Introduction. Biomolecular simulation

Adrian J. Mulholland*

*School of Chemistry, Centre for Computational Chemistry, University of Bristol,
Bristol BS8 1TS, UK*

‘Everything that living things do can be understood in terms of the jiggings and wiggings of atoms’ as Richard Feynman provocatively stated nearly 50 years ago. But how can we ‘see’ this wiggling and jiggling and understand how it drives biology? Increasingly, computer simulations of biological macromolecules are helping to meet this challenge.

Keywords: molecular dynamics; protein; enzyme; DNA; multi-level modelling; quantum mechanics/molecular mechanics

Experiments can provide detailed structures of biological macromolecules such as proteins, but it is hard to study directly how the structures of individual molecules change on short time scales as they function. Similarly, it is not yet possible to study directly by experiment alone the molecular mechanisms of fast processes such as chemical reactions in enzymes or ion transport through membranes. Simulations based on fundamental physics offer the potential of filling in these crucial ‘gaps’, modelling how proteins and other biomolecules move, fluctuate, interact, react and function. Simulated structures can be analysed in atomic detail, to dissect what is important in determining biological activity. Ever more powerful computers, and sophisticated techniques, mean that biomolecular simulations can now do far more than provide pretty pictures: they can show biomolecular machines at work, and analyse the mechanisms by which they function. Physics-based simulations complement experiments in building a molecular-level understanding of biology: they can test hypotheses and interpret and analyse experimental data in terms of interactions at the atomic level. A wide variety of simulation techniques have been developed, applicable to a range of different problems in biomolecular science. Simulations have already shown their worth in helping to analyse how enzymes catalyse biochemical reactions (Garcia-Viloca *et al.* 2004; Mulholland 2005; Warshel *et al.* 2006), and how proteins adopt their functional structures (Daggett 2006). They can help in the design of drugs (Jorgensen 2004; Michel *et al.* 2007) and catalysts (Jiang *et al.* 2008; Röthlisberger *et al.* 2008), and in understanding the molecular basis of disease. And simulations have played a key role in developing the conceptual framework now at the heart of biomolecular science, that is, the understanding that the way that biological molecules move and flex—their dynamics—is central to

their function (Karplus *et al.* 2005), demonstrating the truth of Feynman’s (1963) assertion.

Biomolecular simulation is a vibrant and growing area, making increasingly significant contributions to biology. The papers assembled in this issue of *Interface Focus* provide good examples of a number of different types of applications across the field. van der Kamp *et al.* (2008) provide an overview of biomolecular simulation from its origins over 30 years ago to current state-of-the-art studies and exciting prospects for future developments. Biological macromolecules, such as proteins and nucleic acids, are complex and challenging to model. They are large, often function in heterogeneous environments (e.g. cell membranes) and show complex conformational behaviour over a variety of time scales. Simulations of biomolecular motions—molecular dynamics simulations—are the most widespread type of biomolecular simulation. Such simulations calculate how the atoms of a biomolecule move according to classical (e.g. Newtonian) mechanics. Atomistic simulations can now reach the microsecond time scale for small proteins or oligonucleotides in solution (Freddolino *et al.* 2008; Mura & McCammon 2008); for larger systems, simulations are limited to tens to hundreds of nanoseconds. Using techniques of this sort on supercomputers, it is now possible to simulate, in atomic detail, the dynamics of multi-million-atom biological macromolecular assemblies such as ion channels (Gumbart *et al.* 2005; Biggin & Bond 2008; Lindahl & Sansom 2008) and the protein synthesis machinery of the ribosome (Trobro & Åqvist 2007; Trabuco *et al.* 2008), and to identify functionally relevant large-scale conformational changes in proteins (Karplus *et al.* 2005; Pentikäinen *et al.* 2008).

This *Interface Focus* contains a number of examples of molecular dynamics simulations of biomolecules. Jiranusornkul & Laughton (2008) study the effects of oxidation of guanine in DNA in molecular dynamics simulations of double-stranded DNA by molecular dynamics simulations. They find a significant change in flexibility at the oxidation site, with implications for

*adrian.mulholland@bristol.ac.uk

One contribution of 9 to a Theme Supplement ‘Biomolecular simulation’.

how repair proteins locate such regions of damaged DNA. Yan *et al.* (2008) investigate a drug target, human heat shock protein 90, using molecular dynamics simulations to study the binding of water molecules and a small molecule ligand to the protein. They find that four tightly bound water molecules should be included when modelling the interactions of ligands (potential drugs) to the protein. The water molecules appear to be an essential part of the binding site.

Simulations of the dynamics of large biomolecular systems on long time scales are feasible because they typically use fairly simple, though atomically detailed, ‘ball and spring’ type models. Empirical ‘molecular mechanics’ (MM) potential energy functions of this type represent atoms as spheres with point charges, with chemical bonds treated as springs. Years of careful parameter development have produced MM models that can give good descriptions of protein structure and dynamics. The simplified physical description, though, means that these methods cannot easily be applied to some types of problem: for example modelling chemical reactions (e.g. in enzymes) where treating the bonds as ideal springs would not allow them to break. Chemical bonding is a quantum mechanical phenomenon, so to model the making and breaking of bonds in a chemical reaction, ideally methods taking into account the essential quantum mechanics (QM) are required. QM methods, i.e. which calculate molecular electronic structure, can give very accurate results for small molecules, but are too computer intensive for large biological systems. A solution is provided by hybrid QM/MM methods, which combine a QM (electronic structure) treatment of a small region (e.g. an enzyme active site) with a MM description of the surroundings (e.g. protein and solvent). Approximate (e.g. semi-empirical) QM/MM methods are now fast enough to be used in biomolecular dynamics simulations. At the other end of the spectrum, QM/MM calculations can now be carried out with highly accurate QM methods (Claeyssens 2006), a development that promises to revolutionize enzymology (Mulholland 2007).

This *Interface Focus* contains several studies that use QM/MM methods. Senthilkumar *et al.* (2008) test QM/MM methods for modelling biomolecular hydrogen bonds (with density functional theory level QM methods), in particular the treatment of polarization, and find that these methods give a good description of these interactions, which are central to biomolecular recognition. The widest application of QM/MM methods in biomolecular modelling to date has been in the area of enzyme catalysis (Friesner & Guallar 2005; Mulholland 2005, 2008; Senn & Thiel 2007), and this issue contains two examples of such applications. Ferrer *et al.* (2008) study the important model enzyme lactate dehydrogenase. They examine the effect of an active site mutation, showing how biomolecular modelling can help to interpret the effects of site-directed mutagenesis on enzyme activity. Johannissen *et al.* (2008) examine conformational effects on the proton transfer reaction in aromatic amine dehydrogenase, which is a subject of high interest today (Lodola *et al.* 2007). This enzyme reaction involves quantum tunnelling, an area that is attracting considerable debate

(Allemann *et al.* 2006; Klinman 2006; Sutcliffe *et al.* 2006; Wang *et al.* 2006), a debate in which biomolecular simulation has a crucial role to play (Hammes-Schiffer & Watney 2006; Masgrau *et al.* 2006; Olsson *et al.* 2006). In this *Interface Focus*, Guallar & Wallrapp (2008) demonstrate a different type of application of QM/MM methods, namely a QM/MM approach to finding electron transfer pathways in proteins. They describe investigations of protein–substrate interactions in cytochrome P450cam, ascorbate peroxidase and cytochrome *c* peroxidase, and propose a new approach for studying long range electron transfer between proteins, applied here to the cytochrome *c* peroxidase–cytochrome *c* complex.

To model larger systems and/or longer time scales (e.g. over a microsecond), even simpler models than atomistic molecular dynamics are needed. Coarse-grained models use reduced representations, in which a single particle represents a group of atoms (e.g. an amino acid side chain). Here, Khalid *et al.* (2008) use coarse-grained models in molecular dynamics simulations of DNA–lipid complexes, which are potentially useful as delivery vectors in gene therapy. These workers study the self-assembly of lipid bilayers in the presence of a DNA dodecamer, observing spontaneous bilayer formation of lipid bilayers from initially random systems. They also study transfer of the DNA through the bilayer, finding a high energy barrier to DNA insertion into the bilayer hydrophobic core of the bilayer.

No single biomolecular simulation technique is capable of addressing the whole range of time scales and length scales, and range of types of processes, which arise in biomolecular science. Different techniques have to be used for different types of problem, or for different spatial or temporal scales. Combining different levels of calculation—multi-scale or multi-level modelling—is a hot and increasingly important topic in biomolecular simulation (Sherwood *et al.* in press; Woods & Mulholland in press). QM/MM methods provide an example of a hybrid, multi-level approach (Woods *et al.* 2008). Multi-level biomolecular simulation promises in future to provide exciting new insight into the workings of ‘molecular machines’ such as biological motors and pumps.

This themed supplement arose from the recent CCPB conference ‘Biomolecular Simulation 2008: Frontiers of Biomolecular Simulation’ held in Bristol, UK, 7–9 January 2008. CCPB is a collaborative project, supported by BBSRC, which aims to strengthen and broaden the UK biomolecular simulation community, and also to build links internationally (www.ccpb.ac.uk). Membership of CCPB is free and open to all. Among events organized by CCPB are specialist training workshops to train (particularly younger) researchers in this growing field. CCPB also aims to foster links between computational and experimental scientists: close interaction with experiment will be vital in the ongoing development and exploitation of modelling methods and the results they produce. Given the rapid pace of development (Woods *et al.* 2005), and the sophisticated techniques now available, it seems clear that biomolecular simulation will make

an increasingly important contribution to twenty-first century science.

A.J.M. would like to thank all the contributors to this themed supplement, and Dr Tim Holt (Publishing Editor, *J. R. Soc. Interface*) for handling all the administrative work, and his assistance. A.J.M. thanks EPSRC and BBSRC for support.

REFERENCES

- Allemann, R. K., Evans, R. M., Tey, L., Maglia, G., Pang, J., Rodriguez, R., Shrimpton, P. J. & Swanwick, R. S. 2006 Protein motions during catalysis by dihydrofolate reductases. *Phil. Trans. R. Soc. B* **361**, 1317–1321. (doi:10.1098/rstb.2006.1865)
- Biggin, P. C. & Bond, P. J. 2008 Molecular dynamics simulations of membrane proteins. *Methods Mol. Biol.* **443**, 147–160. (doi:10.1007/978-1-59745-177-2_8)
- Claeysens, F. *et al.* 2006 High-accuracy computation of reaction barriers in enzymes. *Angew. Chem. Int. Ed.* **45**, 6856–6859. (doi:10.1002/anie.200602711)
- Daggett, V. 2006 Protein folding simulation. *Chem. Rev.* **106**, 1898–1916. (doi:10.1021/cr0404242)
- Ferrer, S., Tuñón, I., Moliner, V. & Williams, I. H. 2008 Theoretical site-directed mutagenesis: Asp168Ala mutant of lactate dehydrogenase. *J. R. Soc. Interface* **5**, S217–S224. (doi:10.1098/rsif.2008.0211.focus)
- Feynman, R. 1963 *The Feynman lectures on physics*, vol. 1. Reading, MA: Addison-Wesley Publishing Company.
- Freddolino, P. L., Liu, F., Gruebele, M. & Schulten, K. 2008 Ten-microsecond molecular dynamics simulation of a fast-folding WW domain. *Biophys. J.* **94**, L75–L77. (doi:10.1529/biophysj.108.131565)
- Friesner, R. A. & Guallar, V. 2005 *Ab initio* quantum chemical and mixed quantum mechanics/molecular mechanics (QM/MM) methods for studying enzymatic catalysis. *Annu. Rev. Phys. Chem.* **56**, 389–427. (doi:10.1146/annurev.physchem.55.091602.094410)
- García-Viloca, M., Gao, J., Karplus, M. & Truhlar, D. G. 2004 How enzymes work: analysis by modern rate theory and computer simulations. *Science* **303**, 186–195. (doi:10.1126/science.1088172)
- Guallar, V. & Wallrapp, F. 2008 Mapping protein electron transfer pathways with QM/MM methods. *J. R. Soc. Interface* **5**, S233–S239. (doi:10.1098/rsif.2008.0061.focus)
- Gumbart, J., Wang, Y., Aksimentiev, A., Tajkhorshid, E. & Schulten, K. 2005 Molecular dynamics simulations of proteins in lipid bilayers. *Curr. Opin. Struct. Biol.* **15**, 423–431. (doi:10.1016/j.sbi.2005.07.007)
- Hammes-Schiffer, S. & Watney, J. B. 2006 Hydride transfer catalysed by *Escherichia coli* and *Bacillus subtilis* dihydrofolate reductase: coupled motions and distal mutations. *Phil. Trans. R. Soc. B* **361**, 1365–1373. (doi:10.1098/rstb.2006.1869)
- Jiang, L. *et al.* 2008 De novo computational design of retroaldol enzymes. *Science* **319**, 1387–1391. (doi:10.1126/science.1152692)
- Jiranusornkul, S. & Loughton, C. A. 2008 Destabilization of DNA duplexes by oxidative damage at guanine: implications for lesion recognition and repair. *J. R. Soc. Interface* **5**, S191–S198. (doi:10.1098/rsif.2008.0304.focus)
- Johannissen, L. O., Scrutton, N. S. & Sutcliffe, M. J. 2008 The enzyme aromatic amine dehydrogenase induces a substrate conformation crucial for promoting vibration that significantly reduces the effective potential energy barrier to proton transfer. *J. R. Soc. Interface* **5**, S225–S232. (doi:10.1098/rsif.2008.0068.focus)
- Jorgensen, W. L. 2004 The many roles of computation in drug discovery. *Science* **303**, 1813–1818. (doi:10.1126/science.1096361)
- Karplus, M., Gao, Y. Q., Ma, J. P., van der Vaart, A. & Yang, W. 2005 Protein structural transitions and their functional role. *Phil. Trans. R. Soc. A* **363**, 331–355. (doi:10.1098/rsta.2004.1496)
- Khalid, S., Bond, P. J., Holyoake, J., Hawtin, R. W. & Sansom, M. S. P. 2008 DNA and lipid bilayers: self-assembly and insertion. *J. R. Soc. Interface* **5**, S241–S250. (doi:10.1098/rsif.2008.0239.focus)
- Klinman, J. P. 2006 Linking protein structure and dynamics to catalysis: the role of hydrogen tunnelling. *Phil. Trans. R. Soc. B* **361**, 1323–1331. (doi:10.1098/rstb.2006.1870)
- Lindahl, E. & Sansom, M. S. 2008 Membrane proteins: molecular dynamics simulations. *Curr. Opin. Struct. Biol.* **18**, 425–431. (doi:10.1016/j.sbi.2008.02.003)
- Lodola, A., Mor, M., Zurek, J., Tarzia, G., Piomelli, D., Harvey, J. N. & Mulholland, A. J. 2007 Conformational effects in enzyme catalysis: reaction via a high energy conformation in fatty acid amide hydrolase. *Biophys. J.* **92**, L20–L22. (doi:10.1529/biophysj.106.098434)
- Masgrau, L. *et al.* 2006 Atomic description of an enzyme reaction dominated by proton tunneling. *Science* **312**, 237–241. (doi:10.1126/science.1126002)
- Michel, J., Verdonk, M. L. & Essex, J. W. 2007 Protein-ligand complexes: computation of the relative free energy of different scaffolds and binding modes. *J. Chem. Theor. Comput.* **3**, 1645–1655. (doi:10.1021/ct700081t)
- Mulholland, A. J. 2005 Modelling enzyme reaction mechanisms, specificity and catalysis. *Drug Discov. Today* **10**, 1393–1402. (doi:10.1016/S1359-6446(05)03611-1)
- Mulholland, A. J. 2007 Chemical accuracy in QM/MM calculations on enzyme-catalysed reactions. *Chem. Central J.* **1**, 1–5. (doi:10.1186/1752-153X-1-1)
- Mulholland, A. J. 2008 Computational enzymology: modelling the mechanisms of biological catalysts. *Biochem. Soc. Trans.* **36**, 22–26. (doi:10.1042/BST0360022)
- Mura, C. & McCammon, J. A. 2008 Molecular dynamics of a κ B DNA element: base flipping via cross-strand intercalative stacking in a microsecond-scale simulation. *Nucleic Acids Res.* **36**, 4941–4955. (doi:10.1093/nar/gkn473)
- Pentikäinen, U., Pentikäinen, O. T. & Mulholland, A. J. 2008 Cooperative symmetric to asymmetric conformational transition of the apo-form of scavenger decapping enzyme revealed by simulations. *Proteins: Struct. Funct. Bioinform.* **70**, 498–508. (doi:10.1002/prot.21540)
- Röthlisberger, D. *et al.* 2008 Kemp elimination catalysts by computational enzyme design. *Nature* **453**, 190–195. (doi:10.1038/nature06879)
- Senn, H. M. & Thiel, W. 2007 QM/MM studies of enzymes. *Curr. Opin. Chem. Biol.* **11**, 182–187. (doi:10.1016/j.cbpa.2007.01.684)
- Senthilkumar, K., Mujika, J. I., Ranaghan, K. E., Manby, F. R., Mulholland, A. J. & Harvey, J. N. 2008 Analysis of polarization in QM/MM modelling of biologically relevant hydrogen bonds. *J. R. Soc. Interface* **5**, S207–S216. (doi:10.1098/rsif.2008.0243.focus)
- Sherwood, P., Brooks, B. R. & Sansom, M. S. In press. Multiscale methods for macromolecular simulations. *Curr. Opin. Struct. Biol.* (doi:10.1016/j.sbi.2008.07.003)
- Sutcliffe, M. J. *et al.* 2006 Hydrogen tunnelling in enzyme-catalysed H-transfer reactions: flavoprotein and quinoprotein systems. *Phil. Trans. R. Soc. B* **361**, 1375–1386. (doi:10.1098/rstb.2006.1878)
- Trabuco, L. G., Villa, E., Mitra, K., Frank, J. & Schulten, K. 2008 Flexible fitting of atomic structures into electron microscopy maps using molecular dynamics. *Structure* **16**, 673–683. (doi:10.1016/j.str.2008.03.005)

- Trobro, S. & Åqvist, J. 2007 A model for how ribosomal release factors induce peptidyl-tRNA cleavage in termination of protein synthesis. *Mol. Cell* **27**, 758. (doi:10.1016/j.molcel.2007.06.032)
- van der Kamp, M. W., Shaw, K. E., Woods, C. J. & Mulholland, A. J. 2008 Biomolecular simulation and modelling: status, progress and prospects. *J. R. Soc. Interface* **5**, S173–S190. (doi:10.1098/rsif.2008.0105.focus)
- Wang, L., Goodey, N. M., Benkovic, S. J. & Kohen, A. 2006 The role of enzyme dynamics and tunnelling in catalysing hydride transfer: studies of distal mutants of dihydrofolate reductase. *Phil. Trans. R. Soc. B* **361**, 1307–1315. (doi:10.1098/rstb.2006.1871)
- Olsson, M. H. M., Mavri, J. & Warshel, A. 2006 Transition state theory can be used in studies of enzyme catalysis: lessons from simulations of tunnelling and dynamical effects in lipoxygenase and other systems. *Phil. Trans. R. Soc. B* **361**, 1417–1432. (doi:10.1098/rstb.2006.1880)
- Warshel, A., Sharma, P. K., Kato, M., Xiang, Y., Liu, H. B. & Olsson, M. H. M. 2006 Electrostatic basis for enzyme catalysis. *Chem. Rev.* **106**, 3210–3235. (doi:10.1021/cr0503106)
- Woods, C. J. & Mulholland, A. J. In press. Multiscale modelling of biological systems. In *RSC specialist periodicals reports, chemical modelling: applications and theory* (ed. A. Hinchliffe). Cambridge UK: Royal Society of Chemistry.
- Woods, C. J. *et al.* 2005 Grid computing and biomolecular simulation. *Phil. Trans. R. Soc. A* **363**, 2017–2035. (doi:10.1098/rsta.2005.1626)
- Woods, C. J., Manby, F. R. & Mulholland, A. J. 2008 An efficient method for the calculation of QM/MM free energies. *J. Chem. Phys.* **128**, 014 109. (doi:10.1063/1.2805379)
- Yan, A., Grant, G. H. & Richards, W. G. 2008 Dynamics of conserved waters in human HSp90: implications for drug design. *J. R. Soc. Interface* **5**, S199–S205. (doi:10.1098/rsif.2008.0331.focus)