

THE EFFECT OF PROLONGED TREATMENT WITH SULPHINPYRAZONE ON THROMBOXANE A₂ AND PROSTACYCLIN IN MAN

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We studied the effect of 3 weeks' treatment with 4×200 mg of sulphinpyrazone daily (six healthy volunteers) on proaggregatory thromboxane A₂ (TxA₂) and antiaggregatory prostacyclin (PGI₂). Platelet TxA₂ production was evaluated by measuring its stable metabolite, immunoreactive thromboxane B₂, from serum, and vessel wall PGI₂ production by measuring its stable metabolite, immunoreactive 6-keto-prostaglandin F_{1α} in plasma. The TxA₂ production (initially 209.0 ± 27.1 ng/ml, mean \pm s.e. mean) decreased to about 30% from the second day of the treatment onwards, and it recovered in three days after the discontinuation of the treatment. PGI₂ (initially 33.6 ± 3.6 pg/ml) did not change. The shift of the balance between TxA₂ and PGI₂ to the dominance of antiaggregatory PGI₂ during sulphinpyrazone treatment may be involved with the efficacy of the drug in the secondary prevention of myocardial infarction.

Introduction

Sulphinpyrazone, which originally was developed for the treatment of gout, has proved to be an effective drug also in the secondary prevention of myocardial infarction, although the mechanism of this action is not known (The Anturane Reinfarction Trial Research Group, 1978, 1980; The Anturan Reinfarction Italian Study Group, 1982). Because platelet activation takes place during cardiac ischaemia (Haerem, 1971; Kumpuris *et al.*, 1980; Green *et al.*, 1980), the known effects of sulphinpyrazone on platelets (Smythe *et al.*, 1965; Mustard *et al.*, 1967) may, however, be involved. We studied the effects of sulphinpyrazone on proaggregatory thromboxane A₂ (TxA₂) (Hamberg *et al.*, 1975) and antiaggregatory prostacyclin (PGI₂) (Moncada *et al.*, 1976; Johnson *et al.*, 1976), the balance of which potentially regulates platelet function (Moncada & Vane, 1979).

Methods

We performed this study with six healthy non-smoking volunteers (three females, three males) aged 21–23 years. They were asked to refrain from all medications, other than the study treatments, 2 weeks before and during the trial. On the first 2 days of the experiment the volunteers ingested four

placebo tablets, on days 3–22 four times 200 mg of sulphinpyrazone and on days 23–26 again placebo. Blood samples were drawn at 08.00 h on days 1 and 2 (basal samples), 3 (the first day of sulphinpyrazone, 1 h after ingesting the first 200 mg dose), 4, 5, 8, 12, 17, 22 (the last day of sulphinpyrazone), 23, 24, 25 and 26.

Platelet TxA₂ production was evaluated by measuring its stable metabolite, immunoreactive thromboxane B₂ (i TxB₂) (Hamberg *et al.*, 1975) in serum from blood having clotted at 37°C for 60 min (Viinikka & Ylikorkala, 1980). The amount of i TxB₂ formed in these circumstances correlates well with i TxB₂ generated during induced aggregation in platelet rich plasma (Viinikka & Ylikorkala, 1980). PGI₂ production was determined by assaying its stable metabolite, immunoreactive 6-keto-prostaglandin F_{1α} (i 6-keto-PGF_{1α}) (Johnson *et al.*, 1976) in plasma obtained from blood, which was collected into ice-cold heparinized tubes containing indomethacin (10 μmol/l, final concentration) (Ylikorkala *et al.*, 1981).

Student's *t*-test for paired observations was used for the statistical analysis of the results.

Results

The generation of i TxB₂ by platelets during spontaneous clotting was decreased to about 60% of the initial value in the sample taken 1 h after the first dose

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of sulphinpyrazone. From the second day of the treatment onwards, i TxB₂ production remained on about 30% level of the initial and it recovered in 3 days after the discontinuation of sulphinpyrazone treatment. Plasma i 6-keto-PGF_{1 α} did not change (Figure 1).

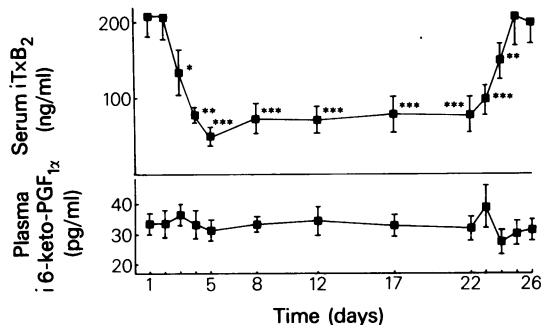


Figure 1 Serum i TxB₂ (mean \pm s.e. mean) and plasma i 6-keto-PGF_{1 α} before, during and after treatment with sulphinpyrazone. Six volunteers ingested 4 \times 200 mg of the drug on days 3–22 and four placebo tablets daily on days 1–2 and 23–26. * P < 0.05; ** P < 0.01; *** P < 0.001 in comparison with the initial level.

Discussion

Increased amounts of TxA₂ are released from activated platelets during cardiac ischaemia (Lewy *et al.*, 1980; Hirsh *et al.*, 1981; Neri Serneri *et al.*, 1981). The increased TxA₂ may further reduce the oxygen delivery of hypoxic myocardium by coronary vasoconstriction and mechanical obstruction of coronary vessels by platelet aggregates. The correlation between the concentration of i TxB₂ in coronary sinus and ventricular ectopic activity (Coker *et al.*, 1981) and the beneficial effects of the inhibition of TxA₂ synthesis in experimental infarction (Smith *et al.*, 1980; Coker & Parratt, 1981; Moschos *et al.*, 1981) further suggest the significance of TxA₂ in myocardial ischaemia. Thus, it is justified to presume that treatments, which decrease TxA₂ synthesis, might also have beneficial effects in myocardial ischaemia.

The data about the effect of sulphinpyrazone on the production of TxA₂ are scanty. Lorenz *et al.* (1981) showed that the generation of i TxB₂ was decreased after four day treatment with sulphinpyrazone. We present here the first data on the effects of long-term sulphinpyrazone treatment and

they show that sulphinpyrazone, as applied in the secondary prevention of myocardial infarction (The Anturane Reinfarction Trial Research Group, 1978, 1980), inhibited more than half of the capacity of platelets to produce TxA₂ in response to thrombin induced aggregation during spontaneous clotting. The inhibition of TxA₂ generation was seen as late as 36 h after the ingestion of the last dose. This supports the hypothesis that sulphinpyrazone is metabolized into a long-acting active metabolite (McDonald *et al.*, 1980).

The data about the effect of sulphinpyrazone on PGI₂ production are scanty, too. Gordon & Pearson (1978) showed that 1 mmol/l of sulphinpyrazone had no effect on porcine endothelial cell PGI₂ production, and Livio *et al.* (1980) found no inhibition of PGI₂-like activity after the administration of 200 mg/kg of sulphinpyrazone into rats. Our data suggests that like in other species, the PGI₂ production also in human is resistant to the action of sulphinpyrazone.

The inhibition of platelet TxA₂ production could theoretically be the result of inhibition of cyclo-oxygenase or of thromboxane synthetase (Moncada & Vane, 1979). The findings that sulphinpyrazone was able to inhibit also the synthesis of other prostaglandins, and that it antagonized the irreversible inhibition of platelet cyclo-oxygenase by acetylsalicylic acid (Ali *et al.*, 1977; McDonald *et al.*, 1980), however, show that cyclo-oxygenase is the target of the action of sulphinpyrazone in the arachidonate cascade. The decreased TxA₂ production together with unchanged PGI₂ generation during sulphinpyrazone treatment seen in this study suggest that human platelet cyclo-oxygenase is more sensitive to the action of the drug than is endothelial cell cyclo-oxygenase. The mechanism of this difference remains, however, to be studied.

In conclusion, our data suggest that during prolonged treatment with sulphinpyrazone the balance between TxA₂ and PGI₂ is shifted to the direction of antiaggregatory, vasodilatory PGI₂. Because this change is likely to be favourable in coronary artery disease, we presume that the reduction of cardiac mortality in the secondary prevention of myocardial infarction by sulphinpyrazone might have been mediated through inhibition of TxA₂ synthesis.

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