Published in final edited form as:

Bioorg Med Chem Lett. 2007 July 15; 17(14): 4049–4052.

Synthesis of 7-Thiaarachidonic Acid as a Mechanistic Probe of Prostaglandin H Synthase-2

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Abstract

The mechanism by which prostaglandin synthase converts arachidonic acid to prostaglandin G₂, creating five new chiral centers in the process, is still incompletely understood. The first radical intermediate has been characterized by EPR spectroscopy but subsequent proposed intermediates have not succumbed to detection. We report the synthesis of 7-thiaarachidonic acid designed to stabilize one of the proposed radical intermediates, which may allow its detection.

Prostaglandin H synthase (PGHS, also called cyclooxygenase) catalyzes the conversion of arachidonic acid to PGG₂ through a radical cascade that is initiated via hydrogen atom abstraction from the C13 position of the substrate by an active site tyrosyl radical (Figure 1).^{1–5} The resulting radical intermediate **I** is then proposed to react with molecular oxygen to form peroxy radical **II**.⁶ A 5-exo trig cyclization onto the C8-C9 double bond leads to formation of intermediate **III**, followed by a second 5-exo trig cyclization to produce the 2.2.1 bicyclic intermediate **IV**. This radical is proposed to undergo a second reaction with oxygen to generate the peroxy radical intermediate **V**, which is converted to PGG₂ by abstraction of a hydrogen atom from tyrosine, thereby regenerating the active form of the enzyme.^{6,7}

Under anaerobic conditions, the initially formed radical intermediate **I** has been characterized as a pentadienyl radical in PGHS-2.^{8–10} Experimental observation of subsequent radicals requires the presence of molecular oxygen as a co-substrate, but under aerobic conditions no radical intermediates are observed. Therefore the proposed carbon centered radicals **II–V** are not present in observable concentrations during aerobic turnover. In order to observe these radical intermediates, the rate determining step would have to be altered by either mutagenesis of the enzyme or modification of the substrate.

A primary kinetic isotope effect observed when tritium was incorporated at the C13 position⁶ indicates that the initial hydrogen atom abstraction is at least partially rate limiting. This is corroborated by computational studies, which have concluded that the formation of intermediates **IV** and **V** require similar activation energies as the hydrogen atom abstraction from C13.^{11,12} This implies that stabilizing either intermediate **III** or **IV** could cause a shift in the rate determining step for the reaction and allow detection of these intermediates.

Stabilization of carbon-centered radicals generated during enzyme catalysis can be achieved in a number of ways.¹³ Resonance delocalization using an adjacent π -system has been

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Supporting Information Available. Experimental procedures and spectral characterization of all synthetic compounds.

frequently used.^{13–18} An alternative approach involves incorporation of a sulfur atom adjacent to the radical site. This approach has been successfully used by Frey and co-workers to investigate the reaction mechanism of lysine 2,3-aminomutase.^{19,20}

In that study the stabilization provided by the sulfur atom was estimated as ~ 9 kcal/mol. The substantial stabilization of an α -sulfide radical over an alkyl radical has also been used in reversible addition fragmentation chain transfer (RAFT) processes to retard the rate of polymerization reactions.^{21–23}

On the basis of these predicted thermodynamic stabilities and the previously calculated barriers for conversion of **III** to **V** we embarked on the preparation of 7-thiaarachidonic acid **1** (Figure 2). It was envisioned that if 7-thiaarachidonic acid were accepted by the enzyme, the proposed reaction mechanism would lead to the generation of radical intermediate **III'**, which would be stabilized by the adjacent sulfur atom and the possibly also by additional resonance stabilization involving the C5-C6 alkene. Previous studies have shown that 7-thiaarachidonic acid is a reversible inhibitor of PGHS-1.²⁴ Inhibition of the PGHS-2 isozyme, for which the pentadienyl radical **I** has been characterized experimentally, has not been investigated. The PGHS-2 active site is larger and is more tolerant to alteration. For instance, acetylation of Ser530 of PGHS-1 by aspirin abolishes all catalytic activity whereas acetylation of the corresponding Ser516 in PGHS-2 does not prevent substrate binding and hydrogen atom abstraction from C13.^{25–29} Similar differences between the two isozymes have also been reported for site directed mutants of active site residues.^{27,30,31}

The synthetic challenge for 7-thiaarachidonic acid lies in the bis(Z-alkenyl)sulfide functionality in a molecule with two additional non-conjugated Z-alkenes. The synthesis of 7-thiaarachidonic acid has been previously reported in the mid 1980s in 17 steps.²⁴ With the development in the intervening time of alternative methodologies, we elected to use a different synthetic route to the target compound. The most direct approach for the synthesis of the bis (Z-alkenyl)sulfide moiety is the stereoselective reduction of a bis(alkynyl)sulfide. Although to the best of our knowledge the reduction of a bis(alkynyl)sulfide has not been reported, several methods exist for reduction of a 1-alkynylsulfide. Using bis(hexenyl)sulfide **2** as a model system, several of these reduction conditions were screened. DIBAL-H reduction of the diyne was accomplished in moderate yield (55 %), but the *E/Z* selectivity was poor (1:1). Hydrogenation conditions previously reported for the reduction of 1-alkynylsulfides or 1-alkynylsulfoxides were also explored. However, Lindlar's catalyst³² and Wilkinson's catalyst³³ did not produce the reduced product in appreciable yield. Finally, alkynyl sulfide reduction utilizing a stoichiometric amount of tantalum chloride and zinc metal was attempted.³⁴ This reaction yielded the reduced bis(hexenyl)sulfide in 60% yield with very good stereocontrol as only the (Z,Z)-isomer was observed by ¹H and ¹³C NMR spectroscopy.

The tantalum-promoted stereoselective reduction is believed to proceed through initial formation of a low-valent tantalum species.³⁵ Addition of the alkyne then generates a metallacyclopentadiene intermediate **VI** (Figure 3). The formation of such tantalum- η^2 -alkyne complexes has been demonstrated by ¹H and ¹³C NMR spectroscopy as well as x-ray crystallography.^{36,37} These air-sensitive intermediates can be hydrolyzed under basic conditions to yield the alkene, generally in good yield.

The low-valent tantalum reduction was attempted next on a more functionalized and asymmetric bis(alkynyl)sulfide. Treatment of alkynes **4** and **5** with *n*-butyllithium followed by slow addition of sulfur dichloride yielded the unsymmetric bis(alkynyl)sulfide **6** (Figure 4).^{38–40} Sulfide **6** was subjected to the tantalum-mediated reaction conditions resulting in formation of the bis(alkenyl)sulfide **7** in 65% yield and excellent stereoselectivity as only the Z,Z-isomer was detected by ¹H NMR spectroscopy. Interestingly, the reaction required the

presence of a terminal hexyne in order to afford reproducible yields of the product. This requirement was discovered serendipitously because the initial attempt at reduction of **6** was conducted with substrate that still contained some **4**. Subsequent reactions with pure **6** were unsuccessful unless 1-hexyne was added. Previous experiments have demonstrated that alkynylsulfides react faster than terminal alkynes with low-valent tantalum metal.⁴¹ Thus, the role of 1-hexyne in the reaction is not understood.

The synthesis of 7-thiaarachidonic acid was completed as depicted in Figure 4. Deprotection of **7** with a 1.0 M solution of tetrabutylammonium fluoride afforded alcohol **8** in good yield. This alcohol was reacted with triphenylphosphine and bromine to produce bromide **9**, which required rapid chromatographic purification as the compound was unstable to silica gel. Heating of the bromide with triphenylphosphine afforded phosphonium salt **10**.

cis-3-Nonenal was synthesized by oxidation of *cis*-nonenol with Dess-Martin periodinane in quantitative yield.⁴² A Wittig reaction between phosphonium salt **10** and *cis*-3-nonenal under *Z*-selective conditions^{43,44} yielded *tert*-butyl thiaarachidonate **11**, which was deprotected with a 1.0 M aqueous solution of lithium hydroxide and DME (1:2, v/v). The resulting 7-thiaarachidonic acid was purified by HPLC and isolated as a single isomer. The ¹H NMR spectrum of the compound was consistent with the spectrum reported by Corey and co-workers.²⁴

One advantage of the tantalum pentachloride reduction is the facile incorporation of deuterium at the C5-C9 alkene positions, which is advantageous for EPR characterization of any substrate based radicals that may be formed with PGHS or human lipoxygenases that also use arachidonic acid as substrate for radical based transformations.^{45,46} The reaction is believed to proceed by basic hydrolysis of a metallocyclopropene intermediate. This step is consistent with the observation that quenching the reaction with a basic solution of deuterium oxide results in deuterium incorporation at both alkene positions for 1,2-disubstituted alkynes.^{35,41} To test whether this labeling strategy could be extended to bis(alkynyl)sulfides, the reduction of **6** was repeated, but a 1.0 M solution of sodium deuterioxide in deuterium oxide was used during the hydrolysis step. The reaction proceeded in good yield and stereocontrol. Based on ¹H NMR integration, the bis(alkenyl)sulfide **7a** contained 97% deuterium at each alkene position. The remaining synthetic steps were identical to the synthetic route for the 7-thiaarachidonic acid. Deprotection with tetrabutylammonium fluoride yielded alcohol **8a**, which was halogenated to form bromide **9a**. Treatment with triphenylphosphine yielded phosphonium salt **10a**. A Wittig reaction then afforded *t*-butyl 7-thia(5,6,8,9-²H₄)arachidonate **11a** in good yield. Final deprotection yielded the desired 7-thia(5,6,8,9-²H₄)arachidonic acid **1a**, which was purified by HPLC to afford the final compound as a single isomer.

In summary, we have developed a new synthetic route to 7-thiaarachidonic acid that marks an improvement over the previously reported route. Our unoptimized synthesis of the target compound was completed in only seven linear steps and allowed the rapid and convenient preparation of deuterium labeled analogs. EPR studies of the interaction of these compounds with PGHS-2 and human lipoxygenases are currently ongoing.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgment

This work was supported by a pre-doctoral fellowship to CMM from the American Heart Association (#0310006Z) and by the National Institutes of Health (GM44911).

References

1. Karthein R, Dietz R, Nastainczyk W, Ruf HH. Eur. J. Biochem 1988;171:313. [PubMed: 2828053]
2. Tsai AL, Hsi LC, Kulmacz RJ, Palmer G, Smith WL. J. Biol. Chem 1994;269:5085. [PubMed: 8106487]
3. Shimokawa T, Kulmacz RJ, DeWitt DL, Smith WL. J. Biol. Chem 1990;265:20073. [PubMed: 2122967]
4. van der Donk WA, Tsai AL, Kulmacz RJ. Biochemistry 2002;41:15451. [PubMed: 12501173]
5. Rouzer CA, Marnett LJ. Chem. Rev 2003;103:2239. [PubMed: 12797830]
6. Hamberg M, Samuelsson B. J. Biol. Chem 1967;242:5336. [PubMed: 6070851]
7. Wei C, Kulmacz RJ, Tsai AL. Biochemistry 1995;34:8499. [PubMed: 7599139]
8. Tsai AL, Kulmacz RJ, Palmer G. J. Biol. Chem 1995;270:10503. [PubMed: 7737984]
9. Peng S, Okeley NM, Tsai AL, Wu G, Kulmacz RJ, van der Donk WA. J. Am. Chem. Soc 2001;123:3609. [PubMed: 11472139]
10. Peng S, Okeley NM, Tsai AL, Wu G, Kulmacz RJ, van der Donk WA. J. Am. Chem. Soc 2002;124:10785. [PubMed: 12207535]
11. Blomberg ML, Blomberg MRA, Siegbahn PEM, van der Donk WA, Tsai AL. J. Phys. Chem. B 2003;107:3297.
12. Silva PJ, Fernandes PA, Ramos MJ. Theoretical Chemistry Accounts 2003;110:345.
13. Stubbe J, van der Donk WA. Chem. Rev 1998;98:705. [PubMed: 11848913]
14. Magnusson OT, Reed GH, Frey PA. J. Am. Chem. Soc 1999;121:9764.
15. Abend A, Bandarian V, Reed GH, Frey PA. Biochemistry 2000;39:6250. [PubMed: 10821701]
16. Magnusson OT, Reed GH, Frey PA. Biochemistry 2001;40:7773. [PubMed: 11425303]
17. Wu W, Booker S, Lieder KW, Bandarian V, Reed GH, Frey PA. Biochemistry 2000;39:9561. [PubMed: 10924153]
18. van der Donk WA, Gerfen GJ, Stubbe J. J. Am. Chem. Soc 1998;120:4252.
19. Wu W, Lieder KW, Reed GH, Frey PA. Biochemistry 1995;34:10532. [PubMed: 7654708]
20. Miller J, Bandarian V, Reed GH, Frey PA. Arch. Biochem. Biophys 2001;387:281. [PubMed: 11370852]
21. Alnajjar MS, Zhang XM, Franz JA, Bordwell FG. J. Org. Chem 1995;60:4976.
22. Coote ML. Macromolecules 2004;37:5023.
23. Coote ML, Henry DJ. Macromolecules 2005;38:1415.
24. Corey EJ, Cashman JR, Eckrich TM, Corey DR. J. Am. Chem. Soc 1985;107:713.
25. Holtzman MJ, Turk J, Shornick LP. J. Biol. Chem 1992;267:21438. [PubMed: 1400457]
26. Lecomte M, Laneuville O, Ji C, DeWitt DL, Smith WL. J. Biol. Chem 1994;269:13207. [PubMed: 8175750]
27. Mancini JA, O'Neill GP, Bayly C, Vickers PJ. FEBS Lett 1994;342:33. [PubMed: 8143845]
28. Schneider C, Brash AR. J. Biol. Chem 2000;275:4743. [PubMed: 10671506]
29. Tsai AL, Palmer G, Wu G, Peng S, Okeley NM, van der Donk WA, Kulmacz RJ. jbc 2002;277:38311.
30. Thuresson ED, Lakkides KM, Rieke CJ, Sun Y, Wingerd BA, Micielli R, Mulichak AM, Malkowski MG, Garavito RM, Smith WL. J. Biol. Chem 2001;276:10347. [PubMed: 11121412]
31. Schneider C, Boeglin WE, Prusakiewicz JJ, Rowlinson SW, Marnett LJ, Samel N, Brash AR. J. Biol. Chem 2002;277:478. [PubMed: 11677234]
32. Beccalli EM, Marchesini A, Pilati T. Tetrahedron 1994;50:12697.
33. Maezaki N, Izumi M, Yuyama S, Sawamoto H, Iwata C, Tanaka T. Tetrahedron 2000;56:7927.
34. Takai K, Miyai J, Kataoka Y, Utimoto K. Organometallics 1990;9:3030.
35. Kataoka Y, Takai K, Oshima K, Utimoto K. Tetrahedron Letters 1990;31:365.
36. Hierso JC, Etienne M. European Journal of Inorganic Chemistry 2000:839.
37. Oshiki T, Tanaka K, Yamada J, Ishiyama T, Kataoka Y, Mashima K, Tani K, Takai K. Organometallics 2003;22:464.

38. Verboom W, Schoufs M, Meijer J, Verkruijsse HD, Brandsma L. Recl. Trav. Chim. Pays-Bas 1978;97:244.
39. Ruecker C, Fritz H. Magn. Reson. Chem 1988;26:1103.
40. Lewis KD, Wenzler DL, Matzger AJ. Org. Lett 2003;5:2195. [PubMed: 12816407]
41. Kataoka Y, Takai K, Oshima K, Utimoto K. J. Org. Chem 1992;57:1615.
42. Wavrin L, Viala J. Synthesis 2002:326.
43. Greenwald R, Chaykovsky M, Corey EJ. J. Am. Chem. Soc 1963;28:1128.
44. Bestmann HJ, Stransky W, Vostrowsky O. Chem. Ber 1976;109:1694.
45. McGinley CM, van der Donk WA. Chem. Commun 2003:2843.
46. Brash AR. J. Biol. Chem 1999;274:23679. [PubMed: 10446122]

Figure 1. Proposed mechanism for the conversion of arachidonic acid to prostaglandin H₂. With the exception of radical I, no spectroscopic evidence is currently available for any of the proposed intermediates.

Figure 2.
Structure of the synthetic target 7-thiaarachidonic acid **1** and the stabilized intermediate radical **III'**.

Figure 3. Test substrate **2** for the stereoselective reduction of bisalkynylsulfides. The metallocyclopropene **VI** has been proposed as an intermediate in the tantalum catalyzed reduction.

Figure 4. Synthetic scheme for the preparation of 7-thia-arachidonic acid and its deuterium-labeled analogs.