

In Vitro Antibacterial Activity of E-0702, a New Semisynthetic Cephalosporin

KANEMASA KATSU,^{1*} KYOSUKE KITOH,² MATSUHISA INOUE,¹ AND SUSUMU MITSUHASHI¹

Department of Microbiology, School of Medicine, Gunma University, Maebashi, Gunma,¹ and Research Laboratories, Eisai Co., Ltd., Bunkyo-ku, Tokyo,² Japan

Received 16 February 1982/Accepted 18 May 1982

E-0702 is an antipseudomonal cephalosporin derivative which has a broad spectrum of antibacterial activity against clinical isolates of gram-positive and gram-negative bacteria. Its in vitro antibacterial activities were less than those of cefoperazone and cefotaxime against *Staphylococcus aureus* and *Staphylococcus epidermidis*, but were significantly high against gram-negative bacteria including the *Pseudomonas* group. It was most characteristic that E-0702 showed high antibacterial activity against *Pseudomonas aeruginosa*. E-0702 was relatively stable to inactivation by plasmid-mediated penicillinases and cephalosporinases produced by gram-negative bacteria.

Many new cephalosporins with a broad antibacterial spectrum have been developed in the past few years. Several of these antibiotics have been in clinical use for the treatment of a wide range of bacterial infections. E-0702, (6R, 7R)-3-[(1 - carboxymethyl - 1H - tetrazol - 5 - yl) - thiomethyl] - 7 - [(2R) - 2 - (6,7 - dihydroxy - 4 - oxo - 4H - 1 - benzopyran - 3 - carboxamido) - 2 - (4 - hydroxyphenyl) - acetamido] - 8 - oxo - 5 - thia - 1 - azabicyclo[4.2.0]oct - 2 - ene - 2 - carboxylic acid disodium salt (Fig. 1), is a new antipseudomonal cephalosporin derivative with a broad antibacterial spectrum, developed by Eisai Co., Ltd., Tokyo, Japan. The current report summarizes the results of studies of the in vitro antibacterial activity and β -lactamase stability of this new cephalosporin with clinical isolates stocked in this laboratory.

MATERIALS AND METHODS

Antibiotics. E-0702 was synthesized in the research laboratories of Eisai Co. As comparative compounds, cefoperazone (CFP) and cefotaxime (CTX) were supplied by Toyama Chemical Co., Japan, and Hoechst Japan Ltd., Japan, respectively. Cephaloridine, cefazolin, and penicillin G (PCG) were obtained commercially.

Organisms. The strains of gram-positive and gram-negative bacteria used in this study were isolated from clinical materials in Japan between 1979 and 1981 and have been maintained since isolation in the Laboratory of Drug Resistance in Bacteria, Gunma University, Japan.

Media. Sensitivity test (ST) agar, ST broth (Nissui Seiyaku Co., Japan), and nutrient agar containing bromothymol blue and lactose were used. For the dilution of cell suspensions, buffered saline containing 0.01% gelatin (1) was used.

Determination of MICs. Minimal inhibitory concentration (MIC) was determined by an agar dilution

method. Twofold serial antibiotic dilutions were made in ST agar, ST agar containing 5% defibrinated horse blood for *Streptococcus pyogenes*, or chocolate ST agar for *Neisseria gonorrhoeae*. The test organism was grown at 37°C overnight in ST broth used for preculture. ST broth containing 0.3% KNO₃ was used for *Pseudomonas* strains. The strains of *N. gonorrhoeae* were grown on chocolate ST agar, and colonies were suspended in buffered saline-0.01% gelatin just before dilution. Overnight cultures of test strains were diluted in buffered saline-0.01% gelatin to a final concentration of about 10⁶ colony-forming units (CFU) per ml, and one loopful (about 0.005 ml) of each bacterial suspension was inoculated on agar plates with an inoculator (Microplanter; Sakuma Seisakusho, Ltd., Tokyo). Plates were incubated for 18 h at 37°C. *N. gonorrhoeae* was incubated in CO₂ for 48 h at 37°C. The MIC was defined as the lowest concentration of antibiotic that allowed no visible growth.

Bactericidal activity. An overnight culture in ST broth at 37°C was diluted in the same fresh broth to about 10³ CFU/ml and incubated with shaking at 37°C. After 2 h of incubation, antibiotics were added to the cultures. Samples were taken at appropriate time intervals, immediately diluted in broth, and plated at several dilutions on bromothymol blue agar. The numbers of colonies were counted after 18 h of incubation at 37°C.

Effect of inoculum size on MICs and MBCs. Each test organism was grown overnight in ST broth at 37°C. Tenfold-diluted cultures were inoculated in ST broth containing twofold serial dilutions of a drug. MICs were read after incubation at 37°C for 18 h. One loopful (about 0.01 ml) of each culture tube in the MIC test series was spotted onto drug-free bromothymol blue agar plates. After incubation at 37°C for 18 h, the minimal bactericidal concentrations (MBCs) of antibiotic were estimated as the lowest concentrations that did not allow the formation of colonies.

Susceptibility to β -lactamase. Chromosomally mediated and plasmid-mediated β -lactamases used in this study were purified by methods described previously

TABLE 1. In vitro antibacterial activities of E-0702 and other cephalosporins

Organism (no. of strains)	Anti-biotic	Range of MIC ($\mu\text{g/ml}$)	MIC ₅₀ ($\mu\text{g/ml}$) ^a	MIC ₉₀ ($\mu\text{g/ml}$) ^a
<i>Staphylococcus aureus</i> (100)	E-0702	3.13–12.5	4.15	5.80
	CFP	0.4–3.13	0.97	1.50
	CTX	0.4–3.13	0.61	1.25
<i>S. epidermidis</i> (50)	E-0702	1.56–50	7.00	28.0
	CFP	0.4–50	1.47	9.50
	CTX	0.8–>100	1.38	12.5
<i>Neisseria gonorrhoeae</i> (53)	E-0702	0.025–1.56	0.09	0.50
	CFP	0.006–0.2	0.02	0.07
	CTX	≤ 0.003 –0.1	0.006	0.04
<i>Escherichia coli</i> (200)	E-0702	≤ 0.003 –3.13	0.01	0.08
	CFP	0.025–100	0.16	2.10
	CTX	0.0125–0.4	0.03	0.04
<i>Klebsiella pneumoniae</i> (100)	E-0702	0.006–3.13	0.02	0.19
	CFP	0.1–25	0.34	2.70
	CTX	0.0125–0.4	0.03	0.07
<i>Enterobacter cloacae</i> (100)	E-0702	0.025–100	0.27	2.20
	CFP	0.05–>100	0.37	17.5
	CTX	0.025–>100	0.11	8.70
<i>Serratia marcescens</i> (100)	E-0702	0.0125–>100	0.06	56.0
	CFP	0.05–>100	0.88	38.2
	CTX	0.05–12.5	0.22	2.30
<i>Proteus mirabilis</i> (50)	E-0702	0.025–6.25	0.05	0.57
	CFP	0.4–3.13	0.82	1.56
	CTX	0.0125–0.2	0.02	0.04
<i>P. vulgaris</i> (50)	E-0702	0.025–50	0.08	0.80
	CFP	0.2–25	1.00	2.35
	CTX	0.0125–3.13	0.02	0.04
<i>P. morganii</i> (50)	E-0702	0.05–12.5	0.14	1.18
	CFP	0.2–25	0.92	5.00
	CTX	0.0125–6.25	0.02	0.20
<i>P. rettgeri</i> (25)	E-0702	0.025–12.5	0.06	0.24
	CFP	0.1–25	0.64	7.40
	CTX	0.025–0.2	0.02	0.09
<i>P. inconstans</i> (21)	E-0702	0.0125–0.8	0.02	0.39
	CFP	0.2–25	1.38	6.25
	CTX	0.0125–0.8	0.05	0.20
<i>Citrobacter freundii</i> (50)	E-0702	0.003–25	0.006	0.40
	CFP	0.05–>100	0.10	17.0
	CTX	0.05–100	0.18	25.0
<i>Salmonella</i> spp. (50)	E-0702	≤ 0.0008 –0.025	0.001	0.005
	CFP	0.2–3.13	0.43	1.00
	CTX	0.0125–0.4	0.06	0.16
<i>Acinetobacter calcoaceticus</i> (100)	E-0702	0.1–3.13	0.29	0.98
	CFP	3.13–>100	26.7	75.0
	CTX	0.4–50	7.45	20.0
<i>Pseudomonas aeruginosa</i> (200)	E-0702	0.025–6.25	0.09	0.45
	CFP	0.8–100	3.30	8.80
	CTX	0.4–>100	10.0	24.5

TABLE 1—Continued

Organism (no. of strains)	Anti-biotic	Range of MIC ($\mu\text{g/ml}$)	MIC ₅₀ ($\mu\text{g/ml}$) ^a	MIC ₉₀ ($\mu\text{g/ml}$) ^a
<i>P. cepacia</i> (50)	E-0702	0.0125–>100	1.26	37.5
	CFP	3.13–>100	21.0	94.0
	CTX	1.56–>100	10.8	42.5
<i>P. maltophilia</i> (50)	E-0702	0.0125–100	0.39	19.8
	CFP	1.56–>100	8.50	44.5
	CTX	3.13–>100	31.5	>100

^a MIC₅₀ and MIC₉₀, Concentrations required to inhibit the growth of 50 and 90%, respectively, of the organisms used.

(3–8, 10, 13–17). β -Lactamase activity was determined by the spectrophotometric method, as described previously (11, 12). The reaction mixture consisted of 3 ml of a 100 μM substrate solution in 50 mM phosphate buffer (pH 7.0) and 50 μl of enzyme solution. Substrate specificity was expressed as the relative rate of hydrolysis of the five antibiotics, taking the absolute rate of PCG or cephaloridine hydrolysis as 100.

RESULTS

Antibacterial activity. The in vitro activity of E-0702 against gram-positive and gram-negative bacteria was compared with those of CFP and CTX (Table 1). Against both *Staphylococcus aureus* and *Staphylococcus epidermidis*, E-0702 was less active than other antibiotics. On the other hand, E-0702 was more active than CFP against gram-negative bacteria except *N. gonorrhoeae*. However, 90% of *N. gonorrhoeae* strains were inhibited by 0.5 μg of E-0702 per ml. The concentrations of E-0702 required to inhibit the growth of 90% of organisms of *Escherichia coli*, *Klebsiella pneumoniae*, and *Enterobacter cloacae* were 0.08, 0.19, and 2.20 $\mu\text{g/ml}$, respectively, indicating that E-0702 was 10- to 25-fold more potent than CFP and similar to CTX. E-0702 was also highly active against *Serratia marcescens*, *Proteus mirabilis*, and indole-positive *Proteus* species. The activity of E-0702 against *S. marcescens* and *Proteus inconstans* was greater than that of CFP and similar to that of CTX. Against *P. mirabilis*, *Proteus morganii*, *Proteus vulgaris*, and *Proteus rettgeri*, E-0702 was some 10-fold more active than CFP but slightly less active than CTX. E-0702 was the most active compound tested against *Citrobacter freundii* and *Salmonella* species.

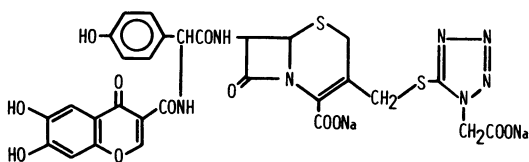


FIG. 1. Chemical structure of E-0702.

The activity of E-0702 against *Pseudomonas aeruginosa* and *Acinetobacter calcoaceticus* was remarkable: 90% of the strains of these species were inhibited by E-0702 at concentrations of 0.45 and 0.98 $\mu\text{g/ml}$, respectively. It was found that E-0702 was also the most effective compound against the other *Pseudomonas* species.

Bactericidal activity. The bactericidal activities of E-0702 were compared with those of CFP against representative strains of *S. marcescens* and *P. aeruginosa* (Fig. 2). Against *S. marcescens* IAM1184, E-0702 and CFP were bactericidal at concentrations of 0.8 and 1.56 $\mu\text{g/ml}$, respectively. Against *P. aeruginosa* PAO1, however, E-0702 displayed a much higher bacte-

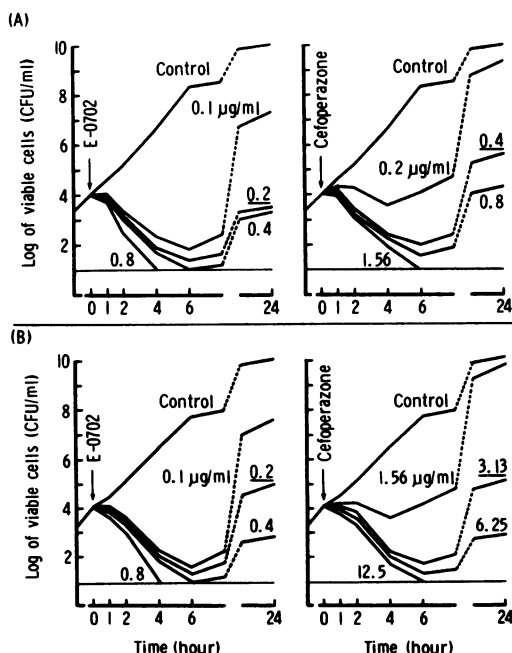


FIG. 2. Bactericidal activity of E-0702 and CFP. (A) *S. marcescens* IAM1184. (B) *P. aeruginosa* PAO1. Underlined numerals indicate the MIC (in micrograms per milliliter) of each drug.

TABLE 2. Bacteriostatic and bactericidal activities of E-0702 at various inoculum sizes

Organism	Inoculum size (CFU)	E-0702		CFP		CTX	
		MIC ($\mu\text{g/ml}$)	MBC ($\mu\text{g/ml}$)	MIC ($\mu\text{g/ml}$)	MBC ($\mu\text{g/ml}$)	MIC ($\mu\text{g/ml}$)	MBC ($\mu\text{g/ml}$)
<i>E. coli</i> NIHJ JC-2	10^8	6.25	6.25	6.25	6.25	6.25	6.25
	10^7	3.13	3.13	3.13	3.13	1.56	3.13
	10^6	0.025	0.05	0.4	0.8	0.1	0.1
	10^5	0.013	0.013	0.2	0.2	0.1	0.1
	10^4	≤ 0.003	0.006	0.1	0.1	0.1	0.1
	10^3	≤ 0.003	≤ 0.003	0.1	0.1	0.1	0.1
	10^2	≤ 0.003	≤ 0.003	0.1	0.1	0.1	0.1
<i>K. pneumoniae</i> PCI-602	10^8	6.25	6.25	50	50	0.4	0.4
	10^7	1.56	1.56	12.5	12.5	0.2	0.2
	10^6	0.2	0.4	0.2	0.2	0.013	0.025
	10^5	0.013	0.025	0.1	0.2	0.006	0.006
	10^4	≤ 0.003	≤ 0.003	0.05	0.05	0.006	0.006
	10^3	≤ 0.003	≤ 0.003	0.025	0.025	≤ 0.003	≤ 0.003
	10^2	≤ 0.003	≤ 0.003	0.013	0.013	≤ 0.003	≤ 0.003
<i>S. marcescens</i> IAM1184	10^8	200	800	400	>800	100	400
	10^7	50	200	100	400	50	200
	10^6	6.25	12.5	6.25	25	0.8	3.13
	10^5	0.4	0.8	1.56	3.13	0.4	0.8
	10^4	0.1	0.4	0.8	3.13	0.2	0.4
	10^3	0.1	0.2	0.8	1.56	0.2	0.4
	10^2	0.05	0.05	0.4	0.4	0.1	0.1
<i>P. aeruginosa</i> PAO1	10^8	>800	>800	>800	>800	>800	>800
	10^7	800	>800	>800	>800	800	>800
	10^6	50	800	100	800	100	800
	10^5	0.1	0.8	6.25	50	12.5	50
	10^4	0.05	0.4	3.13	6.25	12.5	25
	10^3	0.025	0.2	3.13	6.25	12.5	12.5
	10^2	0.006	0.05	3.13	3.13	6.25	12.5

ricidal activity than did CFP.

Effect of inoculum size on MICs and MBCs. The effect of inoculum size on the MICs and MBCs of E-0702 was studied with four species, with the inocula varying between 10^2 and 10^8 CFU (Table 2). A large difference in the values occurred between 10^7 and 10^6 CFU in *E. coli* NIHJ JC-2 and *K. pneumoniae* PCI-602 and between 10^7 and 10^5 CFU in *S. marcescens* IAM1184 and *P. aeruginosa* PAO1, and the values became lower as the inoculum decreased. The MBCs of all the antibiotics were almost identical to the MICs against *E. coli* and *K. pneumoniae*, but 2- to 16-fold higher against *S. marcescens* and *P. aeruginosa*.

Susceptibility to β -lactamase. The maximum rates of hydrolysis (V_{\max}) of the five cephalosporins and PCG are presented in Table 3. E-0702 was stable to various types of cephalosporinases produced by *E. coli* (7), *E. cloacae* (8), *C. freundii* (15), *P. aeruginosa* (10), and *P. rettgeri* (6). The compound also showed a high stability to penicillinase (PCase) types II (2) and III (oxacillin-hydrolyzing PCase; 17) and type IV (carbenicillin-hydrolyzing PCase; 14). However,

E-0702 was partially hydrolyzed by PCase type I (TEM-type PCase; 3), *P. morganii* cephalosporinase (16), and the cefuroxime-hydrolyzing β -lactamases (9) produced by *P. vulgaris* (5), *Pseudomonas cepacia* (4), and *Bacteroides fragilis* (13). The stability of E-0702 to β -lactamase was almost the same as that of CFP.

DISCUSSION

E-0702 is a new parenteral cephalosporin with a broad spectrum of antibacterial activity. This study shows that, with the exception of *N. gonorrhoeae*, E-0702 is more active than CFP against gram-negative bacteria isolated from clinical specimens. A notable characteristic of E-0702 is that this compound has a marked advantage over the other cephalosporins with respect to activity against *P. aeruginosa*, being 20- to 110-fold more active than CFP and CTX. Furthermore, E-0702 also has remarkable activity in vitro against *Salmonella* species, *C. freundii*, and *A. calcoaceticus*. E-0702 is less active in vitro against staphylococci than against gram-negative bacteria.

TABLE 3. Stability of E-0702 to β -lactamases produced by gram-negative bacteria

Organism	Type of β -lactamase	Relative rate of hydrolysis ^a					
		CER	E-0702	CFP	CTX	CEZ	PCG
<i>E. coli</i> W3630(Rms212)	PCase type I	120	54	42	<1	13	100
<i>E. coli</i> W3630(Rms213)	PCase type II	30	<1	1	3	3	100
<i>E. coli</i> W3630(Rte16)	PCase type III	68	3	7	<1	<1	100
<i>P. aeruginosa</i> MI4259(Rms139)	PCase type IV	15	<1	<1	<1	3	100
<i>E. coli</i> GN5482	CSase	100	<1	4	<1	311	63
<i>E. cloacae</i> GN7471	CSase	100	<1	1	<1	100	12
<i>C. freundii</i> GN346	CSase	100	<1	2	<1	116	3
<i>P. aeruginosa</i> GN10362	CSase	100	<1	5	<1	222	29
<i>P. rettgeri</i> GN4430	CSase	100	<1	<1	<1	99	8
<i>P. morgani</i> GN5407	CSase	100	20	1	<1	20	16
<i>S. marcescens</i> GN10857	CSase	100	11	3	<1	198	3
<i>P. cepacia</i> GN11164	CXase	100	153	10	174	156	161
<i>P. vulgaris</i> GN7919	CXase	100	50	15	29	387	20
<i>B. fragilis</i> GN11478	CXase	100	20	8	7	60	50

^a Hydrolysis of each substrate by PCase, cephalosporinase (CSase), and cefuroxime-hydrolyzing β -lactamase (CXase) is expressed as a relative rate of hydrolysis, taking the absolute rate of PCG or cephaloridine (CER) hydrolysis as 100. CEZ, Cefazolin.

Like CFP, E-0702 is relatively stable to various types of β -lactamase and has an inoculum effect in vitro against *E. coli*, *K. pneumoniae*, *S. marcescens*, and *P. aeruginosa*.

Thus, E-0702 has exceptional antibacterial properties which may have clinical significance if pharmacokinetic and toxicological studies demonstrate adequate levels and safety.

LITERATURE CITED

- Curtiss, R., III. 1965. Chromosomal aberrations associated with mutations to bacteriophage resistance in *Escherichia coli*. *J. Bacteriol.* 89:28-40.
- Dale, J. W., and J. T. Smith. 1974. R-factor-mediated β -lactamases that hydrolyze oxacillin: evidence for two distinct groups. *J. Bacteriol.* 119:351-356.
- Egawa, R., T. Sawai, and S. Mitsuhashi. 1967. Drug resistance of enteric bacteria. XII. Unique substrate specificity of penicillinase produced by R-factor. *Jpn. J. Microbiol.* 11:173-178.
- Hirai, K., S. Iyobe, M. Inoue, and S. Mitsuhashi. 1980. Purification and properties of a new β -lactamase from *Pseudomonas cepacia*. *Antimicrob. Agents Chemother.* 17:355-358.
- Matsubara, N., A. Yotsuji, K. Kumano, M. Inoue, and S. Mitsuhashi. 1981. Purification and some properties of a cephalosporinase from *Proteus vulgaris*. *Antimicrob. Agents Chemother.* 19:185-187.
- Matsuura, M., H. Nakazawa, M. Inoue, and S. Mitsuhashi. 1980. Purification and biochemical properties of β -lactamase produced by *Proteus rettgeri*. *Antimicrob. Agents Chemother.* 18:687-690.
- Minami, S., M. Inoue, and S. Mitsuhashi. 1980. Purification and properties of cephalosporinase in *Escherichia coli*. *Antimicrob. Agents Chemother.* 18:77-80.
- Minami, S., M. Inoue, and S. Mitsuhashi. 1980. Purification and properties of a cephalosporinase from *Enterobacter cloacae*. *Antimicrob. Agents Chemother.* 18:853-857.
- Mitsuhashi, S., and M. Inoue. 1981. Mechanisms of resistance to β -lactam antibiotics, p. 41-56. In S. Mitsuhashi (ed.), *Beta-lactam antibiotics*. Japan Scientific Societies Press and Springer-Verlag, Tokyo.
- Murata, T., S. Minami, K. Yasuda, S. Iyobe, M. Inoue, and S. Mitsuhashi. 1981. Purification and properties of cephalosporinase from *Pseudomonas aeruginosa*. *J. Antibiot.* 34:1164-1170.
- Ross, G. W., K. V. Chanter, A. M. Harris, S. M. Kirby, M. J. Marshall, and C. H. O'Callaghan. 1973. Comparison of assay techniques for β -lactamase activity. *Anal. Chem.* 54:9-16.
- Samuni, A. 1975. A direct spectrophotometric assay and determination of Michaelis constants for the β -lactamase reaction. *Anal. Biochem.* 63:17-26.
- Sato, K., M. Inoue, and S. Mitsuhashi. 1980. Activity of β -lactamase produced by *Bacteroides fragilis* against newly introduced cephalosporins. *Antimicrob. Agents Chemother.* 17:736-737.
- Sawada, Y., S. Yaginuma, M. Tai, S. Iyobe, and S. Mitsuhashi. 1976. Plasmid-mediated penicillin beta-lactamases in *Pseudomonas aeruginosa*. *Antimicrob. Agents Chemother.* 9:55-60.
- Tajima, M., Y. Takenouchi, S. Sugawara, M. Inoue, and S. Mitsuhashi. 1980. Purification and properties of β -lactamase from *Citrobacter freundii* GN7391. *J. Gen. Microbiol.* 121:449-456.
- Toda, M., M. Inoue, and S. Mitsuhashi. 1981. Properties of cephalosporinase from *Proteus morgani*. *J. Antibiot.* 34:1469-1475.
- Yaginuma, S., N. Terakado, and S. Mitsuhashi. 1975. Biochemical properties of a penicillin beta-lactamase mediated by R factor from *Bordetella bronchiseptica*. *Antimicrob. Agents Chemother.* 8:238-242.