissolved in CH_2Cl_2 (3 mL), treated with Et_3N (41 μ L, 0.29 mmol), followed by carbonate 15 (10 mg, 0.03 mmol), and stirred at 23 °C for 12 h. The reaction mixture was then concentrated under reduced pressure and the residue was purified by flash chromatography (3% MeOH in $CHCl_3$ as the eluent) to give inhibitor ³¹ (16 mg, 90%) as a white solid, R_f = 0.5 (10% MeOH in $CHCl_3$); ^1H -NMR (500 MHz, $CDCl_3$) δ 1.32 (dd, 1H, J = 5.4, 13.2 Hz), 1.53–1.62 (m, 1H), 2.62 (dd, 1H, J = 11.0, 13.8 Hz), 2.71 (dd, 1H, J = 7.25, 14.6 Hz), 2.84–2.89 (m, 2H), 3.10 (dd, 1H, J = 3.6, 14.0 Hz), 3.61–3.67 (m, 1H), 3.74–3.82 (m, 2H), 3.86 (s, 3H), 3.87 (dd, 2H, J = 5.8, 10.1 Hz), 3.94–3.98 (m, 1H), 4.00–4.05 (m, 2H), 4.27–4.33 (m, 1H), 4.45 (t, 1H, J = 8.8 Hz), 4.99 (q, 1H, J = 5.3, 7.7 Hz), 5.60 (d, 1H, J = 5.3 Hz), 5.68 (d, 1H, J = 9.2 Hz), 6.98 (d, 2H, J = 8.8 Hz), 7.15–7.20 (m, 1H), 7.22–7.27 (m, 4H), 7.69 (d, 2H, J = 8.8 Hz); ^{13}C -NMR (125 MHz, $CDCl_3$) δ 25.78, 43.46, 45.57, 53.23, 54.68, 54.96, 55.62, 56.58, 57.59, 69.60, 71.20, 73.33, 73.58, 109, 31, 114.54, 126.32, 128.32, 129.23, 129.45, 137.85, 155.94, 159.61, 163.38; LRMS-ESI (m/z) $[M+Na]^+$ 628.2; HRMS-ESI (m/z) $[M+Na]^+$ calcd for $C_{28}H_{35}N_3O_{10}S$ 628.1941 found 628.1943.

(2*R*,3*aS*,6*aR*)-hexahydrofuro[2,3-*b*]furan-3-yl-(2*S*,3*R*)-4-(4-amino-*N*((*S*)-2-oxooxazolidin-4-yl)methyl)phenylsulfonamido)-3-hydroxy-1-phenylbutan-2-ylcarbamate 32

A solution of the free amine ²⁹ (15 mg, 0.03 mmol) in 30% trifluoroacetic acid (in CH_2Cl_2 , 3 mL) was stirred at 23 °C for 40 min, then concentrated under reduced pressure. The residue was redissolved in CH_2Cl_2 (3 mL), charged with Et_3N (60 μ L, 0.42 mmol), followed by carbonate 15 (8.9 mg, 0.03 mmol), and stirred at 23 °C for 12 h at 40 °C. The reaction mixture was then concentrated under reduced pressure and the residue was purified by flash chromatography (5% MeOH in $CHCl_3$ as the eluent) to give inhibitor ³² (13 mg, 80%) as a white solid, R_f = 0.32 (10% MeOH in $CHCl_3$); ^1H -NMR (500 MHz, $CDCl_3$) δ 1.33 (dd, 1H, J = 8.5 Hz), 1.53–1.62 (m, 1H), 2.59 (dd, 1H, J = 10.5, 13.9 Hz), 2.66 (dd, 1H, J = 7.4, 14.7 Hz), 2.77 (dd, 1H, J = 8.9, 14.5 Hz), 2.82–2.89 (m, 1H), 3.10 (dd, 1H, J = 3.4, 14.0 Hz), 3.36 (dd, 1H, J = 3.5, 14.3 Hz), 3.44 (dd, 1H, J = 4.5, 14.6 Hz), 3.62–3.67 (m, 1H), 3.72 (dd, 1H, J = 5.0, 9.9 Hz), 3.77 (dt, 1H, J = 1.8, 8.2 Hz), 3.87 (dd, 1H, J = 5.8, 9.9 Hz), 3.92–3.98 (m, 2H), 3.99 (dd, 1H, J = 5.0, 9.0 Hz), 4.15–4.29 (m, 1H), 4.43 (t, 1H, J = 8.8 Hz), 4.97 (q, 1H, J = 5.4, 13.2 Hz), 5.60 (d, 1H, J = 5.3 Hz), 6.66 (d, 2H, J = 8.7 Hz), 7.15–7.18 (m, 1H), 7.20–7.25 (m, 4H), 7.49 (d, 2H, J = 8.6 Hz); ^{13}C -NMR (125 MHz, $CDCl_3$) δ 25.76, 29.56, 34.75, 45.51, 53.33, 54.67, 55.03, 56.62, 67.57, 69.60, 71.18, 73.47, 109.31, 114.02, 124.27, 126.30, 128.30, 129.22, 129.42, 137.86, 151.29, 155.83, 159.65; LRMS-ESI (m/z) $[M+Na]^+$ 613.2; HRMS-ESI (m/z) $[M+Na]^+$ calcd for $C_{27}H_{34}N_4O_9S$ 613.1944 found 613.1938.

Determination of X-ray structure of 19b-bound HIV protease (WT)

The stabilized HIV-1 protease with the substitutions of Q7K, L33I, L63I, C67A and C95A that reduce autoproteolysis and aggregation²⁷ was expressed and purified as described²⁸. These mutations do not alter the inhibitor binding site and the stabilized protease has kinetic parameters and stability indistinguishable from those of the unsubstituted enzyme^{27,28}. Inhibitor **19b** was dissolved in dimethylsulfoxide (DMSO). Crystals were grown by the hanging drop vapor diffusion method using 1:5 molar ratio of protease (at 3.9 mg/mL) to inhibitor. The reservoir contained 0.1 M citrate phosphate buffer, pH 5.0, 0.35 M NaCl and 4% DMSO. Crystals were mounted on a nylon loop and flash-frozen in liquid nitrogen with a

cryoprotectant of 30% (v/v) glycerol. X-ray diffraction data were collected on the SER-CAT beamLine of the Advanced Photon Source, Argonne National Laboratory. Diffraction data were processed using HKL2000 to 1.29 Å resolution.²⁸ Data were reduced in space group $P2_12_12$ with unit cell dimensions of $a = 58.11$ Å, $b = 86.42$ Å, $c = 45.97$ Å with one dimer in the asymmetric unit. The structure was solved by molecular replacement using the CCP4;^{29, 30} using WT/GRL0255 complex (PDB 3DJK) as a starting model.²⁶ The structure was refined using SHELX97³¹ and refitted manually using the molecular graphics program COOT³³. Alternate conformations were modeled for the protease residues where observed in the electron density maps. Anisotropic atomic displacement parameters (B-factors) were refined for all atoms including solvent molecules. Hydrogen atoms were automatically added by SHELXL in the last round of the refinement. The identity of ions and other solvent molecules from the crystallization conditions was deduced from the shape and peak height of the $2F_o - F_c$ and $F_o - F_c$ electron density, the hydrogen bond interactions and interatomic distances. The solvent molecules were one sodium ion, two chloride ions, one glycerol molecule and 207 water (including partial occupancy sites). The final R was 14.1 % for working set and R_{free} was 18.2 % for all data between 10 and 1.29 Å resolution. The rmsd values from ideal bonds and angle distances were 0.013 Å and 0.031 Å, respectively. The average B -factor was 15.9 and 22.7 Å² for protease main chain and side chain atoms, respectively, 14.1 Å² for inhibitor atoms and 32.1 Å² for water atoms. The crystallographic coordinates and structure factors have been deposited in the Protein Databank (PDB) with access code 3H5B.³⁴

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Abbreviations

HIV, human immunodeficiency virus
bis-THF, bis-tetrahydrofuran
Cp-THF, cyclopentanyltetrahydrofuran
PI, protease inhibitor
HAART, highly active antiretroviral therapy
APV, amprenavir
DRV, darunavir
SQV, saquinavir
IDV, indinavir
LPV, lopinavir
RTV, ritonavir

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Figure 1.
Structures of inhibitors **1–4**.

Figure 2.
A stereoview of the major conformation of the X-ray structure of inhibitor **19b**-bound HIV-1 protease.

Figure 3. Protease interactions with the two alternate conformations of the inhibitor pyrrolidine ring. The inhibitor is in green bonds with thick bonds for the major and thin bonds for the minor conformations of the pyrrolidine ring. Hydrogen bonds are shown in dotted lines. Distances between donor and acceptor atoms are shown in Å.

Scheme 1.
Synthesis of lactam containing sulfonamide isosteres

Scheme 2.
Synthesis of lactam containing PIs

Scheme 3.
 Synthesis of sulfonamide isosteres with P1'-oxazolidinone

Scheme 4.
Synthesis of oxazolidinone-derived PIs.

Table 1

Enzymatic inhibitory activity of lactam and oxazolidinone containing inhibitors.

Entry	Inhibitor
1	 17a
2	 17b
3	 18a

Entry

Inhibitor

4

18b

5

19a

Entry

Inhibitor

6

7

8

9

10

11

Entry	Inhibitor
12	 32

^aValues are means of at least two experiments.

^bHuman T-lymphoid (MT-2) cells (2×10^3) were exposed to 100 TCID₅₀S of HIV-1_{Iai} and cultured in the presence of each PI, and IC₅₀ values were determined using the MTT assay. The IC₅₀ values of amprenavir (APV), saquinavir (SQV), and indinavir (IDV) were 0.03 μ M, 0.015 μ M, and 0.03 μ M, respectively.

Table 1
Anti-HIV activity of **19b** against selected clinical isolates highly resistant to multiple protease inhibitors

Virus	Phenotype	EC ₅₀ (μM)				
		IDV	APV	LPV	DRV	19b
HIV-1 _{ERS104pre} (wild-type)	X4	0.028 ± 0.005	0.025 ± 0.006	0.03 ± 0.001	0.0036 ± 0.0002	0.028 ± 0.004
HIV-1 _{tm} (MDR)	X4	>1 (>36)	0.25 ± 0.02 (10)	0.73 ± 0.53 (24)	0.0036 ± 0.0002 (1)	0.029 ± 0.004 (1)
HIV-1 _{tm} (MDR)	R5	>1 (>36)	0.32 ± 0.03 (13)	0.72 ± 0.31 (24)	0.019 ± 0.009 (5)	0.042 ± 0.002 (2)
HIV-1 _C (MDR)	X4	>1 (>36)	0.35 ± 0.03 (14)	0.32 ± 0.01 (11)	0.015 ± 0.001 (4)	0.023 ± 0.007 (1)
HIV-1 _G (MDR)	X4	0.29 ± 0.07 (10)	0.33 ± 0.16 (13)	0.14 ± 0.01 (5)	0.014 ± 0.006 (4)	0.027 ± 0.001 (1)

Amino acid substitutions identified in the protease-encoding region of HIV-1_{ERS104pre}: HIV-1_{TM}, HIV-1_{MM}, HIV-1_C and HIV-1_G as compared to the consensus B sequence cited from the Los Alamos data base include L63P, L101/K14R/R41K/M46L/I54V/L63P/A71V/V82A/L90M/I93L, L101/K43T/M46L/I54V/L63P/A71V/V82A/L90M/Q92K, L101/I15V/K20R/L24I/M36I/M46L/I54V/I62V/L63P/K70Q/V82A/L89M, and L101/V111/T12E/I15V/L191/R41K/M46L/L63P/A71T/V82A/L90M, respectively. The EC₅₀ values were determined by employing PHA-PBM as target cells and the inhibition of p24 *Gag* protein production as an endpoint. All values were determined in duplicate or triplicate, and those shown derived from the results of three independent experiments. Numbers in parentheses represent fold changes of EC₅₀ values against each isolate compared to EC₅₀ values against HIV-1_{ERS104pre}. MDR, multi-drug-resistant.