

CCN proteins: A centralized communication network

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Abstract The CCN family of proteins includes six members presently known as CCN1, CCN2, CCN3, CCN4, CCN5 and CCN6. These proteins were originally designated CYR61, CTGF, NOV, and WISP-1, WISP-2, WISP-3. Although these proteins share a significant amount of structural features and a partial identity with other large families of regulatory proteins, they exhibit different biological functions. A critical examination of the progress made over the past two decades, since the first CCN proteins were discovered brings me to the conclusion that most of our present knowledge regarding the functions of these proteins was predicted very early after their discovery. In an effort to point out some of the gaps that prevent us to reach a comprehensive view of the functional interactions between CCN proteins, it is necessary to reconsider carefully data that was already published and put aside, either because the scientific community was not ready to accept them, or because they were not fitting with the « consensus » when they were published. This review article points to avenues that were not attracting the attention that they deserved. However, it is quite obvious that the six members of this unique family of tetra-modular proteins must act in concert, either simultaneously or sequentially, on the same sites or at different times in the life of living organisms. A better understanding of the spatio-temporal regulation of CCN proteins expression requires considering the family as such, not as a set of single proteins related only by their name. As proposed in this review, there is enough convincing pieces of evidence, at the present time, in favor of these proteins playing a role in the coordination of multiple signaling pathways, and constituting a Centralized Communication Network. Deciphering the hierarchy of regulatory circuits involved in

this complex system is an important challenge for the near future. In this article, I would like to briefly review the concept of a CCN family of proteins and critically examine the progress made over the past 10 years in the understanding of their biological functions and involvement in both normal and pathological processes.

Keywords CCN · CCN proteins · CTGF · CYR61 · NOV · WISP · Inflammation · Transformation · Cell proliferation · Cell signaling · Cell communication · Centralized communication network · Development · Differentiation

Brief historical and structural context

It is always useful to recall the facts that set the ground rules for current ideas in fields where new comers are not always aware of the succession of events that underlie the prevailing concepts. The history of Science contains many examples of major leaders who have seen their contribution vanish over time.¹

The CCN story began in the early 90's with the discovery in mouse, human and chicken of three proteins that were designated CYR61 (for « Cystein Rich »), CTGF (for Connective Tissue Growth Factor) and NOV (for Nephroblastoma Overexpressed) (O'Brien et al. 1990; Bradham et al. 1991; Joliot et al. 1992).

The primary sequence of the genes encoding these proteins revealed that they were very closely related and that they encoded proteins that share partial identity with other major classes of regulatory proteins.

From the predicted amino acid sequences of the proteins, it was also concluded that each of these three proteins was a mosaic assembly of three structural domains that shared

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¹ For example, who still remembers the pivotal contribution of JC Chermann in the identification of the HIV as AIDS associated virus see « <http://nobelchermann.com/> »

partial identity with the N terminus of IGF binding proteins, the type C repeat of the Von Willebrand factor, the type 1 repeat of thrombospondin and a fourth domain, that contained a set of eight cysteines that were shown to form the so-called cystine knot that is present in a whole range of regulatory proteins including growth factors and other secreted proteins. The presence of a sequence encoding a typical signal peptide at the N-terminus of these proteins, strongly suggested that they were secreted, a prediction that was indeed later experimentally confirmed (Fig. 1).

Based on these structural features unique to these three proteins, P. Bork (Bork 1993) considered that they should constitute a new family of proteins which he called the « CCN family of proteins ».

A few years later, with the birth of the International CCN Society, we proposed that the proteins belonging to this family be designated with the CCN acronym followed by a number indicating the chronological order in which they were discovered (Brigstock et al. 2003) (Fig. 2).

Both CCN1 (CYR 61) and CCN2 (CTGF) were induced by cell proliferation (Lau & Nathans, 1985, 1987; Simmons et al., 1989, Brunner et al., 1991, Ryseck et al., 1991), whereas CCN3 (NOV) was inhibited when cells were induced to proliferate. In other words, the genes encoding CCN1 and CCN2 could be classified as Immediate Early Genes, whereas CCN3 was not (Joliot et al. 1992; Scholz et al. 1996).

The fact that NOV was not subject to the same regulatory circuits constituted the very first evidence for CCN3 having distinct biological properties.

The second major difference between CCN3 and the CCN1/CCN2 pair stemmed from the capacity of CCN3 to inhibit cell proliferation and induce cellular transformation when it was truncated at its aminoterminal (Joliot et al. 1992).

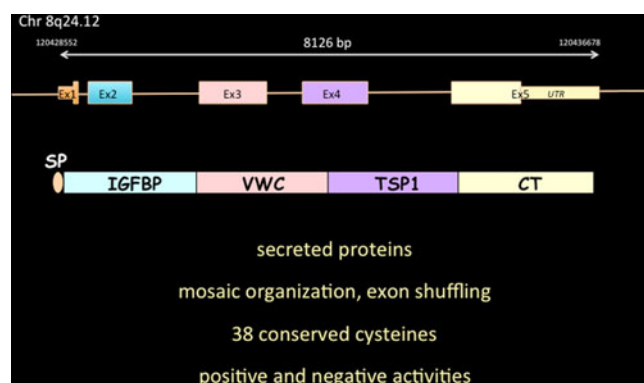


Fig. 1 The prototypic CCN3 protein. The schematic organization of CCN3 exons on the human genome is represented with the corresponding structural domains contained in the full length version of CCN3. SP is for signal peptide. Salient features of CCN proteins are also indicated. See text for details

New nomenclature 2003 *Mol Pathol.* 56(2):127-128.

1p22	CCN 1	Cyr61	cystein rich
6q23	CCN 2	Ctgf	connective tissue growth factor
8q24.1	CCN 3	Nov	nephroblastoma overexpressed
8q24.2	CCN 4	Wisp1	} Wnt-induced Secreted proteins
20q13.1	CCN 5	Wisp2	
6q21-22	CCN 6	Wisp3	

Fig. 2 The CCN family of proteins in human

The inhibition of cell proliferation by CCN3 contrasted with the stimulatory effects of CCN1 and CCN2, therefore suggesting that, although these three proteins might belong to the same group, they were representing both positive and negative regulators of cell proliferation. These observations also argued against the possibility that the three CCN proteins had redundant functions (Perbal 2001).

There are many other examples of protein families containing both positive and negative effectors and it makes a lot of sense for cells to make use of both types of signals to smoothly regulate their biological behavior.

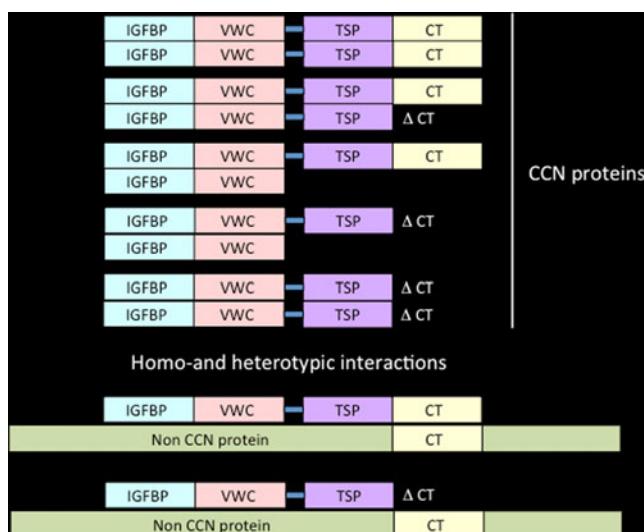
At this stage in the understanding, we could already make the following predictions : 1) as CCN proteins contain a consensus signal peptide, they are likely to be secreted, 2) based on the presence of the IGFBP-like module at their N terminus CCN proteins could possibly interact with IGF signaling, 3) CCN proteins probably exist as large multimeric complexes that regulate cell adhesion and proliferation, via their modules 2 and 3, and 4) the cystine knot contained in their C-terminal module is responsible for the formation of homo and heterodimers.

The presence of a hinge region between the second and third modules of the CCN proteins suggested possible translational processing by proteolytic digestion (Perbal et al. 1999, 2003), as also discussed by D. Brigstock, L. Lau and myself, at the first International Workshop on the family of Genes in Saint-Malo (Ayer-Lelievre et al. 2001)

Interestingly, the amino-truncated form of CCN3 that was expressed in one chicken nephroblastoma cell line, as the result of a MAV1-proviral insertion in the nov gene (Perbal 1994, 1995) was shown to induce morphological transformation of chicken fibroblasts when expressed under the control of a RSV-derived promoter (Joliot et al. 1992). On the contrary, the full length protein expressed in the same conditions induced cell growth arrest.

These observations suggested that the inhibitory effect of CCN3 on cell proliferation required the expression of a full

Since the rCOP-1 protein lacks the C-terminal module of the canonical CCN family members, one can expect it to act as a « dominant negative » effector in the homo and heterotropic multimerization of CCN proteins, either with other members of the CCN family or with proteins which do not



A number of sound reviews have addressed key aspects of the theme since 1999, and have blossomed over the past decade (Brigstock 1999, 2003; Lau and lam 1999; Perbal 2001, 2003, 2004, 2006a; Blom et al. 2002; Planque and Perbal 2003; Bleau et al. 2005; Chaqour and Goppelt-Struebe 2006; Leask and Abraham 2006; Kubota and

Tagigawa 2007; Kleer et al. 2007; Holbourn et al. 2008; Shi-Wen et al. 2008; Hall-Glenn and Lyons 2011, Jun and Lau 2011; Lau 2011; Arnott et al. 2011; Ouellet and Siegel 2012; Mason 2013). Those who are interested in the biological functions of CCN proteins should go back to these articles which contain thoughtful considerations, models, and sometimes provocative ideas. Many views discussed in these articles still represent challenges that should be addressed in the light of the widening interest of the scientific community for the field.

Considering the tetramodular structure of the CCN proteins, it was not much of a surprise that they were found to participate in many essential biological functions including cell communication, control of proliferation, adhesion and migration, regulation of growth, development and differentiation, wound healing, regeneration, and cell death (Yeger and Perbal 2007) (Fig. 4).

In order to better understand the antiproliferative activity of CCN3 that we had established in avian primary chicken embryo fibroblasts we first studied the effects on CCN3 on the progression of cell cycle.

Our initial measurements did not permit us to establish whether CCN3 expression modified the ratio of cell subpopulations at various phases of the cell cycle. When synchronized cell cultures were used, we could establish that the expression of CCN3 interfered with the S/G2 transition of the cell cycle, thereby inducing an artificial accumulation of cells at the S phase (Bleau et al. 2007). These results not only confirmed the antiproliferative activity of CCN3 but they permitted reconciling our present observation with a set of results published by another group who claimed that CCN3 stimulated cell growth, based on BrdU incorporation (Perbal 2008) (Fig. 5).

In spite of our demonstration that the increase in BrdU incorporation was a direct result of the CCN3 inhibitory effect on cell growth, it is still common to find in the

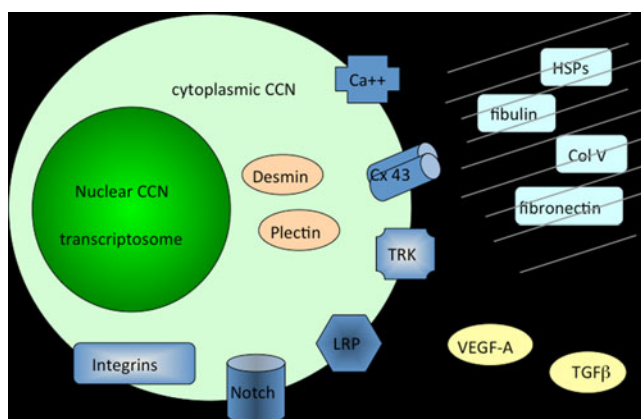


Fig. 4 Schematic drawing of CCN proteins partners. Interactions of CCN proteins with other ligands and regulators are shown here to occur in the extracellular matrix, at the cell membrane and inside the cytoplasm and the nucleus of cells

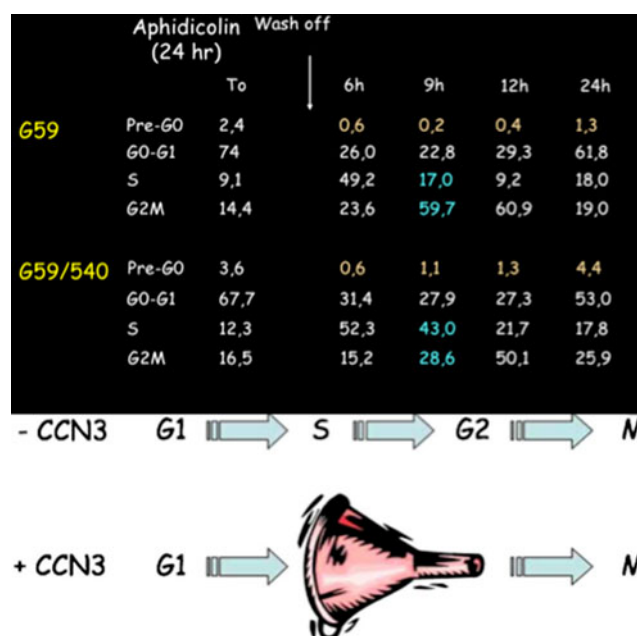


Fig. 5 Inhibition of cell proliferation by CCN3 results from a break at the S/G2 transition in the cell cycle. Synchronized cells that express CCN3 accumulate at the S phase. As a result the cells go less efficiently through the cycle. Data are from Bleau et al. 2007 (see text)

literature, groups who claim that CCN3 shows growth stimulatory properties (based on the BrdU paper !).

The antiproliferative activity of CCN3 has been confirmed in many different cell types, both by our group and others (Gupta et al. 2001; Sakamoto et al. 2002; McCallum et al. 2006; Bleau et al. 2007; Benini et al. 2005; Fukunaga-Kalabis et al. 2006; Planque et al. 2006; van Roeyen et al. 2008; Vallacchi et al. 2008; Shimoyama et al. 2010; Lin et al. 2010). Experiments performed in our laboratory also established that the inhibitory effects on cell growth do not require the two first domains of CCN3 (Planque et al. 2006) (Fig. 6).

Since CCN5 which lacks the CT domain also shows a growth inhibitory effect it is tempting to assign the growth inhibition property to the TSP1 domain of CCN proteins but this needs to be confirmed experimentally.

Along the same line, we could establish that CCN3 also shows an anti-tumorigenic activity, in choriocarcinomas, glioblastomas and Ewing's tumors (Gellhaus et al. 2004; Gupta et al. 2001; Benini et al. 2005). In the case of Ewing's and osteosarcomas, we could also establish that CCN3 is a marker of poor prognosis. Out of 45 patients with primary Ewing's tumors, those who did not express CCN3 in the primary tumors did not develop metastases whereas, 50 % of the patients with primary tumors positive for CCN3 developed metastatic tumors (Manara et al. 2002).

Similar observations were made in our studies on osteosarcomas where we could establish that expression of CCN3 significantly reduced survival (Perbal et al. 2008).

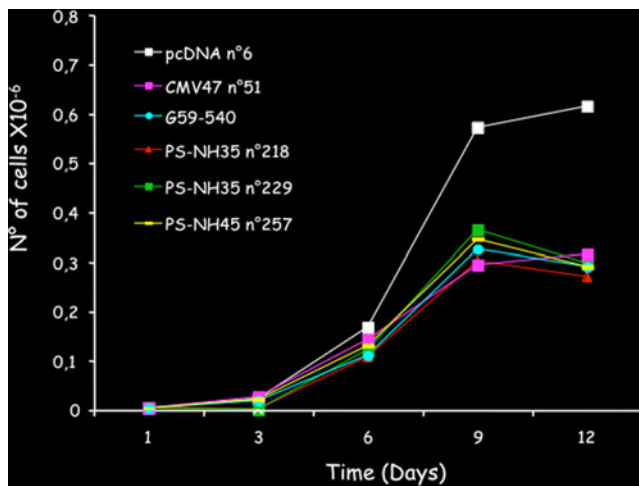


Fig. 6 The C-terminal half of CCN3 is sufficient to promote antiproliferative activity. The drawing shows the growth curves of stably transfected cell lines constructs expressing either the full length CCN3 protein CMV47 and G59-540, the three first domains of CCN3 (NH35), or the C-terminal half of CCN3 (NH45). All constructs induce a dramatic inhibition of cell proliferation. The effects obtained with the NH45 transfected cells indicated that the inhibitory potential is contained within the last two domains of CCN3. Data are from Bleau et al. 2007 (see text)

Pathological conditions are often seen as useful to decipher the role(s) of proteins whose functions may be altered by the environment.

However, one should keep in mind that the protein functions that are uncovered in the context of a given pathological condition might not be comparable to the functions of the same protein in other pathological conditions or in normal conditions.

Thresholds are critical in many instances and the balance between negative and positive signals that is maintained in a normal context might be disrupted in a completely aberrant way in the pathological context.

An illustration of this complexity was shown in a set of two collaborative approaches where the results obtained with skin reconstructs were integrated, and analyzed in the light of the situation encountered in melanomas (Fukunaga-Kalabis et al. 2006, 2008; Vallacchi et al. 2008).

First, the use of skin reconstructs established that CCN3 was essential for the 3D localisation of melanocytes at the basal membrane of normal skin, suggesting that a dysruption of the normal crosstalk between melanocytes and keratinocytes would result in a dermal invasion by melanocytes comparable to the situation that occurs in melanomas. Indeed, CCN3 was found to be inversely correlated with melanoma invasion, as measured in a set of human melanocytic nevi and melanoma samples.

However, in another set of samples, the metastatic potential of melanomas was directly related to the overexpression

of CCN3 and the survival rate was much reduced in patients expressing the nuclear truncated CCN3 species previously identified in other aggressive tumors.

These observations highlighted the difficulties in simply relating the expression of a marker such as CCN3 to the complex biology of tumors, even though the levels of CCN3 expression could be used as an indicator for the staging of tumors and in some cases for a molecularly based therapeutics.

CCN proteins: matricellular proteins only?

Most recently published articles quote in their introductory comments, that CCN proteins are secreted and play their biological functions as essential components of the extracellular matrix. Indeed, for the past decade, the « scientifically-correct » way of thinking in the CCN community, was focused on the these proteins being « matricellular proteins ».

Although there is no doubt that some of the CCN proteins are associated to the extracellular matrix (ECM), the situation may not be that simple.

First of all, published data regarding subcellular localization of the CCN proteins clearly indicate that the distribution of the CCN proteins is not similar for all members of the family. For example, CCN2 is barely detected in the cell culture medium (Ball et al. 1998) and is mainly detected at the cell membrane, whereas newly synthesized CCN1 is not heavily detected in the culture medium, as it associates quickly with the ECM where it appears to be stabilized (half-life of greater than 24 h as compared to 30 min for intracellular and cell surface associated CCN1 (Yang and Lau 1991). We have established that CCN3 is efficiently secreted and detected as a long half-life protein in conditioned medium from CCN3 positive cells. It is also detected at the cell membrane and in the ECM (Joliot et al. 1992, Kyurkchiev et al. 2004).

The subcellular localization of rCop-1 is also informative, as it was neither detected in the conditioned medium of infected cells nor in the ECM, and it appeared to be retained in the cytoplasm of cells (Zhang et al. 1998). These data should be taken in careful consideration as it may inform us not only about the relative importance of each structural module in the secretion process, but also about the fate of the CCN proteins when they are not secreted.

Considering the major influence of the complex environment that constitutes the ECM, and the potential physical and functional interactions of CCN proteins with other matricellular proteins, is it wise and meaningful to study CCN protein function(s) by using a soluble semi-purified protein preparation or to use a test outside of a reconstructed matrix environment when one addresses the matricellular functions of CCN proteins?

The studies that we had performed to better understand the transforming properties of CCN3 in avian nephroblastomas led us to uncover an unexpected aspect of the biological functions for CCN proteins.

First of all, the nucleus of several tumor cell lines stained positive when immunostaining was performed with a CCN3-specific antibody that was raised against the C-terminus of the protein (Perbal 1999, 2004, 2006b). In spite of a whole set of additional controls that were performed in response to reviewers' comments, the CCN community did not receive this piece of data with enthusiasm. Several other growth factors and receptors had been previously shown to reside in the nucleus of cells but obviously, some of the CCN cell biologists did not like to see « secreted » proteins partitioning between the ECM and the nucleus ...

At the same time, IGF binding proteins were also detected in the nucleus. The response of the IGF community was much more positive and a few groups addressed very nicely the problem. It is important to remember, at that stage in the investigations, IGFBPs and CCN proteins were found to share a partial identity at their aminoterminal ends.

Those who are interested in these considerations should read the communication of Professor Rob Baxter in this issue.

A few years after our initial report, a nuclear form of CCN2 was also described (Wahab et al. 2001). From what I understood, and in spite of the high quality of the data that was presented, the publication of this manuscript was not easy.

Other examples of nuclear CCN1, CCN2, CCN3 and CCN5 proteins were reported (Hirschfeld et al. 2009; Sha and Leask 2011; Wiesman et al. 2010; Rittié et al. 2011), but it seems that nuclear CCN proteins do not trigger much interest in the CCN community in spite of accumulating evidence that attributes to nuclear proteins a role in the control of transcription.

A careful examination of the truncated CCN3 protein that was identified in one of the MAV1-induced avian nephroblastoma, led to the conclusion that the aminotruncation of CCN3 might affect the fate of the protein within the tumor cells. Indeed, we could demonstrate that the proviral insertion within the *ccn3* gene removed the sequences encoding the signal peptide, and resulted in a CCN3 protein that was addressed to the nucleus (Planque et al. 2006).

Further experiments established that although it does not contain a typical nuclear localisation signal, the C-terminal module of CCN3 was responsible for the nuclear addressing of CCN3 deprived of its signal peptide (Planque et al. 2006).

One possible interpretation is that the truncated CCN3 protein associates with a carrier that belongs to the group of proteins which contain a cystine knot similar to the one contained in the CT module of CCN3 (see Fig. 3).

In support of this hypothesis it is worth noting that imunogold staining helped to identify CCN3 protein at some nucleus pores, and a careful examination of the images indicated that the gold label was not associated with a vesiculated structure (Thomopoulos et al. 2001). Of course further studies are required to firmly establish these conclusions.

Future directions

This critical overview brings me to address a few questions that are still open and that constitute, in my eyes, very promising fields for future studies.

A single protein and/or complexes ?

Many of us have faced situations where several unexpected protein « bands » are detected on a blot ! Of course there are basic explanations for this result, including the poor quality of the reagents used, a problem that was addressed several times during the CCN workshops by our colleagues. For example, L. Lau raised the need to distribute reference samples of tested reagents and G. Fisher proposed an « ICCNS qualification » that could be given to satisfactory batches of reagents (commercially available or not) once they are tested by reference laboratories.

The use of good reagents also leads to the immunodetection of several other relevant bands in addition to the major canonical one described in the literature.

When this issue was raised at the first CCN Workshop, many participants in the audience admitted that they also observed, larger and shorter bands, that were running at similar apparent molecular weights in different laboratories, using different sources of antibodies and samples.

Although there is no published thorough study of these extra bands, it is generally accepted that they may represent CCN-related proteins. To my knowledge, there is at least one unpublished, as yet, proof of evidence that links a high molecular weight band to the canonical form of a CCN protein.

Ignoring these extra bands is not a satisfactory way of dealing with that important question.

Considering the structural features of the CCN proteins and the presence of sequences known to generate high molecular weight complexes in other proteins, it is possible that complexes resistant to the denaturing agents that are usually used to run gels, are formed between CCN proteins and other components, with the possibility that covalent links might even be involved in some cases.

In my opinion, the existence of CCN isoforms (Perbal 2004, 2009) is an aspect of the CCN biology that has been underestimated for years, and one that will need to be addressed. As post translational modifications of CCN proteins other than glycosylation will be uncovered, it will

become obvious that they are key aspects of the regulatory events governing the biology of these proteins.

The implications of post translational modifications in the regulation of protein interactions with receptors and other partners is well documented in other systems. Recognizing their importance in the CCN field will certainly have profound consequences on the understanding of CCN protein biology

Interactivity and cross regulation

Cross regulation between expression of two CCN proteins was first observed by ChangLong Li in my laboratory, who reported that the induction of CCN2 in adrenocortical tumor cells which express high levels of CCN3, resulted in a dramatic down regulation of CCN3 expression.

Based on this observation, a collaborative study performed with the laboratory of B. Riser confirmed and extended that observation, in establishing that the expression of CCN2 and CCN3 was inversely correlated and that expression of CCN3 inhibited CCN2 (Riser et al. 2009).

Interestingly, CCN2 knocked-down cells express high levels of CCN3, and the antagonistic effects of CCN2 (stimulation of cell growth) and CCN3 (inhibition of cell growth) are key factors in bone and cartilage differentiation (Kawaki et al. 2008).

The isolation of the first true CCN3 Knock Out mice (Shimoyama et al. 2010; Perbal 2007) will permit to better decipher the functional relationships between the CCN2 and CCN3 proteins.

The inverse functional relationship between these two CCN proteins is quite important in the light of their documented opposite effects on cell proliferation and differentiation in various cellular systems.

Indeed, future therapeutic approaches based on CCN protein targeting should take these observations into account in order to selectively alter the expression of CCN2 or CCN3.

One or more receptors? The concept of a Centralized Communication Network (CCN)

In the same vein, the search for a CCN receptor has suffered from a lack of « open minded » approaches. Evidence accumulated quickly in favor of the CCN proteins interacting not with a single receptor, but with a whole variety of receptors on the cell membrane.

The question now is, not to decide whether these represent true receptors, but rather to establish whether the CCN proteins that were shown to interact with receptor A and B are binding to these two at the same time and in the same

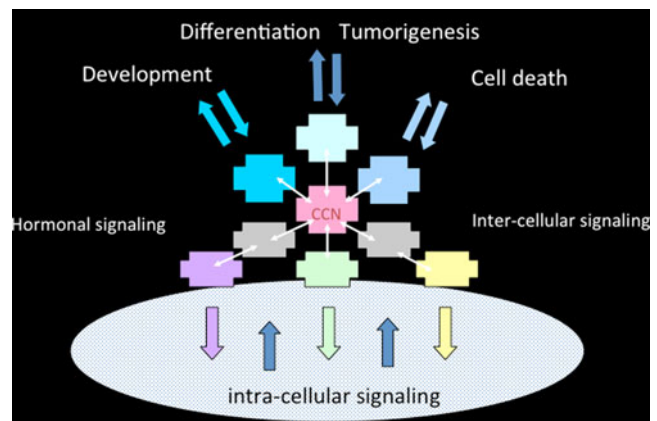


Fig. 7 A model for the CCN Centralized Communication Network. In this model, CCN proteins constitute the control center of a complex array of interactions which are part of several signaling pathways whose action must be tightly coordinated to provoke responses that must be adapted to the variations of the surrounding microenvironment and on another scale, to the variations of the outside medium in which organisms evolve. The coordination of these pathways by CCN proteins is based on functional interactions that have been reported for example with Integrins, Bone Morphogenic Proteins, Notch1, Calcium channels, fibulin 1C, Wnt, Heparan sulfate proteoglycans, decorin, TGFbeta, Tyrosine receptor kinase A, Low-density lipoprotein receptor related protein, and CCN proteins themselves

environment. The simultaneous interaction of CCN proteins with more than one receptor might allow establishing a physical connection, hence a physical support to the coordination, of several distinct regulatory pathways.

Along with this view I had previously proposed that the tetramodular structure of the CCN proteins allows them to engage with a large range of effectors (Perbal 2001), and thereby to act as scaffolds permitting a coordinated and integrated control of pathways that are activated towards the same end point, such as differentiation, growth, cell movement, etc. (Perbal and Perbal 2007) (Fig. 7).

It is more and more obvious now that the interactions of CCN proteins with a multitude of effectors involved in the regulation of key signaling pathways, place the CCN proteins at the center of a Centralized Communication Network (Fig. 8).

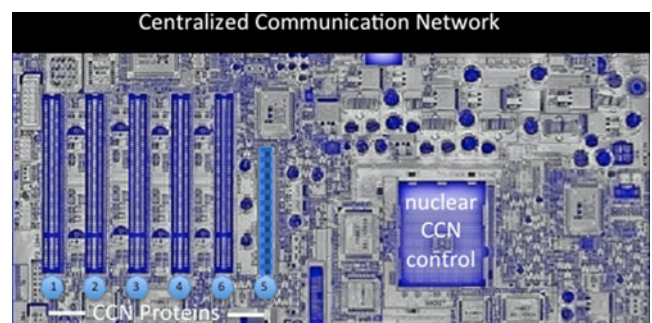


Fig. 8 An artistic representation of the Centralized Communication Network (CCN)

Acknowledgments I am grateful to all the colleagues with whom I could engage very fruitful collaborative projects. The International Workshops on the CCN family of Genes have been the source the inspirational discussions. Again this review provides me the opportunity to deeply thank my wife Annick for her tremendous help in organizing the workshops and my friend and colleague Herman Yeger for his early involvement and support in this venture. The work that was performed in my laboratory was funded by grants from the European Union : PROTHETS (Prognosis and Therapeutic Targets of Ewing Family of Tumors, FP6 Contract 503036), grants from the Ligue Nationale Contre le Cancer, and the French Ministry of Education.

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