

# Update from the 4th Edition of the World Health Organization Classification of Head and Neck Tumours: Hematolymphoid Tumours

Noah A. Brown<sup>1</sup> · Kojo S. J. Elenitoba-Johnson<sup>2</sup>

Received: 23 December 2016 / Accepted: 6 February 2017 / Published online: 28 February 2017  
© Springer Science+Business Media New York 2017

**Abstract** In 2017, the latest revision to the WHO Classification of Head and Neck Tumours will be released. Similar to the 2005 WHO, the codification of hematopoietic and lymphoid neoplasms of the head and neck is included within chapters pertaining to the nasal cavity and paranasal sinuses, the nasopharynx, the larynx, the oral cavity and oropharynx, the neck and the salivary glands. Herein, we describe both changes to the classification of hematolymphoid neoplasms of the head and neck since the 2005 WHO, as well as recent advances in our understanding of the underlying pathogenesis and molecular pathology of these neoplasms.

**Keywords** WHO · 2017 · Revision · Edition · Hematolymphoid · Hematopoietic · Lymphoma

## Introduction

This section applies to hematopoietic and lymphoid neoplasms of the head and neck including the nasal cavity and paranasal sinuses, the nasopharynx, the larynx, the oral cavity and oropharynx, the neck and the salivary glands. While there have been relatively few changes to the

classification of hematolymphoid neoplasms of the head and neck since the 2005 WHO classification of Head and Neck Tumours, there have been significant advances in our understanding of the underlying pathogenesis and molecular pathology of these neoplasms (See Table 1).

## CD30-Positive T-Cell Lymphoproliferative Disorder

CD30-positive T-cell lymphoproliferative disorder (TLPD) encompasses a clinicopathologic spectrum that typically occurs in the oral cavity or occasionally other mucosal sites in the head and neck. This entity was not formally codified in the 2005 WHO classification of Head and Neck Tumours, but has since been added to the current edition. These tumors are analogous to the clinicopathologic spectrum observed in primary cutaneous CD30-positive TLPD. Primarily affecting older adults, these lesions typically present as a mass, often with ulceration [1]. Lesions show sheets of large and atypical neoplastic cells with pleomorphic nuclei, often within a mixed inflammatory background [2–4]. Neoplastic cells show strong uniform CD30, by definition, express CD4 more often than CD8, frequently express cytotoxic markers and are negative for CD56, ALK and EBV [4]. Recently, rearrangements involving *DUSP22* on 6p25.3 have been identified in these lesions, similar to those observed in primary cutaneous TLPD and ALK-negative anaplastic large cell lymphoma [3]. Importantly, most cases of CD30-positive TLPD show complete resolution with local therapy alone [3–5].

Special Issue: World Health Organization Classification Update

✉ Kojo S. J. Elenitoba-Johnson  
Kojo.Elenitoba-Johnson@uphs.upenn.edu

<sup>1</sup> Department of Pathology, University of Michigan Health System, Ann Arbor, MI, USA

<sup>2</sup> Department of Pathology, Perelman School of Medicine at University of Pennsylvania, 609A Stellar Chance Laboratories, 420 Curie Boulevard, Philadelphia, PA 1904, USA

**Table 1** Summary of histologic, immunophenotypic, pathogenic and prognostic features of head and neck hematolymphoid neoplasms

| Diagnosis   | Histologic features  | Immunophenotype   | Molecular/pathogenesis  | Prognosis  |
|---|--|---|---|--|
| CD30-positive T-cell lymphoproliferative disorder       | Analogous to primary cutaneous CD30-positive T-cell lymphoproliferative disorder. Sheets of large, atypical cells with mixed inflammatory background | Strong, uniform CD30. CD4 more often than CD8. Cytotoxic marker expression. Negative for CD56, ALK, EBER  | Some cases with rearrangements of <i>DUSP22</i> on 6p25.3   | Most cases resolve with local therapy alone  |
| Extranodal NK/T-cell lymphoma                           | Angiocentric/angiodestructive with variably atypical T-cells and geographic necrosis   | T-cell antigens, cytotoxic markers, EBER and CD56   | EBV infection with downregulation of MiR-146a and MiR-15a. Mutation of tumor suppressor genes <i>TP53</i> , <i>DDX3X</i> and genes in region of 6q21-23 ( <i>PRDM1</i> , <i>ATG5</i> , <i>AIM1</i> , <i>HACE1</i> and <i>FOXO3</i> ). Activating mutations of <i>JAK3</i> , <i>STAT3</i> , <i>PTPRK</i> | Aggressive behavior with variable prognosis. Invasion of bone or skin, high circulating EBV DNA and EBV + cells in the bone marrow associated with worse prognosis                     |
| Extraosseous plasmacytoma                               | Sheets of variably atypical plasma cells. Extranodal marginal zone lymphoma and plasmablastic lymphoma are important differential diagnoses          | CD138, CD38, MUM1/IRF4. Variable CD79A. CD20 and PAX5 negative  | Similar genetic features as plasma cell myeloma with some differences in <i>IGH</i> translocation partners  | Most cases eradicated by local radiation. Recurrence in ~25%. Progression to plasma cell myeloma in ~15%   |
| Diffuse large B-cell lymphoma (DLBCL)                   | Sheets of large cells with centroblastic, immunoblastic or anaplastic cytology   | Expression of B-cell markers  | Germinal center B-cell: alterations of <i>BCL6</i> , <i>BCL2</i> translocations, <i>EZH2</i> mutations, <i>GNAI3</i> mutations. Activated B-cell: mutations activating BCR/TLR and NFκB pathways ( <i>MYD88</i> , <i>CD79A</i> , <i>CARD11</i> , <i>TNFAIP3</i> )                                       | Aggressive behavior with variable prognosis. Bone marrow involvement, concurrent MYC and BCL2 and/or BCL6 rearrangements and MYC and BCL2 co-expression associated with poor prognosis |
| EBV + mucocutaneous ulcer                               | Circumscribed ulcer with polymorphous infiltrate containing large, atypical B-cells often resembling Reed–Sternberg cells                            | Expression of B-cell markers, EBER positivity   | EBV infection with waning immune defense due to age or iatrogenic immunosuppression   | Spontaneous regression or relapsing–remitting course requiring only conservative management  |
| Classical Hodgkin lymphoma (CHL)                        | Mixed inflammatory background with mononuclear Hodgkin cells and/or multi-nucleated Reed–Sternberg cells   | CD30 positive, CD15 positive in most cases. Weak positivity for PAX5. Negative or weakly positive for CD20 and CD79A  | Crippled B-cell expression program. Constitutive activation of NFκB. EBV in many cases  | Curable in most cases with chemotherapy and radiation  |
| Nodular lymphocyte predominant Hodgkin lymphoma (NLPHL) | Nodules comprised predominantly of small T-cells, studded with large cells with lobulated nuclei (LP cells)  | LP cells express B-cell antigens and are negative for CD30 and CD15. LP cells rosetted by cells with T follicular helper cell immunophenotype. CD21 highlights follicular dendritic meshwork with nodules | Frequent <i>BCL6</i> rearrangements. Somatic hypermutation in <i>PAX5</i> , <i>PIM1</i> , <i>RHOH</i> and <i>MYC</i> . No EBV   | Slow progression with excellent prognosis for low-stage disease. Progression to DLBCL in 3–5% of cases   |

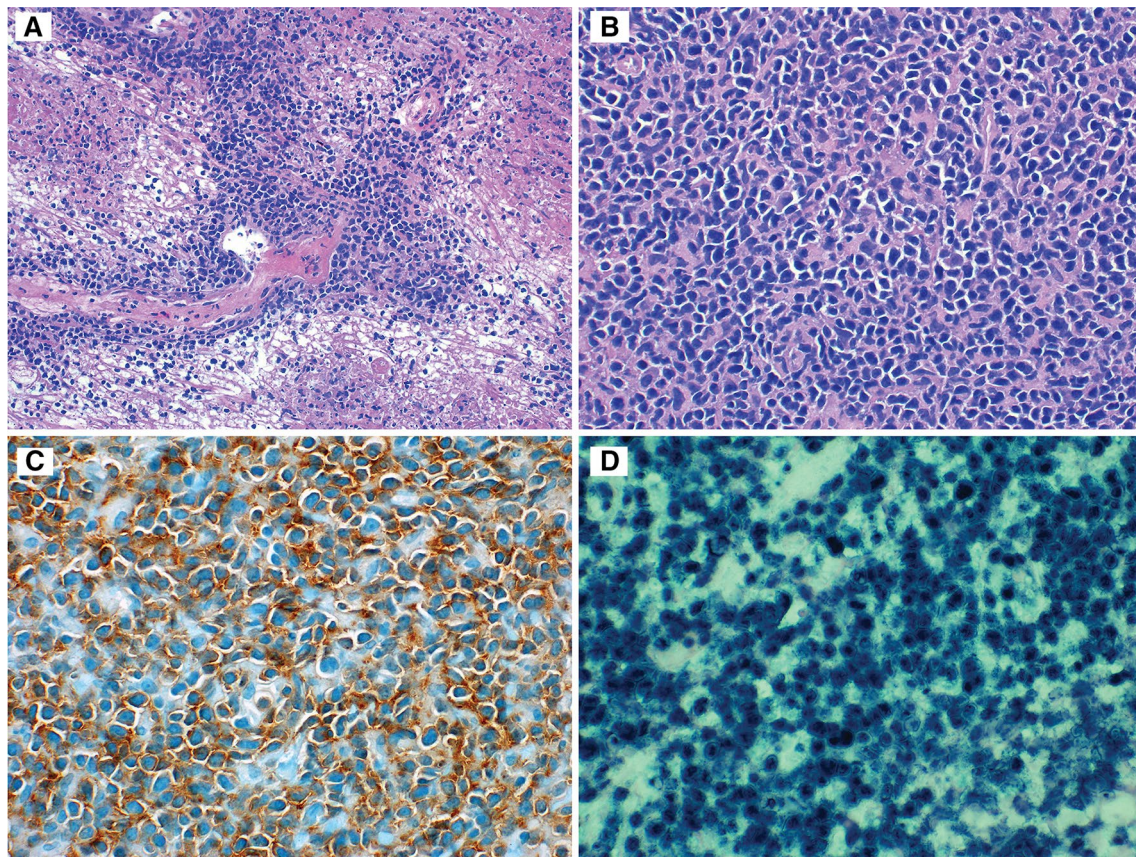
**Table 1** (continued)

| Diagnosis                                   | Histologic features  | Immunophenotype   | Molecular/pathogenesis   | Prognosis   |
|---|--|---|--|---|
| Burkitt lymphoma BL                         | Monomorphic medium-sized cells with squared-off borders, mitotic figures and scattered tingible body macrophages   | Expression of B-cell antigens and germinal center markers. Negative for TdT and negative or weakly positive for BCL2. 100% proliferative index (Ki-67). May be EBER positive, particularly endemic variant  | <i>MYC</i> rearrangements in 90%, most commonly <i>IGH</i> , less frequently <i>IGK</i> or <i>IGL</i> . <i>MYC</i> negative lymphomas resembling Burkitt with 11q aberration recognized as separate provisional entity in 2016 Heme WHO. Endemic cases frequently show EBV, cytomegalovirus and HHV8 infection, expression of EBV lytic genes and recurrent alterations of <i>ARID1A</i> , <i>CCNF</i> and <i>RHOA</i> . Sporadic cases frequently show alterations of <i>MYC</i> , <i>ID3</i> , <i>TCF3</i> and <i>TP53</i> | Highly aggressive but intensive chemotherapy regimens result in cure in up 90%  |
| Follicular lymphoma (FL)                    | Follicular and/or diffuse pattern with centrocytes and centroblasts. Follicles are closely packed with absent or attenuated mantles and lack of polarization or tingible body macrophages. FL in children and young adults (particularly in the tonsil) shows high-grade features i.e. prominent centroblasts or blastoid follicular cells | Expression of B-cell antigens and germinal center markers. Follicles show staining for follicular dendritic cells (CD21, CD23)  | Most cases demonstrate a t(14;18). Tonsillar FL in children and young adults typically lack t(14;18) and <i>BCL6</i> or <i>MYC</i> rearrangements and instead carry <i>MUM1/IRF4</i> rearrangements in 50%   | Low-grade FL (grade 1–2) is indolent and not usually curable. The majority of grade 3 FL show an aggressive clinical course. However, FL in children and young adults - tonsillar in particular - shows a more localized, indolent clinical behavior than conventional grade 3 FL |
| Mantle cell lymphoma (MCL)                  | Small to intermediate-sized, monomorphic cells arranged in a vaguely nodular, diffuse, mantle zone or rarely follicular pattern. No proliferation centers. May have hyalinized vessels and/or epithelioid histiocytes  | Expression of B-cell markers, CD5 and cyclin D1. SOX11 in rare cyclin D1-negative MCL   | Most cases demonstrate t(11;14) <i>CCND1-IGH</i> . Cyclin D1-negative MCL may have rearrangements of <i>CCND2</i> or <i>CCND3</i> . Mutations of ATM and TP53 common   | More aggressive than other small B-cell lymphomas with median survival of 3–5 years. Most cases cannot be cured   |
| T-lymphoblastic leukemia/lymphoma (T-LBL/L) | Small to intermediate-sized cells with fine chromatin, variably prominent nucleoli and scant cytoplasm   | Most positive for cytoplasmic CD3, TdT, CD7 and CD1a; variably positive for CD10; either double positive or double-negative for CD4 and CD8. Early T precursor (ETP) lymphoblastic leukemia/lymphoma is CD7 positive without CD1a or CD8 and positive for one or more myeloid/stem cell markers | Frequent rearrangements involving the T-cell receptor genes. Activation of Notch signalling and loss of CDKN2A common. ETP with frequent mutations of myeloid-associated genes   | Aggressive disease often with more high-risk features than B-LBL/L. Indolent T-lymphoblastic proliferation is a non-neoplastic entity that can mimic T-LBL/L and frequently involves lymphoid tissue of the head and neck   |

**Table 1** (continued)

| Diagnosis   | Histologic features   | Immunophenotype   | Molecular/pathogenesis  | Prognosis   |
|---|---|---|---|---|
| Extranodal marginal zone lymphoma (MALT lymphoma) | Heterogeneous mixture of B-cells including marginal zone (centrocyte-like) cells, monocytoid B-cells, small lymphocytes, plasma cells and scattered immunoblasts and centroblast-like cells. Neoplastic cells typically infiltrate the epithelium forming lymphoepithelial lesions  | Expression of B-cell antigens. Negative for CD5, CD10, BCL6 and cyclin D1   | In salivary glands, lymphoepithelial sialadenitis is a precursor lesion. Monoclonal B cells present in more than half of salivary lesions from Sjögren patients and acquisition of mutations such as in TNFAIP3 associated with lymphoma development. Translocations involving MALT1 including t(14;18) (IGH-MALT1), t(11;18) (API2-MALT1) are uncommon in head and neck MALT lymphomas. Hepatitis C is another predisposing factor. Anecdotal evidence of MALT lymphoma arising in the setting of IgG4-related disease | Variable clinical course. Advance stage, high circulating EBV DNA associated with inferior prognosis  |
| Follicular dendritic cell (FDC) sarcoma           | Mass with pushing border, often arranged in fascicular, whorled or storiform patterns with admixed small lymphocytes. Cells may be spindled, ovoid or epithelioid cells with eosinophilic cytoplasm, indistinct cell borders and finely dispersed chromatin. Nuclear pseudoinclusions, and binucleated or multinucleated cells are common. A subset of cases are associated with hyaline-vascular Castleman disease | Positivity for FDC-associated antigens including CD21, CD23, CD35, clusterin, CXCL13 and podoplanin   | Unknown   | Surgery is potentially curable in early disease, but late recurrence and metastasis can occur. Large tumor size, disseminated disease, extensive necrosis, high mitotic rate and nuclear atypia associated with worse prognosis |
| Plasmablastic lymphoma (PBL)                      | Sheets of large cells resembling immunoblasts   | Expression of plasmacytic markers including CD138, CD38 and IRF4/MUM1. Negative or only weakly positive for CD45, CD20 and PAX5. EBER positive in >80%. Negative for HHV8 | Most frequently in the setting of HIV-related immunosuppression. Associated with EBV infection, and MYC dysregulation with <i>MYC</i> translocation or gains in half of cases   | Very aggressive with most patients dying within a year  |
| Langerhans cell histiocytosis (LCH)               | A mixture of inflammatory cells, including eosinophils, and neoplastic Langerhans cells with folded or grooved nuclei and moderate to abundant cytoplasm  | Positivity for S100, CD1a and CD207 (langerin)  | Activation of MAPK pathway with <i>BRAF</i> V600E in 50% and <i>MAP2K1</i> mutations in 25%   | Highly dependent on staging with >99% survival for unifocal disease and ~33% for multisystem involvement  |
| Extramammary myeloid sarcoma                      | Effacement of tissue architecture by myeloblasts with or without promyelocytic or neutrophilic maturation   | CD68 positivity most common followed by MPO, CD117, lysozyme, CD34 and TdT  | Variety of chromosomal aberrations including monosomy 7, trisomy 8, inv(16) and t(8;21)-particularly in pediatric patients. NPM1 mutations are also found in 16% of cases   | Variably but often unfavorable  |





**Fig. 1** Extranodal NK/T-cell lymphoma **a** Angiocentric growth with surrounding necrosis. **b** Monotonous, intermediate-sized atypical lymphocytes. **c** CD56 positivity. **d** Positivity for EBV in-situ hybridization (EBER)

### Extranodal NK/T-Cell Lymphoma

Extranodal NK/T-cell lymphoma (ENKTL), nasal type is an extranodal lymphoma with a cytotoxic phenotype, associated with EBV. While this entity was recognized in the 2005 WHO, there have been significant advances in our understanding its underlying pathogenesis. This lymphoma most commonly occurs in the upper aerodigestive tract and is more prevalent among East Asians [6] and the indigenous populations of Mexico and Central and South America [7]. Commonly presenting as nasal obstruction and/or epistaxis, ENKTL frequently shows an angiocentric/angiodestructive pattern leading to geographic necrosis. The tumor cells express T cell antigens, cytotoxic markers (TIA1, granzyme B or perforin), EBV RNA (EBER) and frequently CD56 [8] (Fig. 1).

Since the 2005 WHO, our understanding of the molecular pathogenesis of ENKTL has expanded. Several tumor suppressor genes within the commonly deleted region at 6q21-23 have recently been identified including *PRDM1*, *ATG5*, *AIM1*, *HACE1*, and *FOXO3* [9–11]. In addition to *TP53* [12], *DDX3X* was recently identified as another commonly mutated tumor suppressor gene [13]. The JAK/

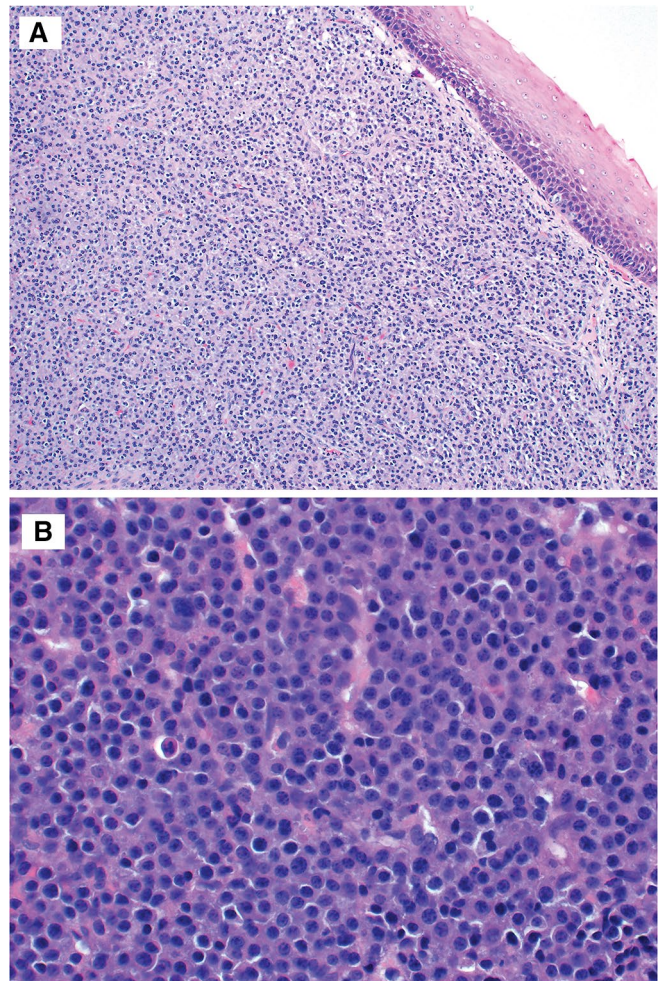
STAT pathway is activated in most cases with frequent genetic alterations of *JAK3*, *STAT3* or *PTPRK* [14–17]. While EBV has long been implicated in the pathogenesis of ENKTL, recent studies have shown that EBV infection downregulated host microRNA and that downregulation of MiR-146a [18] and miR-15a promotes cell proliferation and is associated with a poor prognosis [19]. Recent studies have also highlighted the role of quantitative EBV DNA assay in plasma and PET findings in risk stratification [20].

### Extrasosseous Plasmacytoma

Extrasosseous (extramedullary) plasmacytoma is a mass-forming proliferation of clonal plasma cells in the absence of underlying plasma cell myeloma, as described in the previous WHO. Important differential diagnoses to exclude include plasmablastic lymphoma and extranodal marginal zone lymphoma with extensive plasmacytic differentiation. These tumors frequently involve the upper respiratory tract and appear as sheets of plasma cells expressing markers of plasmacytic differentiation including CD138, CD38 and MUM1/IRF4 (Fig. 2). CD79A is variably expressed



**Fig. 2** Extraosseous (extramedullary) plasmacytoma. **a** A mass formed by monotonous cells with abundant cytoplasm underlying squamous epithelium. **b** Sheets of plasma cells with clumped chromatin and abundant eosinophilic cytoplasm



while CD20 and PAX5 are typically negative. Plasmablastic lymphoma should be considered in the setting of EBV positivity.

Studies have shown similar genetic features between plasma cell myeloma and extraosseous plasmacytoma though differences in *IGH* translocation partners have been reported [21].

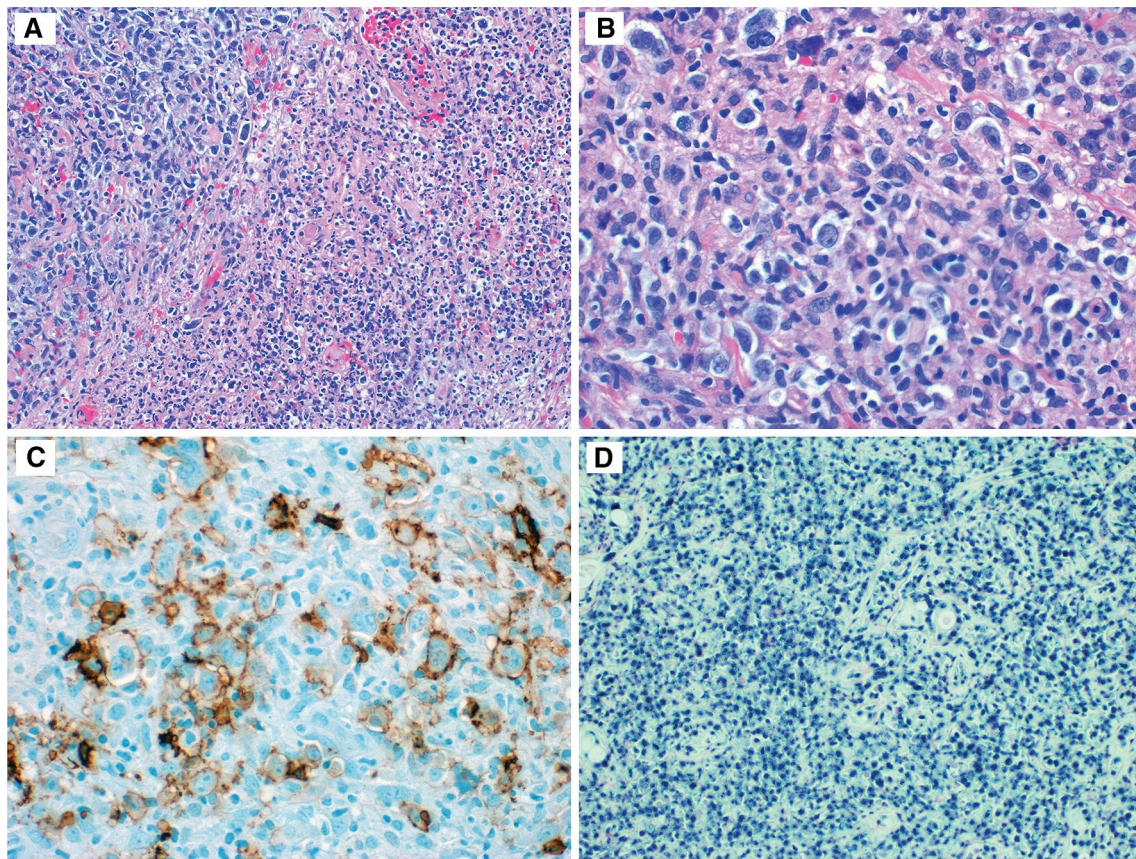
### Diffuse Large B-Cell Lymphoma

Diffuse large B-cell lymphoma (DLBCL) is a diffuse infiltration by intermediate to large-sized B cells with variable cytological features including centroblastic, immunoblastic and anaplastic. DLBCL can occur at any site in the head and neck region. The 2017 Head and Neck WHO does not specifically address the recent advances made in understanding of DLBCL or the separate classifications of DLBCL recognized in the 2016 WHO classification of hematopoietic and lymphoid tumors [22]. In particular, the importance of classifying DLBCL based on cell of origin—activated B-cell (ABC) or germinal center B-cell

(GCB)—is widely accepted based on differences in clinical outcome, gene expression, chromosomal alterations and somatic mutations [23, 24]. Recent next-generation sequencing studies have shown that both GCB and ABC DLBCL frequently have inactivating mutations of *TP53* and immunosurveillance genes, alterations of epigenetic regulators and oncogenic activation of *BCL6*. GCB DLBCL have frequent alterations *EZH2* and *BCL2* translocations similar to follicular lymphoma as well as mutations in the cell motility regulator *GNA13*. In contrast, ABC DLBCL have frequent mutations in genes activating BCR/TLR and NFκB pathways (e.g. *MYD88*, *CD79A*, *CARD11*, *TNFAIP3*). Additional important categories include high grade B-cell lymphoma (HGBL) with rearrangements of *MYC* and *BCL2* and/or *BCL6* which is associated with a poor prognosis [17, 19, 25]. Recent studies have also suggested that “double-expressor lymphomas” (i.e. *MYC* and *BCL2*) may also have a worse outcome [26].

EBV + DLBCL, NOS is also recognized as a distinct entity by the 2016 WHO classification of hematopoietic and lymphoid tumors [17, 27, 28]. The “NOS” designation is intended to exclude other entities with neoplastic





**Fig. 3** EBV+ mucocutaneous ulcer. **a** Ulcerated mandibular gingiva with underlying infiltrate of large atypical cells. **b** Polymorphous infiltrate of large, markedly atypical cells resembling Reed–Sternberg

cells. **c** CD20 positivity consistent with a B-cell process. **d** Uniform positivity for EBV in-situ hybridization (EBER)

EBV+ B-cells such as lymphomatoid granulomatosis. However, in the head and neck region, it is important to consider a EBV+ mucocutaneous ulcer, an indolent condition codified as a provisional entity in the 2016 WHO classification of hematopoietic and lymphoid tumors [17]. These lesions frequently appear as circumscribed oral ulcers in immunocompromised or elderly patients and are characterized by a polymorphous infiltrate with atypical large B-cells often resembling Reed–Sternberg cells [29] (Fig. 3). These ulcers may regress spontaneously or have a relapsing-remitting course, generally requiring only conservative management [24, 30].

As discussed with follicular lymphoma, it is also important to distinguish DLBCL, NOS from Large B-cell lymphoma with IRF4 rearrangement [17]. This lymphoma which generally occurs in children and young adults most frequently involves the cervical lymph nodes and Waldeyer ring.

## Hodgkin Lymphoma

As in the 2005 WHO, classical Hodgkin lymphoma (CHL) is distinct from nodular lymphocyte predominant Hodgkin lymphoma (NLPHL). In CHL, the neoplastic cells (mononuclear Hodgkin cells and multi-nucleated Reed–Sternberg cells) have lost expression of many B-cell specific proteins. They are weakly positive for PAX5 but are typically negative (or only weakly positive) for CD20 and CD79A. CD30 is positive in virtually all cases and CD15 in most. The inflammatory background typically consists of T-cells, plasma cells, histiocytes and eosinophils. In NLPHL, the neoplastic cells have lobulated nuclei, retain B-cell antigen expression are rosetted by T follicular helper cells.

The revision acknowledges that there can be overlap between NLPHL and T-cell histiocyte rich large B-cell lymphoma.

## Burkitt Lymphoma

Burkitt lymphoma (BL) is a B-cell lymphoma with an extremely high proliferation rate, often involving extranodal sites, composed of monomorphic medium-sized cells [31]. The current Head and Neck WHO is largely unchanged from the 2005 edition; although the 2016 WHO classification recognizes a new provisional entity. The diagnosis of BL requires integration of morphology, immunophenotype and genetics. Three variants are recognized: endemic (regions where malaria is endemic), sporadic, and immunodeficiency-associated. The characteristic starry-sky pattern is formed by closely packed neoplastic cells with scattered phagocytizing macrophages. Neoplastic cells are positive for CD20, CD10, BCL6 and MYC and are typically negative for BCL2. Ki-67 typically demonstrates a proliferation index of 100% and EBER is frequently positive.

Approximately 90% of cases demonstrate a rearrangement involving *MYC* (8q24), most commonly with *IGH* (14q32) or less frequently to *IGK* (2p12) or *IGL* (22q11) [19, 32, 33]. Some controversy exists over whether cases without *MYC* rearrangements should be classified as BL. These cases are characterized by 11q alterations including proximal gains and telomeric losses [34, 35]. While these cases resemble BL morphologically, immunophenotypically and by gene expression profiling, they show more complex karyotypes, lower *MYC* expression and may show more cytological pleomorphism. These cases are classified as BL according to the 2017 Head and Neck WHO while the 2016 WHO classification of hematopoietic and lymphoid tumors recognizes a provisional entity designated “Burkitt-like lymphoma with 11q aberration.” [17].

Gene expression profiling studies have shown that endemic and immunodeficiency-associated BL have nearly identical signatures while sporadic cases differ in expression of many genes [36, 37]. Similarly, large-scale sequencing studies have demonstrated differences between endemic and sporadic BL [38, 39]. Endemic cases frequently show both cytomegalovirus and HHV8 infection, expression of EBV lytic genes and recurrent alterations of *ARID1A*, *CCNF* and *RHOA*. By contrast, sporadic cases more frequently show alterations of *MYC*, *ID3*, *TCF3* and *TP53*.

## Follicular Lymphoma

Follicular lymphoma (FL) is a lymphoma comprised of centrocytes and centroblasts with at least a partially follicular pattern. This category may include both pediatric and adult FL which are now acknowledged to have important clinicopathologic differences. In the head and neck, cervical lymph nodes are the most common site with Waldeyer ring involvement typically representing secondary tonsillar

extension from nodal disease [40]. Isolated oropharyngeal FL is rare, most often occurring in children and young adults. In keeping with a germinal center phenotype, cells are positive for CD10 and BCL6. BCL2 is positive in 85–90% of cases.

The revised WHO acknowledges the differences between FL in the tonsil of children and young adults and other cases of FL. FL in this setting typically shows large expansile follicles, positivity for MUM1/IRF4, a high proliferative index and prominent centroblasts or blastoid follicular center cells rather than classic centrocytes [41, 42]. While most FL cases bear the t(14;18) translocation, MUM1/IRF4-positive FL in children and young adults typically lack the t(14;18) translocation as well as *BCL6* and *MYC* rearrangements and instead carries translocations involving *MUM1/IRF4* in approximately 50% of cases [29, 30]. The 2016 WHO classification of hematopoietic and lymphoid tumors separately classifies these cases as “pediatric-type” follicular lymphoma and avoids classifying these cases as conventional grade 3 FL given the generally localized indolent behavior of these cases [17, 29, 30]. In addition, a separate provision entity designated “large B-cell lymphoma with IRF4 rearrangement” has been designated [17, 29, 43]. This entity most frequently occurs in Waldeyer ring and/or cervical lymph nodes and is meant to include lymphomas with follicular, follicular and diffuse, or pure diffuse growth patterns that would otherwise be designated FL grade 3B or DLBCL. This lymphoma is considered more aggressive than other pediatric-type FL, though patients still seem to respond well to treatment [31]. Additional recent findings include the discovery of recurrent mutations of *CREBBP*, *KMT2D* and *EZH2* occurring early in the pathogenesis of FL [44–46].

## Mantle Cell Lymphoma

Mantle cell lymphoma (MCL) is an aggressive B-cell lymphoma of small to medium-sized cells. As described in the 2005 WHO, lymphoma cells usually carry a translocation involving *CCND1* leading to cyclin D1 overexpression. However, the current WHO recognizes cases of cyclin D1-negative MCL. The head and neck region is the second most common extranodal site with MCL accounting for 2.6% of all Waldeyer ring non-Hodgkin lymphomas [47–49]. MCL may show a diffuse, vaguely nodular or mantle-zone pattern and is typically positive for IgD, CD20, CD5 and cyclin D1.

The revised WHO points out that SOX11 is useful in identifying rare cyclin D1-negative cases of MCL [50, 51]. While most cases have t(11;14) *CCND1-IGH* translocations, cyclin-D1 negative cases may have rearrangements



of *CCND2* or *CCND3* [52, 53]. Mutations in many other genes including *ATM* and *TP53* are also common [54].

### T-Lymphoblastic Leukemia/Lymphoma

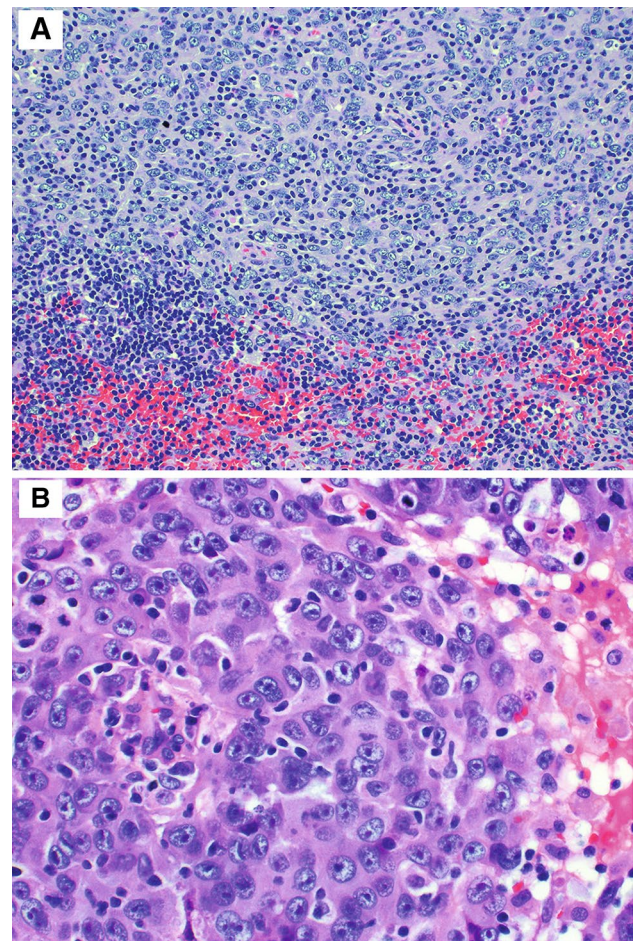
T-lymphoblastic leukemia/lymphoma (T-LBL/L) is a neoplasm of lymphoblasts committed to T-cell lineage that rarely occurs in the head and neck region [55]. This entity was recognized in the 2005 WHO, but the current edition points out important diagnostic distinctions and recent molecular discoveries. T-LBL/L consists of small to medium-sized cells with fine chromatin, variably conspicuous nucleoli and scant cytoplasm. Lymphoblasts are typically positive for cytoplasmic CD3, TdT, CD7 and CD1a; variably positive for CD10; and either double-positive or double negative for CD4 and CD8. Early T precursor (ETP) acute lymphoblastic leukemia (ALL) is recognized as a provisional entity in the 2016 WHO classification for hematopoietic and lymphoid tumors [56]. This leukemia is characterized by CD7 without CD1a or CD8 and positivity for one or more myeloid/stem cell markers [57]. A high frequency of mutations in myeloid-associated genes is also associated with ETP ALL [58–60].

Many cases of T-LBL/L have an abnormal karyotype with frequent translocations involving the T-cell receptor genes [61]. Activation of Notch signaling and loss of *CDKN2A* are also common [62].

As the revised WHO points out, it is important to distinguish T-LBL/L from indolent T-lymphoblastic proliferation which is now well recognized as a non-neoplastic entity that can mimic T-LBL/L and frequently involves lymphoid tissue of head and neck [63]. Histologically, these lesions resemble normal thymic cortex without thymic epithelium.

### Follicular Dendritic Cell Sarcoma

Follicular dendritic cell (FDC) sarcoma is a rare tumor of nodal and extranodal sites that exhibits phenotypic features similar to FDC. No significant changes have been made in the current classification of this entity. While FDC sarcoma accounts for less than 1% of head and neck tumors, the head and neck is the most common site of involvement for this tumor—most frequently cervical lymph nodes or Waldeyer ring. The tumor displays a pushing border with spindle, ovoid or epithelioid cells, often arranged in fascicular, whorled or storiform growth patterns with admixed small lymphocytes (Fig. 4). The cells have eosinophilic cytoplasm with indistinct cell borders and elongated nuclei with finely dispersed chromatin. Nuclear pseudoinclusions, and binucleated or multinucleated cells are common. A subset of cases are associated with hyaline-vascular Castleman

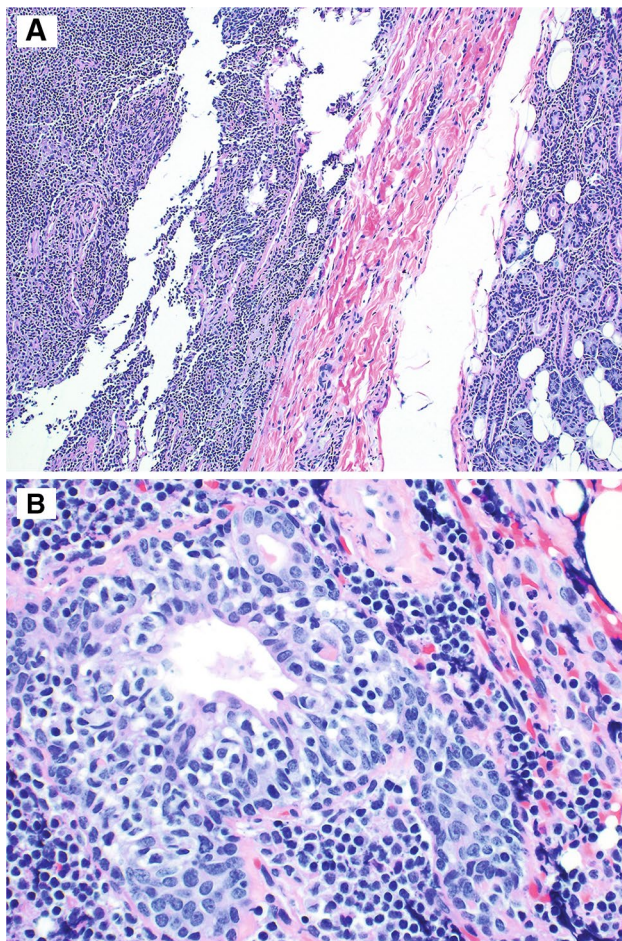


**Fig. 4** Follicular dendritic cell sarcoma. **a** Proliferation of spindle to epithelioid cells within a lymph node with a pushing border. **b** Sheets of large, epithelioid cells with prominent nucleoli, abundant eosinophilic cytoplasm and indistinct cell boundaries, lightly infiltrated by small lymphocytes

disease. Tumor cells are positive for FDC associated antigens including CD21, CD23, CD35, clusterin, CXCL13 and podoplanin. The pathogenesis of FDC sarcoma remains unknown.

### Extranodal Marginal Zone Lymphoma of Mucosa-Associated Lymphoid Tissue (MALT Lymphoma)

Extranodal marginal zone lymphoma of mucosa-associated lymphoid tissue (MALT lymphoma) remains largely unchanged in the 2017 WHO. This is an indolent B-cell lymphoma composed of heterogeneous mixture of B-cells including marginal zone (centrocyte-like) cells, monocytoid B-cells, small lymphocytes, plasma cells and scattered immunoblasts and centroblast-like cells [64] (Fig. 5). The neoplastic cells typically infiltrate the epithelium forming



**Fig. 5** Extranodal marginal zone lymphoma of mucosa-associated lymphoid tissue (MALT lymphoma). **a** Parotid gland containing a mass comprised of small lymphocytes. **b** Lymphoepithelial lesions in which glandular tissue is distorted and destroyed by invading neoplastic lymphocytes

Lymphoepithelial lesions. The head and neck is the second most frequent site of MALT lymphoma, most frequently involving the ocular adnexa, salivary glands, and less commonly Waldeyer ring [65, 66]. MALT lymphoma cells are positive for B-cell antigens and are typically negative for CD5, CD10, BCL6 and cyclin D1. The revised WHO points out that IRTA1, a marker of marginal zone differentiation is positive in the majority of cases [67]. In salivary glands, lymphoepithelial sialadenitis is a precursor lesion for MALT lymphoma [68]. Monoclonal B cells are present in more than half of salivary lesions from Sjögren syndrome patients and the acquisition of mutations such as in *TNFAIP3* has been associated with lymphoma development [69]. However, translocations involving *MALT1* including t(14;18) (*IGH-MALT1*), t(11;18) (*API2-MALT1*) which are

commonly observed in MALT lymphomas of other sites including the stomach, are uncommon in head and neck MALT lymphomas [70]. Hepatitis C is another predisposing factor [71] and there is anecdotal evidence of MALT lymphoma arising in the setting of IgG4-related disease [72, 73].

### Plasmablastic Lymphoma

Plasmablastic lymphoma (PBL) is a proliferation of large cells resembling immunoblasts with a plasmacytic immunophenotype. Previously codified as a distinctive form of DLBCL, this neoplasm is now recognized as a completely separate entity. PBL most commonly occurs in the oral cavity but can occur in other extranodal sites including the oropharynx, nasopharynx and sinonasal tract [74, 75]. Lymph nodes are occasionally involved. PBL most frequently occurs in the setting of HIV-related immunosuppression.

PBL is associated with EBV infection and *MYC* dysregulation with *MYC* translocations or gains occurring in approximately half of cases [20, 76, 77].

### Langerhans Cell Histiocytosis

Langerhans cell histiocytosis (LCH) is a neoplastic proliferation of Langerhans cells with a peak incidence between 3 and 5 years of age [78]. Similar to other hematolymphoid neoplasms, significant advances in understanding of the pathogenesis of this disease have been made since 2005. Head and neck involvement most commonly affects the skull, orbit, jaw, scalp, periauricular skin, cervical lymph nodes, paranasal sinuses and oral mucosa [79]. Histologically, lesions contain a mixture of inflammatory cells and neoplastic cells with folded or grooved nuclei [80]. These cells express S100, CD1a and CD207 (langerin).

LCH is now known to result from activation of the mitogen activated protein kinase (MAPK) pathway resulting from *BRAF* V600E mutations in approximately half of cases [81] and *MAP2K1* mutations in approximately one quarter [82].

### Extramedullary Myeloid Sarcoma

Extramedullary myeloid sarcoma is mass consisting of myeloid blasts occurring outside of the bone marrow. There have not been significant changes in description of this entity since the 2005 WHO. This disease can occur de novo or be associated with acute myeloid leukemia or another myeloid neoplasm [83]. The oral cavity is the most common site, but any head and neck site may be involved [84].



A variety of chromosomal aberrations have been reported including monosomy 7, trisomy 8, inv(16) and t(8;21)—particularly in pediatric patients [28]. *NPM1* mutations are also found in 16% of cases [85].

## References

- Ficarra G, Prignano F, Romagnoli P. Traumatic eosinophilic granuloma of the oral mucosa: a CD30+ (Ki-1) lymphoproliferative disorder? *Oral Oncol*. 1997;33(5):375–9.
- Alobeid B, Pan LX, Milligan L, Budel L, Frizzera G. Eosinophil-rich CD30+ lymphoproliferative disorder of the oral mucosa. A form of “traumatic eosinophilic granuloma”. *Am J Clin Pathol*. 2004;121(1):43–50.
- Sciallis AP, Law ME, Inwards DJ, McClure RF, Macon WR, Kurtin PJ, Dogan A, Feldman AL. Mucosal CD30-positive T-cell lymphoproliferations of the head and neck show a clinicopathologic spectrum similar to cutaneous CD30-positive T-cell lymphoproliferative disorders. *Mod Pathol*. 2012;25(7):983–92.
- Wang W, Cai Y, Sheng W, Lu H, Li X. The spectrum of primary mucosal CD30-positive T-cell lymphoproliferative disorders of the head and neck. *Oral Surg Oral Med Oral Pathol Oral Radiol*. 2014;117(1):96–104.
- Agarwal M, Shenjere P, Blewitt RW, Hall G, Sloan P, Pigadas N, Banerjee SS. CD30-positive T-cell lymphoproliferative disorder of the oral mucosa—an indolent lesion: report of 4 cases. *Int J Surg Pathol*. 2008;16(3):286–90.
- Aoki R, Karube K, Sugita Y, Nomura Y, Shimizu K, Kimura Y, Hashikawa K, Suefuji N, Kikuchi M, Ohshima K. Distribution of malignant lymphoma in Japan: analysis of 2260 cases, 2001–2006. *Pathol Int*. 2008;58(3):174–82.
- Laurini JA, Perry AM, Boilesen E, Diebold J, MacLennan KA, Müller-Hermelink HK, Nathwani BN, Armitage JO, Weisenburger DD. Classification of non-Hodgkin lymphoma in Central and South America: a review of 1028 cases. *Blood*. 2012;120(24):4795–801.
- Pongpruttipan T, Sukpanichnant S, Assanasen T, Wannakrairot P, Boonsakan P, Kanoksil W, Kayasut K, Mitarnun W, Khuhapinant A, Bunworasate U, Puavilai T, Bedavanija A, Garcia-Herrera A, Campo E, Cook JR, Choi J, Swerdlow SH. Extranodal NK/T-cell lymphoma, nasal type, includes cases of natural killer cell and  $\alpha\beta$ ,  $\gamma\delta$ , and  $\alpha\beta/\gamma\delta$  T-cell origin: a comprehensive clinicopathologic and phenotypic study. *Am J Surg Pathol*. 2012;36(4):481–99.
- Iqbal J, Kucuk C, Deleeuw RJ, Srivastava G, Tam W, Geng H, Klinkebiel D, Christman JK, Patel K, Cao K, Shen L, Dybkaer K, Tsui IF, Ali H, Shimizu N, Au WY, Lam WL, Chan WC. Genomic analyses reveal global functional alterations that promote tumor growth and novel tumor suppressor genes in natural killer-cell malignancies. *Leukemia*. 2009;23(6):1139–51.
- Karube K, Nakagawa M, Tsuzuki S, Takeuchi I, Honma K, Nakashima Y, Shimizu N, Ko YH, Morishima Y, Ohshima K, Nakamura S, Seto M. Identification of FOXO3 and PRDM1 as tumor-suppressor gene candidates in NK-cell neoplasms by genomic and functional analyses. *Blood*. 2011;118(12):3195–204.
- Küçük C, Hu X, Iqbal J, Gaulard P, Klinkebiel D, Cornish A, Dave BJ, Chan WC. HACE1 is a tumor suppressor gene candidate in natural killer cell neoplasms. *Am J Pathol*. 2013;182(1):49–55.
- Quintanilla-Martinez L, Kremer M, Keller G, Nathrath M, Gamboa-Dominguez A, Meneses A, Luna-Contreras L, Cabras A, Hoefler H, Mohar A, Fend F. p53 Mutations in nasal natural killer/T-cell lymphoma from Mexico: association with large cell morphology and advanced disease. *Am J Pathol*. 2001;159(6):2095–105.
- Jiang L, Gu ZH, Yan ZX, Zhao X, Xie YY, Zhang ZG, Pan CM, Hu Y, Cai CP, Dong Y, Huang JY, Wang L, Shen Y, Meng G, Zhou JF, Hu JD, Wang JF, Liu YH, Yang LH, Zhang F, Wang JM, Wang Z, Peng ZG, Chen FY, Sun ZM, Ding H, Shi JM, Hou J, Yan JS, Shi JY, Xu L, Li Y, Lu J, Zheng Z, Xue W, Zhao WL, Chen Z, Chen SJ. Exome sequencing identifies somatic mutations of DDX3X in natural killer/T-cell lymphoma. *Nat Genet*. 2015;47(9):1061–6.
- Huang Y, de Reyniès A, de Leval L, Ghazi B, Martin-Garcia N, Travert M, Bosq J, Brière J, Petit B, Thomas E, Coppo P, Marafioti T, Emile JF, Delfau-Larue MH, Schmitt C, Gaulard P. Gene expression profiling identifies emerging oncogenic pathways operating in extranodal NK/T-cell lymphoma, nasal type. *Blood*. 2010;115(6):1226–37.
- Koo GC, Tan SY, Tang T, Poon SL, Allen GE, Tan L, Chong SC, Ong WS, Tay K, Tao M, Quek R, Loong S, Yeoh KW, Yap SP, Lee KA, Lim LC, Tan D, Goh C, Cutcutache I, Yu W, Ng CC, Rajasegaran V, Heng HL, Gan A, Ong CK, Rozen S, Tan P, Teh BT, Lim ST. Janus kinase 3-activating mutations identified in natural killer/T-cell lymphoma. *Cancer Discov*. 2012;2(7):591–7.
- Coppo P, Gouilleux-Gruart V, Huang Y, Bouhlal H, Bouamar H, Bouchet S, Perrot C, Vieillard V, Dartigues P, Gaulard P, Agbaliika F, Douay L, Lassoued K, Gorin NC. STAT3 transcription factor is constitutively activated and is oncogenic in nasal-type NK/T-cell lymphoma. *Leukemia*. 2009;23(9):1667–78.
- Chen YW, Guo T, Shen L, Wong KY, Tao Q, Choi WW, Au-Yeung RK, Chan YP, Wong ML, Tang JC, Liu WP, Li GD, Shimizu N, Loong F, Tse E, Kwong YL, Srivastava G. Receptor-type tyrosine-protein phosphatase  $\kappa$  directly targets STAT3 activation for tumor suppression in nasal NK/T-cell lymphoma. *Blood*. 2015;125(10):1589–600.
- Paik JH, Jang JY, Jeon YK, Kim WY, Kim TM, Heo DS, Kim CW. MicroRNA-146a downregulates NF $\kappa$ B activity via targeting TRAF6 and functions as a tumor suppressor having strong prognostic implications in NK/T cell lymphoma. *Clin Cancer Res*. 2011;17(14):4761–71.
- Komabayashi Y, Kishibe K, Nagato T, Ueda S, Takahara M, Harabuchi Y. Downregulation of miR-15a due to LMP1 promotes cell proliferation and predicts poor prognosis in nasal NK/T-cell lymphoma. *Am J Hematol*. 2014;89(1):25–33.
- Kim SJ, Choi JY, Hyun SH, Ki CS, Oh D, Ahn YC, Ko YH, Choi S, Jung SH, Khong PL, Tang T, Yan X, Lim ST, Kwong YL, Kim WS, Asia Lymphoma Study Group. Risk stratification on the basis of Deauville score on PET-CT and the presence of Epstein–Barr virus DNA after completion of primary treatment for extranodal natural killer/T-cell lymphoma, nasal type: a multicentre, retrospective analysis. *Lancet Haematol*. 2015;2(2):e66–e74.
- Bink K, Haralambieva E, Kremer M, Ott G, Beham-Schmid C, de Leval L, Peh SC, Laeng HR, Jütting U, Hutzler P, Quintanilla-Martinez L, Fend F. Primary extramedullary plasmacytoma: similarities with and differences from multiple myeloma revealed by interphase cytogenetics. *Haematologica*. 2008;93(4):623–6.
- Swerdlow SH, Campo E, Pileri SA, Harris NL, Stein H, Siebert R, Advani R, Ghielmini M, Salles GA, Zelenetz AD, Jaffe ES. The 2016 revision of the World Health Organization classification of lymphoid neoplasms. *Blood*. 2016;127(20):2375–90.
- Young RM, Shaffer AL, Phelan JD, Staudt LM. B-cell receptor signaling in diffuse large B-cell lymphoma. *Semin Hematol*. 2015;52(2):77–85.
- Roschewski M, Staudt LM, Wilson WH. Diffuse large B-cell lymphoma-treatment approaches in the molecular era. *Nat Rev Clin Oncol*. 2014;11(1):12–23.



25. Karube K, Campo E. MYC alterations in diffuse large B-cell lymphomas. *Semin Hematol*. 2015;52(2):97–106.
26. Johnson NA, Slack GW, Savage KJ, Connors JM, Ben-Neriah S, Rogic S, Scott DW, Tan KL, Steidl C, Sehn LH, Chan WC, Iqbal J, Meyer PN, Lenz G, Wright G, Rimsza LM, Valentino C, Brunhoeber P, Grogan TM, Brazier RM, Cook JR, Tubbs RR, Weisenburger DD, Campo E, Rosenwald A, Ott G, Delabie J, Holcroft C, Jaffe ES, Staudt LM, Gascoyne RD. Concurrent expression of MYC and BCL2 in diffuse large B-cell lymphoma treated with rituximab plus cyclophosphamide, doxorubicin, vincristine, and prednisone. *J Clin Oncol*. 2012;30(28):3452–9.
27. Dojcinov SD, Venkataraman G, Pittaluga S, Wlodarska I, Schrager JA, Raffeld M, Hills RK, Jaffe ES. Age-related EBV-associated lymphoproliferative disorders in the Western population: a spectrum of reactive lymphoid hyperplasia and lymphoma. *Blood*. 2011;117(18):4726–35.
28. Nicolae A, Pittaluga S, Abdullah S, Steinberg SM, Pham TA, Davies-Hill T, Xi L, Raffeld M, Jaffe ES. EBV-positive large B-cell lymphomas in young patients: a nodal lymphoma with evidence for a tolerogenic immune environment. *Blood*. 2015;126(7):863–72.
29. Dojcinov SD, Venkataraman G, Raffeld M, Pittaluga S, Jaffe ES. EBV positive mucocutaneous ulcer—a study of 26 cases associated with various sources of immunosuppression. *Am J Surg Pathol*. 2010;34(3):405–17.
30. Hart M, Thakral B, Yohe S, Balfour HH Jr, Singh C, Spears M, McKenna RW. EBV-positive mucocutaneous ulcer in organ transplant recipients: a localized indolent post-transplant lymphoproliferative disorder. *Am J Surg Pathol*. 2014;38(11):1522–9.
31. Molyneux EM, Rochford R, Griffin B, Newton R, Jackson G, Menon G, Harrison CJ, Israels T, Bailey S. Burkitt's lymphoma. *Lancet*. 2012;379(9822):1234–44.
32. Bernheim A, Berger R, Lenoir G. Cytogenetic studies on Burkitt's lymphoma cell lines. *Cancer Genet Cytogenet*. 1983;8(3):223–9.
33. Pelicci PG, Knowles DM, Magrath I, Dalla-Favera R. Chromosomal breakpoints and structural alterations of the c-myc locus differ in endemic and sporadic forms of Burkitt lymphoma. *Proc Natl Acad Sci USA*. 1986;83(9):2984–8.
34. Salaverria I, Martin-Guerrero I, Wagener R, Kreuz M, Kohler CW, Richter J, Pienkowska-Grela B, Adam P, Burkhardt B, Claviez A, Damm-Welk C, Drexler HG, Hummel M, Jaffe ES, Küppers R, Lefebvre C, Lisfeld J, Löffler M, Macleod RA, Nagel I, Oschlies I, Rosolowski M, Russell RB, Rymkiewicz G, Schindler D, Schlesner M, Scholtysik R, Schwaenen C, Spang R, Szczepanowski M, Trümper L, Vater I, Wessendorf S, Klapper W, Siebert R, Molecular Mechanisms in Malignant Lymphoma Network. Project, Berlin-Frankfurt-Münster Non-Hodgkin Lymphoma Group. A recurrent 11q aberration pattern characterizes a subset of MYC-negative high-grade B-cell lymphomas resembling Burkitt lymphoma. *Blood*. 2014;123(8):1187–98.
35. Ferreiro JF, Morscio J, Dierickx D, Marcelis L, Verhoef G, Vandenbergh P, Tousseyn T, Wlodarska I. Post-transplant molecularly defined Burkitt lymphomas are frequently MYC-negative and characterized by the 11q-gain/loss pattern. *Haematologica*. 2015;100(7):e275–e9.
36. Dave SS, Fu K, Wright GW, Lam LT, Kluin P, Boerma EJ, Greiner TC, Weisenburger DD, Rosenwald A, Ott G, Müller-Hermelink HK, Gascoyne RD, Delabie J, Rimsza LM, Brazier RM, Grogan TM, Campo E, Jaffe ES, Dave BJ, Sanger W, Bast M, Vose JM, Armitage JO, Connors JM, Smeland EB, Kvaloy S, Holte H, Fisher RI, Miller TP, Montserrat E, Wilson WH, Bahl M, Zhao H, Yang L, Powell J, Simon R, Chan WC, Staudt LM, Lymphoma/Leukemia Molecular Profiling Project. Molecular diagnosis of Burkitt's lymphoma. *N Engl J Med*. 2006;354(23):2431–42.
37. Piccaluga PP, De Falco G, Kustagi M, Gazzola A, Agostinelli C, Tripodo C, Leucci E, Onnis A, Astolfi A, Sapienza MR, Bellan C, Lazzi S, Tumwine L, Mawanda M, Ogwang M, Calbi V, Formica S, Califano A, Pileri SA, Leoncini L. Gene expression analysis uncovers similarity and differences among Burkitt lymphoma subtypes. *Blood*. 2011;117(13):3596–608.
38. Schmitz R, Young RM, Ceribelli M, Jhavar S, Xiao W, Zhang M, Wright G, Shaffer AL, Hodson DJ, Buras E, Liu X, Powell J, Yang Y, Xu W, Zhao H, Kohlhammer H, Rosenwald A, Kluin P, Müller-Hermelink HK, Ott G, Gascoyne RD, Connors JM, Rimsza LM, Campo E, Jaffe ES, Delabie J, Smeland EB, Ogwang MD, Reynolds SJ, Fisher RI, Brazier RM, Tubbs RR, Cook JR, Weisenburger DD, Chan WC, Pittaluga S, Wilson W, Waldmann TA, Rowe M, Mbulaiteye SM, Rickinson AB, Staudt LM. Burkitt lymphoma pathogenesis and therapeutic targets from structural and functional genomics. *Nature*. 2012;490(7418):116–20.
39. Abate F, Ambrosio MR, Mundo L, Laginestra MA, Fuligni F, Rossi M, Zairis S, Gazaneo S, De Falco G, Lazzi S, Bellan C, Rocca BJ, Amato T, Marasco E, Etebari M, Ogwang M, Calbi V, Ndede I, Patel K, Chumba D, Piccaluga PP, Pileri S, Leoncini L, Rabadan R. Distinct viral and mutational spectrum of endemic Burkitt lymphoma. *PLoS Pathog*. 2015;11(10):e1005158.
40. Armitage JO, Weisenburger DD. New approach to classifying non-Hodgkin's lymphomas: clinical features of the major histologic subtypes. Non-Hodgkin's Lymphoma Classification Project. *J Clin Oncol*. 1998;16(8):2780–95.
41. Liu Q, Salaverria I, Pittaluga S, Jegalian AG, Xi L, Siebert R, Raffeld M, Hewitt SM, Jaffe ES. Follicular lymphomas in children and young adults: a comparison of the pediatric variant with usual follicular lymphoma. *Am J Surg Pathol*. 2013;37(3):333–43.
42. Louissaint A Jr, Ackerman AM, Dias-Santagata D, Ferry JA, Hochberg EP, Huang MS, Iafrate AJ, Lara DO, Pinkus GS, Salaverria I, Siddique Z, Siebert R, Weinstein HJ, Zukerberg LR, Harris NL, Hasserjian RP. Pediatric-type nodal follicular lymphoma: an indolent clonal proliferation in children and adults with high proliferation index and no BCL2 rearrangement. *Blood*. 2012;120(12):2395–404.
43. Salaverria I, Philipp C, Oschlies I, Kohler CW, Kreuz M, Szczepanowski M, Burkhardt B, Trautmann H, Gesk S, Andrusiewicz M, Berger H, Fey M, Harder L, Hasenclever D, Hummel M, Loeffler M, Mahn F, Martin-Guerrero I, Pellissery S, Pott C, Pfreundschuh M, Reiter A, Richter J, Rosolowski M, Schwaenen C, Stein H, Trümper L, Wessendorf S, Spang R, Küppers R, Klapper W, Siebert R, Molecular Mechanisms in Malignant Lymphomas Network Project of the Deutsche Krebshilfe, German High-Grade Lymphoma Study Group, Berlin-Frankfurt-Münster-NHL trial group. Translocations activating IRF4 identify a subtype of germinal center-derived B-cell lymphoma affecting predominantly children and young adults. *Blood*. 2011;118(1):139–47.
44. Bödör C, Grossmann V, Popov N, Okosun J, O'Riain C, Tan K, Marzec J, Araf S, Wang J, Lee AM, Clear A, Montoto S, Matthews J, Iqbal S, Rajnai H, Rosenwald A, Ott G, Campo E, Rimsza LM, Smeland EB, Chan WC, Brazier RM, Staudt LM, Wright G, Lister TA, Elemento O, Hills R, Gribben JG, Chelala C, Matolcsy A, Kohlmann A, Haferlach T, Gascoyne RD, Fitzgibbon J. EZH2 mutations are frequent and represent an early event in follicular lymphoma. *Blood*. 2013;122(18):3165–8.
45. Okosun J, Bödör C, Wang J, Araf S, Yang CY, Pan C, Boller S, Cittaro D, Bozek M, Iqbal S, Matthews J, Wrench D, Marzec J, Tawana K, Popov N, O'Riain C, O'Shea D, Carlotti E, Davies A, Lawrie CH, Matolcsy A, Calaminici M, Norton A, Byers RJ, Mein C, Stupka E, Lister TA, Lenz G, Montoto S, Gribben

- JG, Fan Y, Grosschedl R, Chelala C, Fitzgibbon J. Integrated genomic analysis identifies recurrent mutations and evolution patterns driving the initiation and progression of follicular lymphoma. *Nat Genet.* 2014;46(2):176–81.
46. Green MR, Kihira S, Liu CL, Nair RV, Salari R, Gentles AJ, Irish J, Stehr H, Vicente-Dueñas C, Romero-Camarero I, Sanchez-Garcia I, Plevritis SK, Arber DA, Batzoglou S, Levy R, Alizadeh AA. Mutations in early follicular lymphoma progenitors are associated with suppressed antigen presentation. *Proc Natl Acad Sci USA.* 2015;112(10):E1116–E25.
  47. Ambinder AJ, Shenoy PJ, Nastoupil LJ, Flowers CR. Using primary site as a predictor of survival in mantle cell lymphoma. *Cancer.* 2013;119(8):1570–7.
  48. Lee SJ, Suh CW, Lee SI, Kim WS, Lee WS, Kim HJ, Choi CW, Kim JS, Shin HJ, Consortium for Improving Survival of Lymphoma. Clinical characteristics, pathological distribution, and prognostic factors in non-Hodgkin lymphoma of Walden's ring: nationwide Korean study. *Korean J Intern Med.* 2014;29(3):352–60.
  49. Bracci PM, Benavente Y, Turner JJ, Paltiel O, Slager SL, Vajdic CM, Norman AD, Cerhan JR, Chiu BC, Becker N, Cocco P, Dogan A, Nieters A, Holly EA, Kane EV, Smedby KE, Maynadié M, Spinelli JJ, Roman E, Glimelius B, Wang SS, Sampson JN, Morton LM, de Sanjosé S. Medical history, lifestyle, family history, and occupational risk factors for marginal zone lymphoma: the InterLymph Non-Hodgkin Lymphoma Subtypes Project. *J Natl Cancer Inst Monogr.* 2014;2014(48):52–65.
  50. Caballero D, Campo E, López-Guillermo A, Martín A, Arranz-Sáez R, Giné E, López A, González-Barca E, Canales MÁ, González-Díaz M, Orfao A. Clinical practice guidelines for diagnosis, treatment, and follow-up of patients with mantle cell lymphoma. Recommendations from the GEL/TAMO Spanish Cooperative Group. *Ann Hematol.* 2013;92(9):1151–79.
  51. Nakashima MO, Durkin L, Bodo J, Lin J, Quintanilla-Martinez L, Fu K, Hsi ED. Utility and diagnostic pitfalls of SOX11 monoclonal antibodies in mantle cell lymphoma and other lymphoproliferative disorders. *Appl Immunohistochem Mol Morphol.* 2014;22(10):720–7.
  52. Wlodarska I, Dierickx D, Vanhentenrijk V, Van Roosbroeck K, Pospíšilová H, Minnei F, Verhoef G, Thomas J, Vandenberghe P, De Wolf-Peeters C. Translocations targeting CCND2, CCND3, and MYCN do occur in t(11;14)-negative mantle cell lymphomas. *Blood.* 2008;111(12):5683–90.
  53. Salaverria I, Royo C, Carvajal-Cuenca A, Clot G, Navarro A, Valera A, Song JY, Woroniecka R, Rymkiewicz G, Klapper W, Hartmann EM, Sujobert P, Wlodarska I, Ferry JA, Gaulard P, Ott G, Rosenwald A, Lopez-Guillermo A, Quintanilla-Martinez L, Harris NL, Jaffe ES, Siebert R, Campo E, Beà S. CCND2 rearrangements are the most frequent genetic events in cyclin D1(-) mantle cell lymphoma. *Blood.* 2013;121(8):1394–402.
  54. Beà S, Valdés-Mas R, Navarro A, Salaverria I, Martín-García D, Jares P, Giné E, Pinyol M, Royo C, Nadeu F, Conde L, Juan M, Clot G, Vizán P, Di Croce L, Puente DA, López-Guerra M, Moros A, Roue G, Aymerich M, Villamor N, Colomo L, Martínez A, Valera A, Martín-Subero JI, Amador V, Hernández L, Rozman M, Enjuanes A, Forcada P, Muntanola A, Hartmann EM, Calasanz MJ, Rosenwald A, Ott G, Hernández-Rivas JM, Klapper W, Siebert R, Wiestner A, Wilson WH, Colomer D, López-Guillermo A, López-Otín C, Puente XS, Campo E. Landscape of somatic mutations and clonal evolution in mantle cell lymphoma. *Proc Natl Acad Sci USA.* 2013;110(45):18250–5.
  55. Wright D, McKeever P, Carter R. Childhood non-Hodgkin lymphomas in the United Kingdom: findings from the UK Children's Cancer Study Group. *J Clin Pathol.* 1997;50(2):128–34.
  56. Arber DA, Orazi A, Hasserjian R, Thiele J, Borowitz MJ, Le Beau MM, Bloomfield CD, Cazzola M, Vardiman JW. The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. *Blood.* 2016;127(20):2391–405.
  57. Coustan-Smith E, Mullighan CG, Onciu M, Behm FG, Raimondi SC, Pei D, Cheng C, Su X, Rubnitz JE, Basso G, Biondi A, Pui CH, Downing JR, Campana D. Early T-cell precursor leukaemia: a subtype of very high-risk acute lymphoblastic leukaemia. *Lancet Oncol.* 2009;10(2):147–56.
  58. Zhang J, Ding L, Holmfeldt L, Wu G, Heatley SL, Payne-Turner D, Easton J, Chen X, Wang J, Rusch M, Lu C, Chen SC, Wei L, Collins-Underwood JR, Ma J, Roberts KG, Pounds SB, Ulyanov A, Becksfort J, Gupta P, Huether R, Kriwacki RW, Parker M, McGoldrick DJ, Zhao D, Alford D, Espy S, Bobba KC, Song G, Pei D, Cheng C, Roberts S, Barbato MI, Campana D, Coustan-Smith E, Shurtleff SA, Raimondi SC, Kleppe M, Cools J, Shimano KA, Hermiston ML, Doulatov S, Eppert K, Laurenti E, Notta F, Dick JE, Basso G, Hunger SP, Loh ML, Devidas M, Wood B, Winter S, Dunsmore KP, Fulton RS, Fulton LL, Hong X, Harris CC, Dooling DJ, Ochoa K, Johnson KJ, Obenauer JC, Evans WE, Pui CH, Naeve CW, Ley TJ, Mardis ER, Wilson RK, Downing JR, Mullighan CG. The genetic basis of early T-cell precursor acute lymphoblastic leukaemia. *Nature.* 2012;481(7380):157–63.
  59. Neumann M, Heesch S, Schlee C, Schwartz S, Gökbuget N, Hoelzer D, Konstantin NP, Ksienzyk B, Vosberg S, Graf A, Krebs S, Blum H, Raff T, Brüggemann M, Hofmann WK, Hecht J, Bohlander SK, Greif PA, Baldus CD. Whole-exome sequencing in adult ETP-ALL reveals a high rate of DNMT3A mutations. *Blood.* 2013;121(23):4749–52.
  60. Neumann M, Coskun E, Fransecky L, Mochmann LH, Bartram I, Sartangi NF, Heesch S, Gökbuget N, Schwartz S, Brandts C, Schlee C, Haas R, Dührsen U, Griesshammer M, Döhner H, Ehninger G, Burmeister T, Blau O, Thiel E, Hoelzer D, Hofmann WK, Baldus CD. FLT3 mutations in early T-cell precursor ALL characterize a stem cell like leukemia and imply the clinical use of tyrosine kinase inhibitors. *PLoS One.* 2013;8(1):e53190.
  61. Gaux C, Cools J, Michaux L, Vandenberghe P, Hagemeijer A. Cytogenetics and molecular genetics of T-cell acute lymphoblastic leukemia: from thymocyte to lymphoblast. *Leukemia.* 2006;20(9):1496–510.
  62. Van Vlierberghe P, Ferrando A. The molecular basis of T cell acute lymphoblastic leukemia. *J Clin Invest.* 2012;122(10):3398–406.
  63. Ohgami RS, Arber DA, Zehnder JL, Natkunam Y, Warnke RA. Indolent T-lymphoblastic proliferation (iT-LBP): a review of clinical and pathologic features and distinction from malignant T-lymphoblastic lymphoma. *Adv Anat Pathol.* 2013;20(3):137–40.
  64. Isaacson PG, Du MQ. MALT lymphoma: from morphology to molecules. *Nat Rev Cancer.* 2004 Aug;4(8):644–53.
  65. Suh C, Huh J, Roh JL. Extranodal marginal zone B-cell lymphoma of mucosa-associated lymphoid tissue arising in the extracranial head and neck region: a high rate of dissemination and disease recurrence. *Oral Oncol.* 2008;44(10):949–55.
  66. Wenzel C, Fiebig W, Dieckmann K, Formanek M, Chott A, Raderer M. Extranodal marginal zone B-cell lymphoma of mucosa-associated lymphoid tissue of the head and neck area: high rate of disease recurrence following local therapy. *Cancer.* 2003;97(9):2236–41.
  67. Falini B, Agostinelli C, Bigerna B, Pucciarini A, Pacini R, Tabarrini A, Falcinelli F, Piccioli M, Paulli M, Gambacorta M, Ponzoni M, Tiacchi E, Ascani S, Martelli MP, Dalla Favera R, Stein H, Pileri SA. IRTA1 is selectively expressed in nodal and extranodal marginal zone lymphomas. *Histopathology.* 2012;61(5):930–41.

68. Nocturne G, Mariette X. Sjögren Syndrome-associated lymphomas: an update on pathogenesis and management. *Br J Haematol*. 2015;168(3):317–27.
69. Chanudet E, Ye H, Ferry J, Bacon CM, Adam P, Müller-Hermelink HK, Radford J, Pileri SA, Ichimura K, Collins VP, Hamoudi RA, Nicholson AG, Wotherspoon AC, Isaacson PG, Du MQ. A20 deletion is associated with copy number gain at the TNFA/B/C locus and occurs preferentially in translocation-negative MALT lymphoma of the ocular adnexa and salivary glands. *J Pathol*. 2009;217(3):420–30.
70. Streubel B, Simonitsch-Klupp I, Müllauer L, Lamprecht A, Huber D, Siebert R, Stolte M, Trautinger F, Lukas J, Püspök A, Formanek M, Assanasen T, Müller-Hermelink HK, Cerroni L, Raderer M, Chott A. Variable frequencies of MALT lymphoma-associated genetic aberrations in MALT lymphomas of different sites. *Leukemia*. 2004;18(10):1722–6.
71. Arcaini L, Burcheri S, Rossi A, Paulli M, Bruno R, Passamonti F, Brusamolino E, Molteni A, Pulsoni A, Cox MC, Orsucci L, Fabbri A, Frezzato M, Voso MT, Zaja F, Montanari F, Merli M, Pascutto C, Morra E, Cortelazzo S, Lazzarino M. Prevalence of HCV infection in nongastric marginal zone B-cell lymphoma of MALT. *Ann Oncol*. 2007;18(2):346–50.
72. Ochoa ER, Harris NL, Pilch BZ. Marginal zone B-cell lymphoma of the salivary gland arising in chronic sclerosing sialadenitis (Küttner tumor). *Am J Surg Pathol*. 2001;25(12):1546–50.
73. Cheuk W, Yuen HK, Chan AC, Shih LY, Kuo TT, Ma MW, Lo YF, Chan WK, Chan JK. Ocular adnexal lymphoma associated with IgG4+ chronic sclerosing dacryoadenitis: a previously undescribed complication of IgG4-related sclerosing disease. *Am J Surg Pathol*. 2008;32(8):1159–67.
74. Delecluse HJ, Anagnostopoulos I, Dallenbach F, Hummel M, Marafioti T, Schneider U, Huhn D, Schmidt-Westhausen A, Reichart PA, Gross U, Stein H. Plasmablastic lymphomas of the oral cavity: a new entity associated with the human immunodeficiency virus infection. *Blood*. 1997;89(4):1413–20.
75. Castillo JJ, Bibas M, Miranda RN. The biology and treatment of plasmablastic lymphoma. *Blood*. 2015;125(15):2323–30.
76. Valera A, Balagué O, Colomo L, Martínez A, Delabie J, Tadmeh-Heath L, Jaffe ES, Campo E. IG/MYC rearrangements are the main cytogenetic alteration in plasmablastic lymphomas. *Am J Surg Pathol*. 2010;34(11):1686–94.
77. Boy SC, van Heerden MB, Babb C, van Heerden WF, Willem P. Dominant genetic aberrations and coexistent EBV infection in HIV-related oral plasmablastic lymphomas. *Oral Oncol*. 2011;47(9):883–7.
78. Nicholson HS, Egeler RM, Nesbit ME. The epidemiology of Langerhans cell histiocytosis. *Hematol Oncol Clin North Am*. 1998;12(2):379–84.
79. Irving RM, Broadbent V, Jones NS. Langerhans' cell histiocytosis in childhood: management of head and neck manifestations. *Laryngoscope*. 1994;104(1):64–70.
80. Pileri SA, Grogan TM, Harris NL, Banks P, Campo E, Chan JK, Favera RD, Delsol G, De Wolf-Peters C, Falini B, Gascoyne RD, Gaulard P, Gatter KC, Isaacson PG, Jaffe ES, Kluin P, Knowles DM, Mason DY, Mori S, Müller-Hermelink HK, Piris MA, Ralfkiaer E, Stein H, Su JJ, Warnke RA, Weiss LM. Tumours of histiocytes and accessory dendritic cells: an immunohistochemical approach to classification from the International Lymphoma Study Group based on 61 cases. *Histopathology*. 2002;41(1):1–29.
81. Badalian-Very G, Vergilio JA, Degar BA, MacConaill LE, Brandner B, Calicchio ML, Kuo FC, Ligon AH, Stevenson KE, Kehoe SM, Garraway LA, Hahn WC, Meyerson M, Fleming MD, Rollins BJ. Recurrent BRAF mutations in Langerhans cell histiocytosis. *Blood*. 2010;116(11):1919–23.
82. Brown NA, Furtado LV, Betz BL, Kiel MJ, Weigelin HC, Lim MS, Elenitoba-Johnson KS. High prevalence of somatic MAP2K1 mutations in BRAF V600E-negative Langerhans cell histiocytosis. *Blood*. 2014;124(10):1655–8.
83. Pileri SA, Ascani S, Cox MC, Campidelli C, Bacci F, Piccioli M, Piccaluga PP, Agostinelli C, Asioli S, Novero D, Bisceglia M, Ponzoni M, Gentile A, Rinaldi P, Franco V, Vincelli D, Pileri A Jr, Gasbarra R, Falini B, Zinzani PL, Baccarani M. Myeloid sarcoma: clinico-pathologic, phenotypic and cytogenetic analysis of 92 adult patients. *Leukemia*. 2007;21(2):340–50.
84. Zhou J, Bell D, Medeiros LJ. Myeloid sarcoma of the head and neck region. *Arch Pathol Lab Med*. 2013;137(11):1560–8.
85. Falini B, Lenze D, Hasserjian R, Coupland S, Jaehne D, Soupir C, Liso A, Martelli MP, Bolli N, Bacci F, Pettrossi V, Santucci A, Martelli MF, Pileri S, Stein H. Cytoplasmic mutated nucleophosmin (NPM) defines the molecular status of a significant fraction of myeloid sarcomas. *Leukemia*. 2007;21(7):1566–70.